Sensing of magnetic field effects in radical-pair reactions using a quantum sensor

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Magnetic field effects (MFE) in certain chemical reactions have been well established in the last five decades and are attributed to the evolution of transient radical pairs whose spin dynamics are determined by local and external magnetic fields. The majority of existing experimental techniques used to probe these reactions only provide ensemble averaged reaction parameters and spin chemistry, hindering the observation of the potential presence of quantum coherent phenomena at the single molecule scale. Here, considering a single nitrogenvacancy (NV) center as quantum sensor, we investigate the prospects and requirements for the detection of MFEs on the spin dynamics of radical pairs at the scale of a single and small ensembles of molecules. We employ elaborate and realistic models of radical pairs, considering their coupling to the local spin environment and the sensor. For two model systems, we derive signals of MFE detectable even in the weak coupling regime between a radical pair and an NV quantum sensor and observe that the dynamics of certain populations, as well as coherence elements, of the density matrix of the radical pair are directly detectable. Our investigations will provide important guidelines for the potential detection of spin chemistry of biomolecules at the scale of a single and small ensembles of molecules.

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I. INTRODUCTION

Investigating the fundamental role of spin interactions in magnetic field effects (MFE) in chemical reactions has a long history [1–3]. The study of spin-chemical effects provides important insights about structure, kinetics and magnetic properties of transient intermediate chemical species [4,5] and are explored in various interdisciplinary applications including sensitivity enhancement in nuclear magnetic resonance (NMR) [6], quantum computing [7], avian magnetic compass [8], and solar energy conversion kinetics in photosynthetic systems [9].

It is astounding that spin interactions can have decisive effects on the fate of chemical reactions as the energy of spin transitions typically is orders of magnitude smaller than the thermal energy [4]. However, spin dependent MFEs in chemical systems, also known as the radical-pair mechanism (RPM) [2,8,10], rely on the creation of transient paramagnetic species in a nonequilibrium state called radicals, which are chemical species with an odd number of electrons. In the RPM, the radical pair (RP) is spatially separated but in a spin-correlated state and the recombination of the radicals

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back to the molecular precursor state is spin selective. The influence of an external magnetic field then occurs in terms of a modulation of the spin dynamics and consequently an alteration in the yield of products formed from the various spin states [11,12].

The formulation of the RPM started in the late 1960s to explain nonequilibrium magnetic resonance spectra of chemical reactions of organic molecules [13-19], while the recent interest is fuelled by investigations of the RPM as the most plausible mechanism for MFE in biological reaction kinetics [20,21]. These include flavoproteins related to DNA photolyases, the involvement of cryptochromes in circadian rhythms [22,23] and their proposed role in animal magnetoreception [24]. MFEs have been recorded in cryptochromes [11] and seem to fulfill the structural and dynamical requirements of the RPM [8]. Considerable interest exists also in the role of radical pairs in chemical kinetics in photosynthetic reaction centres [9,25,26]. Flavoenzymes [27]—Flavin-based enzymes—are responsible for catalytic functions in diverse biological reactions [28] and the involvement of various RPs in these reactions is debated in recent discussions [29,30].

Existing experimental techniques for *in vitro* probing of MFE in RP dynamics in chemical reactions rely on averaged signals collected from a large ensemble of molecules. These techniques include time-resolved electron paramagnetic resonance (TREPR) spectroscopy [31,32], and optical methods based on absorption [33–36] and fluorescence detection [37–41]. Studies of RP reactions conducted using these techniques can only provide ensemble averaged information of spin dynamics along with requiring a large quantity (few microliters) of potentially precious biological samples.

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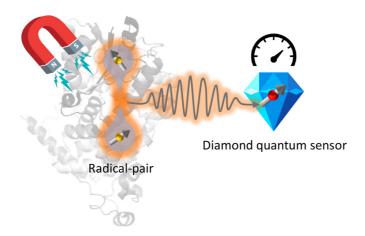


FIG. 1. Simplified illustration for sensing of MFEs of an RP reaction using a NV quantum sensor in diamond. An RP on a host molecule is formed in a spin-correlated state (yellow). Subject to local and external fields, the distinct RP evolution induces a signal that is detectable by a nearby NV center (red) using appropriate sensing sequences.

It is therefore imperative to instead consider single-molecule detection techniques in order to reveal potential quantum coherent spin evolution otherwise hidden by ensemble averaging [42–44]. Specifically the single negatively charged nitrogen-vacancy (NV) center in diamond [45] has attracted significant interest as potential candidate for the detection of spin-chemical effects of RP reactions at the single molecule scale [43,44]. This is achieved due to the excellent biocompatibility of diamond [46,47], the attainable nanometer scale spatial resolution and remarkable sensitivity to external electromagnetic fields of a single NV center [48–52].

In the present work, we discuss a realistic avenue for the detection of MFE associated with RPs at the scale of a single and small ensembles (≤ 100) of molecules using a single NV center in diamond (Fig. 1). We investigate both the weak and the strong coupling regime between the NV center and RPs. Compared to the simplified model employed by Finkler *et al*. [44] neglecting important spin interactions and considering only the strong coupling regime, we here use an elaborate model governing realistic RP spin dynamics. We include up to three nuclear spins per radical with maximum anisotropic hyperfine coupling along with dipolar and exchange coupling between the radicals. Applying standard sensing protocols, we derive measurable signals received by an NV center and quantify magnetic field dependent RP dynamics. We show that in the weak coupling regime, the signal generated by an RP on a single molecule is comfortably within the sensitivity limit of state-of-the-art shallow NV centers and dynamics of certain populations, as well as coherences, of the RP density matrix is directly accessible. Nontrivial features of RP spin dynamics can be observed depending on direction and magnitude of a bias magnetic field. Further, we observe that signal features tend to average out when we consider a small ensemble of RPs, highlighting the importance of singlemolecule detection. In the strong coupling regime, we find that although there is an opportunity to probe various RP spin state populations individually, detection becomes challenging especially in larger biomolecules due to the increased number of unavoidable and spectrally indistinguishable hyperfine interactions within the RP.

The paper is organized as follows. In Sec. II, we introduce the theoretical framework for the RP and interaction with the NV center. In Sec. III, we describe the numerical simulations performed with realistic RP systems and present the main results. We conclude in Sec. IV and discuss possible future directions.

II. DESCRIPTION OF THE MODEL

The general situation considered for the model is illustrated in Fig. 2(a). Molecules hosting RPs are distributed above the diamond surface and an NV center is situated at a depth d from the surface, where d is typically in the range 5–50 nm depending on the preparation method [53,54]. We choose to work in the NV frame (shown in the figure) and the total Hamiltonian of the whole system in this frame is given as

$$H = H_{\rm NV} + H_{\rm RP} + H_{\rm C},\tag{1}$$

where H_{NV} , H_{RP} , and H_{C} are the Hamiltonians governing the dynamics of the NV center, the RP, and coupling between the NV center and the RP, respectively.

We consider the C_{3v} symmetric, negatively charged NV⁻ center (hereafter called NV) that consists of ground (³A, total spin s = 1), metastable (${}^{1}A$, s = 0) and excited (${}^{3}E$, s=1) states [45]. The ground state triplet $\{|0\rangle, |\pm 1\rangle|\}$ is split at zero magnetic field with $D_{\rm ZFS} = 2.87\,\rm GHz$ [48]. The $|+1\rangle$ and $|-1\rangle$ can be further split by application of an external magnetic field, thus making all the three states accessible by application of appropriate control fields. Upon optical excitation typically with 532 nm light, the NV center might relax via a spin-dependent intersystem crossing [55], a process that allows for initialization in $|0\rangle$ and spin-state dependent fluorescence contrast between $|0\rangle$ and the $|\pm 1\rangle$ states. These properties combined with an external magnetic field can be exploited to isolate $|0\rangle \leftrightarrow |+1\rangle$ or $|0\rangle \leftrightarrow |-1\rangle$ as a two-level system with effective spin s = 1/2. In addition, the NV center interacts with nuclear spins, predominantly (abundance of 99.6% [56]) the intrinsic ¹⁴N with total spin s = 1, hyperfine coupling tensor A_N and quadrupolar coupling $Q_{\rm N} \simeq -5.01\,{\rm MHz}$ [56], causing further splitting of the $|\pm 1\rangle$ states. Due to the axial symmetry of the NV center, $A_{\rm N}$ can be expressed in a diagonal form in the principal axis system of the NV axis with diagonal elements $[A_{\rm N}^{\perp} \simeq$ -2.7 MHz, $A_{\rm N}^{\perp}$, $A_{\rm N}^{\parallel} \simeq -2.16$ MHz] [56]. Taking the above details into account, the relevant part of the NV center Hamiltonian can be written as

$$H_{\rm NV} = D_{\rm ZFS} J_z^2 + \gamma_e (\vec{B_0} \cdot \vec{J}) + \vec{J} \cdot A_{\rm N} \cdot I_{\rm N}, \tag{2}$$

where γ_e is the gyromagnetic ratio of an electron, $\vec{J} = [J_x, J_y, J_z]$ and $\vec{I}_N = [I_{Nx}, I_{Ny}, I_{Nz}]$ are, respectively, spin operators of the NV center and ^{14}N nuclear spin, and $\vec{B}_0 = [B_{0x}, B_{0y}, B_{0z}]$ is the vector of the externally applied magnetic field on the NV center. The relatively large zero-field splitting allows to make the secular approximation $(|D_{ZFS} + \gamma_e B_{0z}| \gg A_N^{\perp})$ where we can ignore all terms involving electron-nuclear spin flip flops (containing A_N^{\perp}). Further, we assume $B_{0z} \gg B_{0x}$ and B_{0y} and consequently choose $|0\rangle \leftrightarrow |+1\rangle$ (hereafter

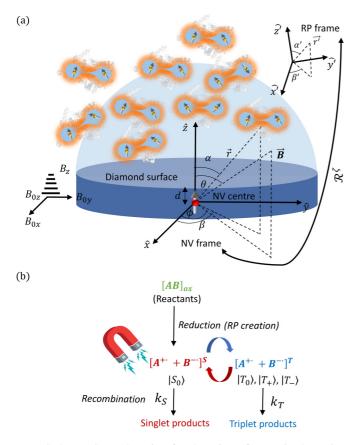


FIG. 2. (a) General setting for detection of RP spin dynamics using an NV center situated at a depth d under the diamond surface. The light blue hemisphere shows the sensing volume of the NV center and an RP situated in this volume contribute to the detectable signal by the NV center. Without loss of generality, we choose to work in the axis system where normal to the diamond surface is aligned along the NV axis ([1 1 1] crystal axis), named "NV frame" here. The origin of the coordinate system $(\hat{r}', \alpha', \beta')$ associated with the molecule that hosts an RP, named "RP frame," is situated at location($\vec{r}, \zeta = \{\alpha, \beta\}$) with respect to the NV center. The two axis systems are related by the rotation matrix \mathcal{R}^{ζ} , which is different for each RP. The magnetic field is applied in direction (θ, ϕ) . When RP spin dynamics is probed at very low magnetic fields (close to Earth's), a gradient is required for the magnetic field in the z direction to ensure that the two level approximation of the NV center energy levels is valid. (b) Simplified description of the RPM. The two factors, (i) magnetic field dependent interconversion between the singlet state and triplet states, and (ii) spin dependent product yield, make the RPM a plausible mechanism behind MFE in RP reactions.

denoted $|1\rangle)$ as a two-level system. Now the NV center Hamiltonian simplifies to

$$H_{\text{NV}} = (D_{\text{ZFS}} + \gamma_e B_{0z}) J_z + A_N^{\parallel} J_z I_{Nz}.$$
 (3)

The RP consists of two radicals each containing an unpaired electron. A simplified reaction for the creation and recombination of radicals is shown in Fig. 2(b). The radicals can be generated either by electron transfer or chemical bond breaking from the oxidized state of the molecule. The reduction can be facilitated by various mechanisms, for example, by photoexcitation with light of appropriate wavelength (reduction of flavin

based systems [57]) or by chemical means (Haber-Weiss reaction [58]). Depending on the internal molecular dynamics, the unpaired electrons in the radicals are born in either singlet (s=0) $(|S_0\rangle = [|\uparrow\downarrow\rangle - |\downarrow\uparrow\rangle]/\sqrt{2})$ or triplet (s=1) $(|T_0\rangle = [|\uparrow\downarrow\rangle + |\downarrow\uparrow\rangle]/\sqrt{2}, |T_+\rangle = (|\uparrow\uparrow\rangle, |T_-\rangle = (|\downarrow\downarrow\rangle)$ states, where $|\uparrow\rangle$ and $|\downarrow\rangle$ denote the eigenstate of the Pauli z matrix. After creation, the singlet and triplet states interconvert among each other under the RP Hamiltonian, which for the RP situated at location $[r, \zeta = \{\alpha, \beta\}, \text{ see Fig. 2(a)}]$ is given by [59]

$$H_{\text{RP}}^{\zeta} = -\gamma_{e} [\vec{B}^{\zeta}.(\vec{S}_{1} + \vec{S}_{2})] - 2J_{\text{ex}}^{\zeta} \vec{S}_{1} \cdot \vec{S}_{2} + \vec{S}_{1} \cdot \boldsymbol{D}^{\zeta} \cdot \vec{S}_{2} + \sum_{i=1}^{N_{1}} \vec{S}_{1} \cdot \boldsymbol{A}_{1i}^{\zeta} \cdot \vec{I}_{1i} + \sum_{i=1}^{N_{2}} \vec{S}_{2} \cdot \boldsymbol{A}_{2j}^{\zeta} \cdot \vec{I}_{2j},$$
(4)

where \vec{S}_1 and \vec{S}_2 are the spin operators of the unpaired electrons of the first and second radicals, respectively. The \vec{B}^{ζ} denotes the external magnetic field applied on the RP and it is directed as indicated by angles (θ, ϕ) . The superscript ζ is a mere indication of the parameter value for the radical pair that is relevant. The radicals couple to each other via exchange interaction (coupling constant $J_{\rm ex}^{\zeta}$) and dipolar interaction (coupling tensor D^{ζ}). Each unpaired electron is further surrounded by a set of nuclei in the radical and interact with them via hyperfine coupling. The $\vec{I}_{1i}(A_{1i}^{\zeta})$ and $\vec{I}_{2j}(A_{2j}^{\zeta})$ are spin operators (hyperfine coupling tensors) of the ith nuclei coupled to the unpaired electron in the first and jth nuclei coupled to the unpaired electron in the second radical, respectively. The magnitude of hyperfine coupling is typically in the range of 2.8–28 MHz [60] in organic radicals, whereas the strength of the dipolar and exchange interaction is dependent on the separation between the radicals [61]. Usually the dipolar and hyperfine coupling tensors are simulated or measured in a coordinate frame associated with the molecule that hosts the RP, hereafter referred to as the RP frame. The NV and RP frames are related by a rotation matrix \mathcal{R}^{ζ} , i.e., $H_{\text{RP}}^{\zeta} = \mathcal{R}^{\dagger \zeta} H_{\text{RP}}^{\zeta} \mathcal{R}^{\zeta}$, where H_{RP}^{\prime} is the RP Hamiltonian in the RP frame. All the terms in Eq. (4) except the hyperfine interaction terms commute with the singlet state projector. Thus the spin exchange symmetry is broken, resulting in interconversion between the singlet and triplet states, since the singlet and triplet states are antisymmetric and symmetric under spin exchange, respectively.

Due to interaction with the surrounding environment, after the creation, along with the interconversion dynamics, the spin-correlated RP recombine back to the equilibrium state. The rates of recombination depend on whether the RP is in the singlet (rate constant k_S) or the triplet state (rate constant k_T). Products are thus formed with spin-state dependent yield and can be altered by the application of a suitable magnetic field, with the RPM as result [8]. The recombination dynamics can be modeled by treating the RP as an open quantum system with Lindbladian

$$\mathcal{L}_{\text{rec}}[\cdot] = -\left\{ \left(\frac{k_S}{2} |S_0\rangle \langle S_0| + \frac{k_T}{2} \sum_{i=+,0,-} |T_i\rangle \langle T_i| \right) \otimes I^{\otimes N}, \cdot \right\},$$
(5)

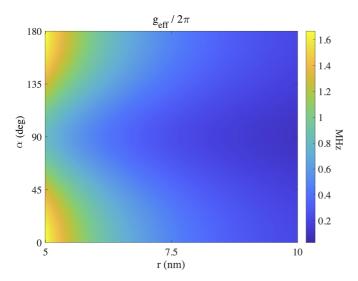


FIG. 3. The effective coupling strength $g_{\rm eff}(r,\zeta)/2\pi = D_r(r)\sqrt{D_{cx}(\zeta)^2 + D_{cz}(\zeta)^2}/\pi$ for various RPs situated at (r,α) $(\beta = 0 \text{ is assumed})$.

where $N = N_1 + N_2$ is total number of nuclei in the RP and $\{.\}$ denotes the anticommutator.

The electron spin of the NV center and the RP is coupled via dipolar interaction and the corresponding Hamiltonian can be written as

$$H_{\rm C}(r,\zeta) = \vec{J}.D_c(r,\zeta).(\vec{S}_1 + \vec{S}_2),$$
 (6)

where D_c is the dipolar coupling tensor in the NV frame which depends on the location of the RP $\zeta = \{\alpha, \beta\}$ and the NV center distance to the RP. Under the secular approximation $|D_{\rm ZFS} + \gamma_e B_z| \gg |D_c|$ and assumption $|B_z| \gg \{|B_x|, |B_y|\}$, the coupling Hamiltonian simplifies to

$$H_{C}(r,\zeta) = J_{z}D_{r}(r) \sum_{i=x,y,z} D_{ci}(\zeta)(S_{1i} + S_{2i})$$

$$= J_{z}D_{r}(r) \sum_{i=x,y,z} \tilde{S}_{i}(\zeta), \tag{7}$$

where $D_r(r) = -\frac{\mu_0 \gamma_e^2 \hbar}{4\pi |r|^3}$, $D_{cx}(\zeta) = 3 \sin 2\alpha \cos \beta/2$, $D_{cy}(\zeta) = 3 \sin 2\alpha \sin \beta/2$, $D_{cz}(\zeta) = 3 \cos^2 \alpha - 1$ and $\tilde{S}_i(\zeta) = D_{ci}(\zeta)(S_{1i} + S_{2i})$. The trace norm $g_{\rm eff}(r,\zeta) = ||D_r(r)\sum_i \tilde{S}_i(\zeta)|| = 2D_r(r)\sqrt{D_{cx}(\zeta)^2 + D_{cy}(\zeta)^2 + D_{cz}(\zeta)^2}$ represents the effective RP-NV coupling (Fig. 3) and can be determined using techniques developed in recent works for three-dimensional molecular localization using the NV center [62–65]. We also assume the same dipolar coupling D_{ci} between the NV and both radicals, which is justified in the regime where the NV-RP distance |r| is much larger than the RP separation $r_{\rm RP}$. Note that the coupling Hamiltonian can not be approximated to contain only the $J_z \tilde{S}_z$ term [44,66] as the strength of cross terms $(J_z \tilde{S}_x)$ and $J_z \tilde{S}_y$ may not be smaller compared to various other parameters in $H_{\rm RP}^{\zeta}$ at low external magnetic fields.

Now, after establishing the general framework, the protocol to study RP dynamics using the NV center is as follows.

(1) Prepare the NV center and the RP in a known initial state $\rho(0) = \rho_{NV}(0) \otimes \rho_{RP}(0)$. The initial state of the

unpaired electrons in the RP is either the $|S_0\rangle$ or $|T_0\rangle$ state as described before, while the nuclei start in the maximally mixed state $I^{\otimes N}/2^N$. The initial state of the NV center can be controlled and depends on the applied sensing scheme.

- (2) Solve the quantum dynamics in the NV frame using the master equation: $\frac{d\rho(t)}{dt} = -i[H, \rho(t)] \mathcal{L}_{rec}[\rho(t)]$. Since the NV center has the capability of detecting magnetic signal directly originating from interconversion dynamics of the RP instead of relying on the product yield based detection, we can simplify the Lindbladian by assuming $k_S = k_T = k$, thus giving $\mathcal{L}_{rec}(\cdot) = -k\{I^{\otimes (N+2)}, \rho(t)\}$ [59]. Further, the rate k may be allowed to include decoherence of the electron spins of the NV center as well because their typical timescale is similar to recombination rates in organic RPs which is in the order of tens of μ s.
- (3) Trace out the RP to calculate the state of the NV center at any time t: $\rho_{NV}(t) = \text{Tr}_{\text{RP}}[\rho(t)]$ and investigate signatures of the RP evolution on the states of the NV center.

Depending on r, α , and β , the coupling between an NV center and an RP varies (Fig. 3). The resulting NV-RP dynamics can be divided in two dynamical regimes:

weak coupling :
$$g_{\text{eff}}(r, \zeta) < \Gamma$$
, (8)

strong coupling :
$$g_{\text{eff}}(r, \zeta) > \Gamma$$
, (9)

where $\Gamma = \frac{1}{\pi T_2}$ and T_2 is the dephasing time of the NV center. T_2 depends on factors including applied sensing sequence and magnetic field [67], proximity to surface impurities, presence of paramagnetic defects and nuclear spins around the NV center [45]. Typical values of T_2 range from a few microseconds to a few milliseconds depending mainly on the applied sensing sequence, the isotopic purity of the diamond sample used [68] and the surface termination [69], making both weak and strong coupling regimes achievable in practice.

III. SIMULATIONS

In this section, we present details and results of the simulations. We consider two RP systems, respectively with isotropic and anisotropic hyperfine coupling tensors. We investigate MFEs in RP systems in the weak as well as strong coupling regimes and discuss appropriate detection strategies. In the weak coupling regime, we study the MFE as a function of strength and direction of magnetic field due to a single as well as a small ensemble of RP. For the strong coupling regime, we point out potential challenges in the regime of single molecule detection that arise for larger biomolecules.

A. RP systems

The two RP systems we investigate are the following.

- (1) Flavin adenine dinucleotide—tryptophan (FAD• TrPH•+) with anisotropic hyperfine couplings, which has been subjected to various MFE studies using absorption spectroscopy in recent years [11]. This system is suggested as one possible candidate enabling avian magnetoreception [8].
- (2) Pyrene-N,N-dimethylaniline (PY• DMA•+) RP, widely studied [70] and with isotropic hyperfine couplings [71].

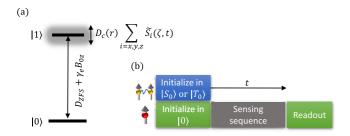


FIG. 4. Energy level structure of an NV center in the weak coupling regime. The RP causes a relative phase between $|0\rangle$ and $|1\rangle$ states of the NV center, this relative phase is proportional to $H_C(r,\zeta,t)$. The sequences to detect the RP dynamics in each regime are shown in (b). The RP is born in $|S_0\rangle$ or $|T_0\rangle$ state and the NV is initialized in $|0\rangle$ state. Then an appropriate sensing sequence is applied which is chosen based on the frequency profile of $X_i(r,\omega,\zeta)$. During sensing, the NV center acquires a phase which contains the information about the RP dynamics which is then read out by a projective measurement.

B. Weak coupling regime

In the weak coupling regime, no observable splitting of the NV energy levels occurs upon interaction with the RP and the effect of spin dynamics of the RP on the NV center dynamics then appears only as a time-dependent classical magnetic field. The coupling Hamiltonian between the NV and a single RP simplifies to

$$H_{C}(r,\zeta,t) = J_{z}D_{r}(r) \sum_{i=x,y,z} \langle \tilde{S}_{i}(\zeta,t) \rangle, \qquad (10)$$

where $\langle O(t) \rangle = \text{Tr}[O\ e^{-iH_{\text{RP}}^{\zeta}t}\ \rho_{\text{RP}}(0)\ e^{iH_{\text{RP}}^{\zeta}t}]$ with $O \in \{\tilde{S}_i\}$, i = x, y, z. The detectable magnetic signal generated from an RP situated at a location (r, ζ) can now be calculated as (in units of Tesla):

$$X_i(r,\zeta,t) = -\frac{D_r(r)}{\gamma_e} \langle \tilde{S}_i(\zeta,t) \rangle = \frac{\mu_0 \gamma_e \hbar}{4\pi |r|^3} \langle \tilde{S}_i(\zeta,t) \rangle$$
 (11)

where i = x, y, z. Due to the $1/|r|^3$ scaling and as indicated in Fig. 2, only an RP within the NV sensing volume will contribute to the field $X_i(r, \zeta, t)$. For estimating the field of a small ensemble as discussed in Sec. III B 2, Eq. (11) can easily be modified by summing over the contributions from individual RPs located within the sensing volume of the NV, a situation similar to nanoscale NMR [54]. The summation for the ensemble is finite since the contribution of RPs outside the sensing volume to $X_i(r, \zeta, t)$ will be negligible.

The maximum signal is generated when $\mathcal{R}^{\zeta} = \mathcal{I}$, where \mathcal{I} is the identity matrix, i.e. when the RP frame is aligned with the NV frame. The possibility of achieving such a condition is discussed in Appendix A 1.

A simple protocol to detect the above signal is shown in Fig. 4. First, we initialize the NV center in $|0\rangle$ and the RP in either the $|S_0\rangle$ or $|T_0\rangle$ state. An appropriate sensing sequence is then applied on the NV. For the range of relevant bias fields 0–20 mT considered here, the $|S_0\rangle$ and $|T_0\rangle$ transition energies are $<< D_{\rm ZFS} - \gamma_e B$, and effects of the NV drive sequence on the RP are thus negligible. The choice of the NV sequence depends on the Fourier spectrum of the detectable signal,

denoted by $X_i(r, \zeta, \omega)$ here. Finally, a projective measurement is made on the NV to yield information about the accumulated phase, which is proportional to $X_i(r, \zeta, t)$, acquired during the sensing sequence. As we are considering the weak coupling regime with effective NV-RP coupling $g_{\rm eff}$ is smaller than rates in $H_{\rm RP}^{\zeta}$ and since both NV and RP are initialized in each iteration of the sequence, it is also assumed that back-action of the NV on the RP can be neglected.

As evident from Eq. (10), only specific combinations of RP density matrix elements corresponding to $\{S_{ji}, j = 1, 2 \text{ and } i = x, y, z\}$ contribute to X(t) and are directly measurable by the NV center. Explicitly, in the singlet-triplet basis, the directly measurable elements are

$$S_{1z} + S_{2z} = |T_{+}\rangle\langle T_{+}| - |T_{-}\rangle\langle T_{-}|,$$
 (12)

which measures the magnetization corresponding to the population difference of outer triplet states and

$$S_{1x} + S_{2x} = \sqrt{2} \operatorname{Re}(|T_{+}\rangle\langle T_{0}| + |T_{-}\rangle\langle T_{0}|),$$

$$S_{1y} + S_{2y} = \sqrt{2} \operatorname{Im}(|T_{+}\rangle\langle T_{0}| + |T_{-}\rangle\langle T_{0}|),$$
 (13)

which measure the magnetization corresponding to the coherences between the outer and central triplet states. Thus the detection of magnetization corresponding to elements $S_{1x} + S_{2x}$ and $S_{1y} + S_{2y}$ can reveal involvement of quantum coherent phenomenon in RP dynamics in the singlet-triplet basis. The full state tomography of the density matrix of RP, however, requires conversion of various elements onto directly detectable ones [Eqs. (12) and (13)] and is beyond the scope of this work.

We consider RPs to be statistically distributed above the surface of the diamond sample. The coupling strength of an RP outside the sensing radius of three-four times the depth d of the NV center [cf. Fig. 2(a)] decreases by an order of magnitude compared to RPs on the surface. The number of RPs falling in the sensing volume depends on the size of the host protein. For the case of proteins with many amino acids and large molecular weights (>500 kDa, the average radius >5 nm [72]), only one of them might fall in the sensing volume with high probability. On the other hand, for the case of smaller proteins (<50 kDa, the average radius being 2–3 nm), tens of RPs can be accommodated in the sensing volume. The following two subsections discuss how the generated signal behaves in these two cases.

1. RP on a single molecule

To study RP dynamics at the single-molecule level, we assume $\mathcal{R}^{\zeta} = I$ and $\beta = 0$ without loss of generality. We use three nuclei with maximum anisotropic hyperfine couplings for each of the radicals in (FAD^{•-} - TrPH^{•+}): N₅, N₁₀, and H₆ for FAD^{•-}, and N₁, H₁, and H_{\beta1} for TrPH^{•+}) [60]. The used $J_{\rm ex}^{\zeta}$ and D^{ζ} couplings correspond to FAD^{•-} - TrPH^{•+} RP in *Drosophila melanogaster* cryptochrome protein [59]. The recombination rate is assumed to be $k = 2 \times 10^5$ Hz, which is of the order of the observed spin relaxation time in behavioural studies in migratory birds [73]. To calculate the total signal received (or phase accumulated by the NV center) at a given strength and direction of the applied magnetic field,

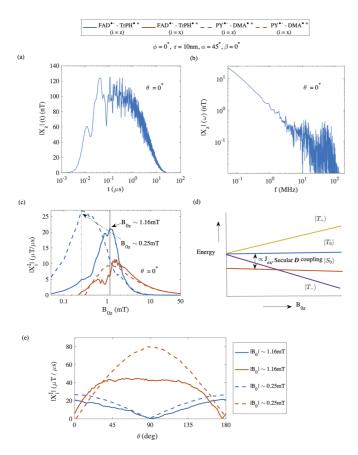


FIG. 5. Signal received by the NV center from a single RP molecule of model systems FAD•- - TrPH•+ (solid lines) and PY•- - DMA•+ RP) (dashed lines). Time (a) and frequency (b) profile of the signal at magnetic field $B_{0z}=1.16 \mathrm{mT}$ for FAD•- - TrPH•+ RP, where maximum of time integrated signal $|X_z^I|$ and $|X_x^I|$ is obtained as plotted in (c). $\theta=0^\circ$ is assumed in (a), (b), and (c). (d) Level dynamics of the RP in singlet-triplet basis as a function of the strength of the applied magnetic field. (e) Dependence of the integrated signal on the angle of the applied magnetic field at magnetic field strength $|B_0|=1.16 \mathrm{mT}$ and $|B_0|=0.25 \mathrm{mT}$, respectively, for FAD•- - TrPH•+ and PY•- - DMA•+ RP.

we define the time integrated signal per second as

$$X_i^I(r,\zeta) = \frac{1}{T} \sum_t X_i(r,\zeta,t), \tag{14}$$

where T is the total time of the sequence. In Fig. 5, we show the strength of the magnetic signal received by the NV from an RP on a single molecule at a distance r=10 nm. For such shallow NV centres, the achievable sensitivity is of the order of a few nT/ $\sqrt{\rm Hz}$ to a slowly alternating signal [66,74]. For the case of $\beta=0$, as we assume here, the detectable signal is only generated by X_x^I and X_z^I , which, respectively, corresponds to the dynamics of RP density matrix elements $S_{1x}+S_{2x}$ and $S_{1z}+S_{2z}$.

To determine the suitable NV sensing sequence to probe the generated signal, Fig. 5(a) plots the temporal evolution of $|X_z|(t)$ for a bias magnetic field that maximizes the time integrated signal $|X_z^I|$ [see Fig. 5(c)]. The unpaired electrons in the RP are initialized in the singlet-state whose population

decays with time as it is converted into other density matrix elements under the evolution of $H_{\rm RP}^{\zeta}$, thus resulting in the build-up of $|X_z^I|$. The generated signal (hundreds of nT) is within the sensitivity achieved for <10 nm deep NV centres. The corresponding spectrum is shown in Fig. 5(b). Because we are interested in low frequency dynamics (kHz to tens of MHz) resulting from the dipolar and hyperfine coupling terms (typically 1–10 mT) in $H_{\rm RP}^{\zeta}$, the maximum component of the signal in the frequency domain is limited to 500 MHz. The cutoff frequency of these oscillations observed is in the order of 10MHz. Sequences including Ramsey, spin-echo, and dynamical decoupling are thus suitable for probing such RP dynamics [67].

Variation with strength of the magnetic field. To study the dependence of the signal on the strength of the applied magnetic field, we set $\theta=0$, i.e., $B_{0x}=0$ and $B_{0y}=0$ so that there is no spin mixing of the NV center states keeping the two-level approximation valid and $B_{0z}(=B_z)$ is varied from 0 to 50 mT. The dependence of the time-integrated signal $|X_z^I|$ and $|X_x^I|$ is plotted as a function of B_{0z} in Fig. 5(c).

The features of the time-integrated signal $|X_z^I|$ can be qualitatively explained by the dynamics of the RP under H_{RP}^{ζ} in singlet-triplet basis as shown in Fig. 5(d). At $B_{0z} \approx 0$, all three triplet states are almost degenerate and the energy gap between $|S_0\rangle$ and $|T_0\rangle$ states is proportional to $J_{\rm ex}^{\zeta}$ and the secular part of D^{ζ} . With the unpaired electrons in the RP initialized in the singlet state, population transfer between the spin states occurs due to the evolution of the RP under $H_{\rm RP}^{\zeta}$ containing hyperfine interactions with the surrounding nuclei [Eq. (4)]. However, for small B_{0z} , the RP only generates a very low signal $|X_z^I|$ as the outer triplet states $(|T_{\pm}\rangle)$ remain close to degeneracy. As B_{0z} increases, the outer triplet states move apart and give rise to additional manifolds for singlet-triplet mixing [75,76]. This opens the possibility of information (population and coherence) transfer among $|S_0\rangle$ and $|T_0\rangle$ states to only one of the outer triplet states, creating a population imbalance between outer triplet states and giving rise to an increase in signal $|X_z^I|$. At a certain B_{0z} value, there is a maximum transfer between singlet-triplet oscillations to population difference of outer triplet states, giving rise to the peaklike features in $|X_z^I|$ (for example the peak at ~ 1.16 and $\sim 0.25 \,\mathrm{mT}$, respectively, for FAD $^{\bullet-}$ - TrPH $^{\bullet+}$ RP and $PY^{\bullet -}$ - DMA $^{\bullet +}$ RP). A further increase in B_{0z} results in energetic isolation of the outer triplet states which causes steady decrease in the probability of information transfer between them, and $|S_0\rangle$ and $|T_0\rangle$ states, eventually vanishing at very high magnetic fields $|B_{0z}| \gg |H_{\rm RP}^{\zeta}|/\gamma_e$. A similar argument as above can be used to qualitatively explain features in the dynamics of $|X_r^I|$.

Variation with direction of the magnetic field. In Fig. 5(e), we investigate the signal dependence on the direction of the applied magnetic field. Here we restrict the magnetic field to the XZ plane by assuming $\phi=0$ as the following discussion holds for the general case. Although B_{0x} is nonzero for certain values of θ , its magnitude should be less than that required for mixing of the $|1\rangle$ and $|-1\rangle$ levels of the NV center. We fix the magnetic field strength to ~ 1.16 and ~ 0.25 mT, respectively, for FAD*- - TrPH*- RP and PY*- - DMA*+ RP where the maximum MFE is expected based on the results shown in

Fig. 5(c). The dependence of the time-integrated signal on θ is plotted in Fig. 5(e).

Once again, using the level dynamics under $H_{\rm RP}^{\zeta}$ in the singlet-triplet basis, a qualitative explanation of various features of the generated signals is possible using the following arguments: (i) for a given magnetic field, θ close to 90° essentially means presence of an extra channel for singlet-triplet mixing due to nonzero B_{0x} and B_{0y} and (ii) at θ close to 90° , the outer triplet states are almost degenerate while maximum splitting occurs at θ close to 0° or 180° . The shape of $|X_x^I|$ is a direct consequence of argument (i) as nonzero transverse fields close to 90° increase the probability of singlet-triplet as well as triplet-triplet mixing. The signal $|X_z^I|$ is a consequence of argument (ii) as higher splitting of outer triplet states opens new manifolds for information transfer between them and $|S_0\rangle$ and $|T_0\rangle$ states as described earlier.

For parameter values chosen here and described above, we do not observe the recently discovered spike feature [60] in the generated signals $|X_z^I|$ and $|X_x^I|$ presumably due to the high value of magnetic field used, and the nonzero exchange and dipolar coupling in the H_{RP}^{ζ} [77]. However, the spike feature can be observed when we consider a simpler RP system with only one nuclear spin in each RP in earth's magnetic field as summarized in Appendix A 2.

2. Ensemble of RPs

Assuming the minimum size of the protein to be 3-4 nm (small protein with molecular weight <50 kDa [72]), \approx 100 RPs ($\xi = 5 \times 10^{-2} \text{ nm}^{-3}$) might fall in the NV sensing volume. In contrast, standard absorption or EPR based studies require orders of magnitude larger number of molecules (micro litres of sample) in order to get observable signals which might cause masking of certain features of the RPM due to averaging [44]. In Fig. 6, we plot the mean and variance of the generated signal as a function of strength and direction of applied magnetic field for an ensemble of RPs, where we neglected dipolar coupling between RPs on different molecules. Such dipolar couplings are expected to give rise to altered RP dynamics and even yield unique magnetic field effects. We consider two types of ensembles. (i) All the RPs are oriented along the direction of the NV axis, i.e., $\mathcal{R}^{\zeta} = I$ (solid lines in Fig. 6), and (ii) all RPs are randomly oriented with respect to the NV axis (dashed lines in Fig. 6), i.e., $\mathcal{R}^{\zeta} = R_{x}(\gamma_{1})R_{y}(\gamma_{2})R_{z}(\gamma_{3})$, where $R_{x}(\gamma_{1}), R_{y}(\gamma_{2})$, and $R_z(\gamma_3)$, respectively, are rotation matrices about the x, y, and z axes with randomly chosen Euler angles γ_1 , γ_2 , and γ_3 . For both cases, r, α , and β were sampled uniformly from the range [5–20 nm], $[0, \pi/2]$, and $[0, 2\pi]$, respectively, assuming spherical geometry for the distribution of RPs.

For the uniformly oriented ensemble (i), the mean of the generated signal is obviously amplified in contrast to the randomly oriented ensemble in (ii) where the features due to the RP spin dynamics are almost entirely removed because of random averaging of the signal over various realizations of \mathcal{R}^{ζ} . The variance of the generated signal as a function of strength of the magnetic field is within the sensitivity range of a shallow NV center for both types of ensembles and the MFE behavior akin to the single-molecule case [Fig. 5(c)] is still observable after averaging. On the contrary, although there is

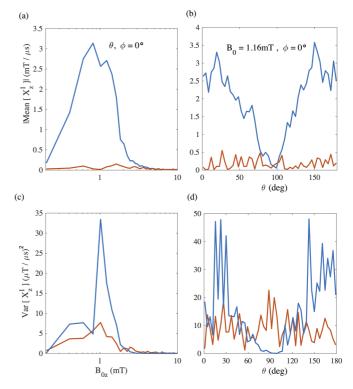


FIG. 6. Dynamics of the statistical signal generated by an ensemble of RPs in the sensing volume of the NV center. To reduce computation time, two nuclear spins were included for each radical: N_5 , N_{10} for $FAD^{\bullet-}$, and N_1 , H_1 for $TrPH^{\bullet+}$. MFE against variation in [(a) and (c)] strength of magnetic field [(b) and (d)] direction of the magnetic field. The blue line indicate the average MFE when the coordinate frames of all the RPs are aligned ($\mathcal{R}^{\varsigma} = I$) with the NV frame, while the red lines show the average MFE from a randomly oriented ensemble (mean and variance of 50 random realizations of \mathcal{R}^{ς}).

an observable variance signal when θ is varied, the shape of the variation is erratic for ensemble (ii) with no similarity to the single molecule case [Fig. 5(f)]. Therefore, although the ensemble (ii) is simply realized by drop casting a molecule sample on the surface of the diamond, detecting a MFE in this arrangement is more challenging. The situation might be compared to absorption or EPR studies where the size of the probed ensemble is very large (micro litres of sample) and hence MFE detection relies on statistical signals generated from a very small number of molecules. This limits the detection of MFE to a few percent [35] along with causing wastage of precious biological samples. Hence, it becomes important to appropriately position the proteins in the desired orientation with respect to the NV center to maximize the probability of successful detection of an MFE (Appendix A 1).

C. Strong coupling regime

In the strong coupling regime, the energy levels of the NV and RP mix, giving rise to a level structure as illustrated in Fig. 7. Due to the s=1 structure of the NV center, the RP components of eigenstates in $|0\rangle$ and $|1\rangle$ are quantized along different axes determined by the strength of the coupling tensor. The total Hamiltonian in this regime can be

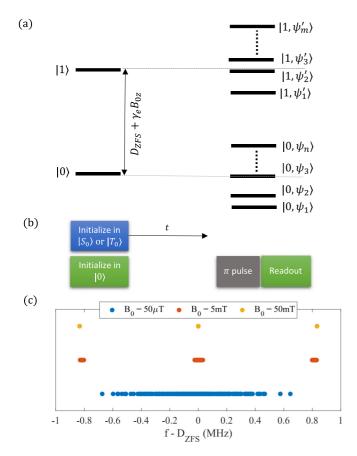


FIG. 7. (a) Energy level structure in the strong coupling regime. The RP components of an eigenstate in the $|0\rangle$ manifold $(|\psi_i\rangle)$ are determined by $H_{\rm RP}^\zeta$ alone whereas in the eigenstate $|1\rangle$ manifold, $(|\psi_i'\rangle)$ are determined by both $H_{\rm RP}^\zeta$ and H_c together. The sequence to detect RP dynamics in this regime is shown in (b). (c) The number (each circle represent one peak) of peaks in the spin resonance spectrum of the NV center resulting from interaction of the NV center with a single RP as a function of applied magnetic field. Here we only considered the resonances resulting for the case n=m, i.e., transitions corresponding to $|+1,\psi_n'\rangle \rightarrow |0,\psi_n\rangle$.

reduced to

$$H = H_{\text{NV}} + H_{\text{RP}}^{\zeta} |0\rangle\langle 0| + \left[H_{\text{RP}}^{\zeta} + D_r \sum_{i=x,y,z} \tilde{S}_i \right] |1\rangle\langle 1|. \quad (15)$$

Depending on the magnitude of various couplings in H_{RP}^{ζ} and type of the nuclei in the RP, the $|0\rangle$ and $|1\rangle$ states of the NV center are further split, as shown in Fig. 7(a). For example, for the RP consisting of only spin -1/2 nuclei, NV center levels split into a maximum of 2^{N+2} levels. The level structure results in resonance lines in the magnetic resonance spectrum of the NV center corresponding to transitions $|+1, \psi'_m\rangle \rightarrow |0, \psi_n\rangle$, where $|\psi'_m\rangle$ and $|\psi_n\rangle$ are respectively eigenstates of H_{RP}^{ζ} and $H_{\text{RP}}^{\zeta} + D_r \sum_i \tilde{S}_i$. The magnitude of the resonance lines is proportional to the transition dipole moment and population difference of the states connected by the transition. Therefore monitoring it using a simple sequence as shown in Fig. 7(b) may facilitate a probing of the dynamics of the populations of H_{RP}^{ζ} .

There are a couple of important details about the detection of RP dynamics in this regime which should be pointed out.

- (1) Since we wish to study RP dynamics exclusively under an applied magnetic field, a pulsed sensing sequence [Fig. 7(b)] is preferred as it ensures that the NV center stays in the $|0\rangle$ state during the time of evolution, implying no backaction of the NV on the RP. In a Ramsay type of sequence where the NV center is prepared in $(|0\rangle + |1\rangle)/\sqrt{2}$, the RP experiences an additional induced field from the $|1\rangle$ state component [44] which is different for each RP, complicating the probing of MFEs.
- (2) As the size of the biomolecule increases, the number of unavoidable hyperfine interactions also grows. As a consequence, the spectral features will start to overlap, especially when the applied magnetic fields are comparable to the hyperfine couplings in $H_{\rm RP}^{\zeta}$ [see Fig. 7(c) for FAD*-TrPH*]. In this case, it becomes challenging to access the various electronic transitions of the RP separately. One could consider decoupling the nuclear spin bath [78,79] during the readout time by driving the RP at a frequency larger than the strongest hyperfine coupling, however the short readout duration (typically ≈ 300 ns for an NV center [45,80]) will render the averaging inefficient.

In the light of the second argument, although the detection of RP dynamics in the strong coupling regime may be challenging, it offers an opportunity to track various populations of the RP density matrix individually and the possibility of quantum control of the RP using an NV center [44].

IV. CONCLUSION AND OUTLOOK

In summary, we analysed the realistic prospects of MFE detection at the single molecule scale in RP reactions in biomolecules using a single NV center in diamond. To realistically describe the biomolecular spin dynamics, our RP model includes three nuclear spins per radical that have the largest hyperfine couplings. Including dipolar and exchange coupling between the two electrons forming the radical pair, the RP spin dynamics become correlated and can not be simulated by treating each of the radicals independently [43]. Depending on distance and orientation between NV and RP, two coupling regimes of dynamics can be considered requiring accordingly tailored NV sensing approaches. We find that in the weak coupling regime, RP dynamics can be seen as a classical magnetic field by the NV. This regime is most suitable for large biomolecules with unavoidable and spectrally indistinguishable spin interactions. The signal generated even from a single RP is well within the sensitivity achievable by state of the art NV centers and distinct features of RP spin dynamics are thus observable at the single molecule and a small ensemble $(\leq 100 \text{ molecules}) \text{ scale.}$

The simulations performed in our work can be further improved by incorporating more nuclear spins and using the recently developed coherent state sampling method [81,82] for efficient computation of spin dynamics. Adding to the recent theoretical works [42–44], we expect our analysis to pave the way towards experimental detection of MFE in biomolecules at the single molecule level and unravel the importance of quantum coherent effects in biochemical processes [20,21].

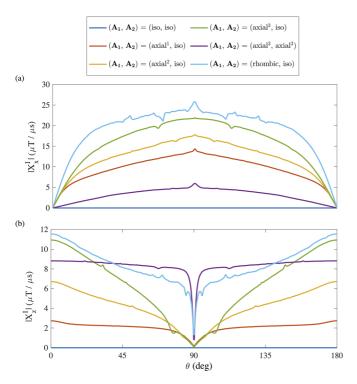


FIG. 8. The generated signal $|X_x^I|$ and $|X_z^I|$ for various types of hyperfine coupling tensors for the model system. Here "iso," "axial¹," "axial²," "axial³," and "rhombic" stand for hyperfine principle components (A_{xx}, A_{yy}, A_{zz}) , respectively, (0.5, 0.5, 0.5), (-0.09, -0.09, 1.76), (-0.2, -0.2, 1.76), (-0.39, -0.39, 1.76), and (-0.39, 0, 1.76) mT. The exchange coupling is chosen to be $J_{\rm ex}=0.375$ mT and diplolar coupling tensor $(\textbf{\textit{D}})$ is assumed to be zero as the effect is similar to exchange coupling. The lifetime of the RP is 5 μ s.

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APPENDIX

1. Orientation of RP with respect to NV

As described in the main text, the orientation of the molecule hosting the RP is crucial for obtaining maximum signal. Recent studies involving investigation of structure and motion of single proteins using NV centers have employed either statistical placement of proteins via diamond surface functionalization [66,83,84] or controlled alignment via attaching the protein to a solid host and then using a nanopositioning system, for example, a tuning fork scanning probe of an atomic force microscope [85]. The latter method is preferred because, in the former case, there are two main

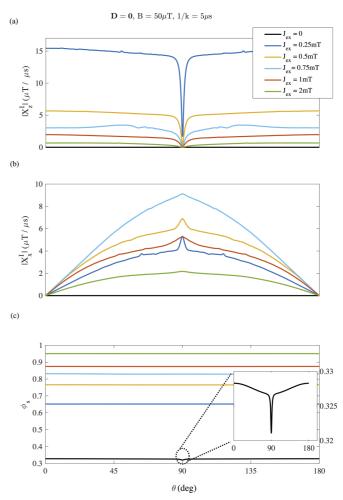


FIG. 9. The generated signal $|X_x^I|$ and $|X_z^I|$, and singlet yield [86] for various values of exchange coupling for the model system. Here $A_1 = A_2$ and chosen according to the case axial³ (see caption of Fig. 8).

disadvantages: (i) the RP might end up in an orientation where one or more coupling parameters $(D_{ci}, i = x, y, z)$ are zero, thereby reducing the coupling to the NV center (ii). The functionalization of the diamond surface may not be possible for all types of proteins.

2. Spike features in the generated signal

Here we consider a simple RP model system with only one nuclear spin in each radical (we drop the ζ notation for convenience in the following), with hyperfine tensors A_1 and A_2 , to analyze which parameter ranges give rise to spikelike features of the generated signal as a function of the direction of the applied magnetic field. This feature has been suggested to be behind the precision of an avian magnetic compass [60].

In Fig. 8, we plot the generated signal $|X_x^I|$ and $|X_z^I|$, respectively, in (a) and (b) for various types of hyperfine coupling tensors chosen by varying principal axis components set (A_{xx}, A_{yy}, A_{zz}) . For the anisotropic hyperfine coupling case, the principal axis system is that of the N₅ in FAD^{• [60]}. A qualitative description of the seen behavior can be provided by looking at the symmetry of the RP Hamiltonian.

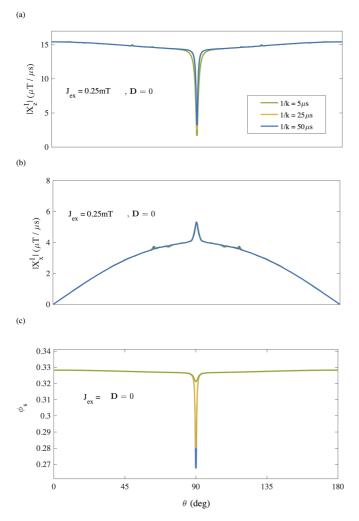


FIG. 10. The generated signal $|X_x^I|$ and $|X_z^I|$, and singlet yield for increasing values of RP lifetime. Here $B_0 = 50 \,\mu\text{T}$ and $A_1 = A_2$ and chosen according to the case axial³ (see caption of Fig. 8).

In the absence of dipolar coupling, there are three interactions in the RP Hamiltonian, namely, Zeeman, exchange and hyperfine. Zeeman (same magnetic field on all the spins) and exchange interactions are symmetrical with respect to electronic spin exchange while hyperfine interaction is symmetry breaking in general. When the hyperfine interaction is also assumed to be isotropic and $A_1 = A_2$, the full Hamiltonian is symmetric, and the singlet and triplet states are eigenstates. As a result, there is no singlet-triplet oscillations possible for isotropic hyperfine coupling as the sub-spaces of different symmetry remain unconnected and as consequence, $|X_{\rm r}^I|$ and $|X_{\tau}^{I}|$ remains zero. As the anisotropy is included in the Hamiltonian, a nonzero signal is generated along with appearance of spike features with the most prominent around $\theta = 90^{\circ}$. Again, the amplitude of this spike is within the sensitivity achieved for <10 nm deep NV centres. Now we analyze the behavior of the generated signal and spike as a function of various parameters of the RP Hamiltonian.

The amplitude of the spike increases and it becomes narrower as the principal components A_{xx} and A_{yy} are increased. This observation hints that the origin of the spike might also be similar, i.e., an avoided crossing of energy levels as a function of direction of the magnetic field. However, further investigations are required to draw firm conclusions.

In Fig. 9, we show the generated signal $|X_x^I|$ and $|X_z^I|$ along with the singlet yield $\phi_s = kdt \sum_t \text{Tr}[\rho_{RP}(t)|S_0\rangle\langle S_0|]$ [86] (c) with increasing value of J_{ex} . Using the symmetry argument, as J_{ex} is increased, the Hamiltonian eigenstate get closer to singlet and triplet states, and reduction (increase) in the signal (singlet yield) is observed. The spike feature in the generated signal at $\theta = 90^\circ$ is most prominent when J_{ex} is comparable to the A_{xx} and A_{yy} principal component of the hyperfine coupling. This behavior hints that the spike in the generated signal depends on exchange interaction (and also dipolar in general) as well as anisotropic hyperfine interaction. This is in contrast to the singlet yield where the spike depends strongly on hyperfine anisotropy as it disappears even for small J_{ex} , comparable to A_{xx} and A_{yy} parameters [77].

In Fig. 10, we show the generated signal $|X_z^I|$ and $|X_z^I|$ along with the singlet yield [86] (c) with increasing value of RP lifetime τ . A contrasting behavior is observed in the generated signal and singlet yield in addition to the former being relatively less sensitive to the lifetime of the radical. This observation seems to suggest that unpaired electrons prefer to stay in the singlet state compared to other populations and coherences of the RP. However, again, further studies are required in this direction to draw concrete conclusions which are beyond purview of current work.

3. Experimental parameters

Here, we provide an order of magnitude estimation of the parameters for acquisition of MFE effects, that would be required applying a diamond NV sensor. We consider a single NV center shallow implanted below the surface of a flat diamond sample that yields a photon count rate of $\approx 100 \text{ k/s}$ and a spin readout contrast of $\approx 30\%$. The sensitivity of a Ramsey sequence can be estimated via [87]

$$\eta_{DC} = \frac{\hbar}{g\mu_B} \sigma_R \sqrt{\frac{\tau + t_I + t_R}{\tau^2}},\tag{A1}$$

where \hbar is the reduced Planck's constant, g is the Lande factor, μ_B is the Bohr magneton, and $t_I \approx 1$ μs and $t_R \approx 1$ μs are NV initialization and readout times. For such configuration, the spin-readout noise $\sigma_R \approx 30$ (in units of the spin projection noise) and with $\tau = T_2^*$, a sensitivity $\eta_{DC} \approx 1$ $\mu THz^{-1/2}$ is obtained. Thus a total acquisition time $T \approx 20$ s (corresponding to 5×10^6 single repetitions) is required to resolve a field with an amplitude around 500 nT. Depending on resolution, an experimental sweep will thus take at least several minutes.

^[1] U. E. Steiner and T. Ulrich, Magnetic field effects in chemical kinetics and related phenomena, Chem. Rev. **89**, 51 (1989).

^[2] J. R. Woodward, Radical pairs in solution, Prog. React. Kinet. Mech. 27, 165 (2002).

- [3] C. R. Timmel and K. B. Henbest, A study of spin chemistry in weak magnetic fields, Philos. Trans. R. Soc. London. Series A: Math. Phys. Eng. Sci. 362, 2573 (2004).
- [4] H. Hayashi, Introduction to Dynamic Spin Chemistry: Magnetic Field Effects on Chemical and Biochemical Reactions (World Scientific, Singapore, 2004), Vol. 8.
- [5] P. J. Hore, K. L. Ivanov, and M. R. Wasielewski, Spin chemistry, J. Chem. Phys. 152, 120401 (2020).
- [6] J. H. Lee, Y. Okuno, and S. Cavagnero, Sensitivity enhancement in solution NMR: Emerging ideas and new frontiers, J. Magn. Reson. 241, 18 (2014).
- [7] B. K. Rugg, M. D. Krzyaniak, B. T. Phelan, M. A. Ratner, R. M. Young, and M. R. Wasielewski, Photodriven quantum teleportation of an electron spin state in a covalent donor– acceptor–radical system, Nat. Chem. 11, 981 (2019).
- [8] P. J. Hore and H. Mouritsen, The radical-pair mechanism of magnetoreception, Annu. Rev. Biophys. 45, 299 (2016).
- [9] M. T. Colvin, A. B. Ricks, A. M. Scott, A. L. Smeigh, R. Carmieli, T. Miura, and M. R. Wasielewski, Magnetic field-induced switching of the radical-pair intersystem crossing mechanism in a donor- bridge- acceptor molecule for artificial photosynthesis, J. Am. Chem. Soc. 133, 1240 (2011).
- [10] C. T. Rodgers and P. J. Hore, Chemical magnetoreception in birds: The radical pair mechanism, Proc. Natl. Acad. Sci. USA 106, 353 (2009).
- [11] E. W. Evans, C. A. Dodson, K. Maeda, T. Biskup, C. J. Wedge, and C. R. Timmel, Magnetic field effects in flavoproteins and related systems, Interface Focus 3, 20130037 (2013).
- [12] A. R. Jones, Magnetic field effects in proteins, Mol. Phys. 114, 1691 (2016).
- [13] R. Kaptein and L. J. Oosterhoff, Chemically induced dynamic nuclear polarization III (anomalous multiplets of radical coupling and disproportionation products), Chem. Phys. Lett. 4, 214 (1969).
- [14] R. Kaptein and J. L. Oosterhoff, Chemically induced dynamic nuclear polarization II: (relation with anomalous ESR spectra), Chem. Phys. Lett. 4, 195 (1969).
- [15] H. R. Ward and R. G. Lawler, Nuclear magnetic resonance emission and enhanced absorption in rapid organometallic reactions, J. Am. Chem. Soc. **89**, 5518 (1967).
- [16] R. W. Fessenden and R. H. Schuler, Electron spin resonance studies of transient alkyl radicals, J. Chem. Phys. **39**, 2147 (1963)
- [17] G. L. Closs, Mechanism explaining nuclear spin polarizations in radical combination reactions, J. Am. Chem. Soc. **91**, 4552 (1969).
- [18] G. L. Closs and A. D. Trifunac, Theory of chemically induced nuclear spin polarization. III. effect of isotropic g shifts in the components of radical pairs with one hyperfine interaction, J. Am. Chem. Soc. 92, 2183 (1970).
- [19] S. K. Wong, D. A. Hutchinson, and J. K. S. Wan, Chemically induced dynamic electron polarization. II. A general theory for radicals produced by photochemical reactions of excited triplet carbonyl compounds, J. Chem. Phys. 58, 985 (1973).
- [20] Y. Kim, F. Bertagna, E. M. D'Souza, D. J. Heyes, L. O. Johannissen, E. T. Nery, A. Pantelias, A. S.-P. Jimenez, L. Slocombe, M. G. Spencer *et al.*, Quantum biology: An update and perspective, Quantum Rep. 3, 80 (2021).
- [21] A. Marais, B. Adams, A. K. Ringsmuth, M. Ferretti, J. M. Gruber, R. Hendrikx, M. Schuld, S. L. Smith, I. Sinayskiy,

- T. P. J. Krüger *et al.*, The future of quantum biology, J. R. Soc. Int. **15**, 20180640 (2018).
- [22] P. Emery, R. Stanewsky, J. C. Hall, and M. Rosbash, A unique circadian-rhythm photoreceptor, Nature (London) 404, 456 (2000).
- [23] S. Panda, J. B. Hogenesch, and S. A. Kay, Circadian rhythms from flies to human, Nature (London) 417, 329 (2002).
- [24] R. Wiltschko, C. Nießner, and W. Wiltschko, The magnetic compass of birds: The role of cryptochrome, Front. Physiol. 12, 667000 (2021).
- [25] A. Marais, I. Sinayskiy, F. Petruccione, and R. van Grondelle, A quantum protective mechanism in photosynthesis, Sci. Rep. 5, 8720 (2015).
- [26] I. K. Kominis, Quantum measurement corrections to cidnp in photosynthetic reaction centers, New J. Phys. 15, 075017 (2013).
- [27] V. Joosten and W. J. H. van Berkel, Flavoenzymes, Current Opin. Chem. Biology 11, 195 (2007).
- [28] P. Pimviriyakul and P. Chaiyen, Overview of flavin-dependent enzymes, *The Enzymes* (Elsevier, 2020), Vol. 47, pp. 1–36.
- [29] H. L. Messiha, T. Wongnate, P. Chaiyen, A. R. Jones, and N. S. Scrutton, Magnetic field effects as a result of the radical pair mechanism are unlikely in redox enzymes, J. R. Soc. Inter. 12, 20141155 (2015).
- [30] D. Crotty, G. Silkstone, S. Poddar, R. Ranson, A. Prina-Mello, M. T. Wilson, and J. M. D. Coey, Reexamination of magnetic isotope and field effects triphosphate adenosine production bv creatine kinase. Proc. Natl. Acad. Sci. USA 109, (2012).
- [31] R. Bittl and S. Weber, Transient radical pairs studied by time-resolved EPR, Biochim. Biophys. Acta Bioenerg. **1707**, 117 (2005).
- [32] T. Biskup, Time-resolved electron paramagnetic resonance of radical pair intermediates in cryptochromes, Mol. Phys. 111, 3698 (2013).
- [33] K. B. Henbest, K. Maeda, P. J. Hore, M. Joshi, A. Bacher, R. Bittl, S. Weber, C. R. Timmel, and E. Schleicher, Magnetic-field effect on the photoactivation reaction of *Escherichia coli* DNA photolyase, Proc. Natl. Acad. Sci. USA 105, 14395 (2008).
- [34] K. Maeda, S. R. T. Neil, K. B. Henbest, S. Weber, E. Schleicher, P. J. Hore, S. R. Mackenzie, and C. R. Timmel, Following radical pair reactions in solution: A step change in sensitivity using cavity ring-down detection, J. Am. Chem. Soc. 133, 17807 (2011).
- [35] K. Maeda, A. J. Robinson, K. B. Henbest, H. J. Hogben, T. Biskup, M. Ahmad, E. Schleicher, S. Weber, C. R. Timmel, and P. J. Hore, Magnetically sensitive light-induced reactions in cryptochrome are consistent with its proposed role as a magnetoreceptor, Proc. Natl. Acad. Sci. USA 109, 4774 (2012).
- [36] D. M. W. Sheppard, J. Li, K. B. Henbest, S. R. T. Neil, K. Maeda, J. Storey, E. Schleicher, T. Biskup, R. Rodriguez, S. Weber *et al.*, Millitesla magnetic field effects on the photocycle of an animal cryptochrome, Sci. Rep. **7**, 42228 (2017).
- [37] C. A. Dodson, C. J. Wedge, M. Murakami, K. Maeda, M. I. Wallace, and P. J. Hore, Fluorescence-detected magnetic field effects on radical pair reactions from femtolitre volumes, Chem. Commun. 51, 8023 (2015).
- [38] E. W. Evans, J. Li, J. G. Storey, K. Maeda, K. B. Henbest, C. A. Dodson, P. J. Hore, S. R. Mackenzie, and C. R. Timmel,

- Sensitive fluorescence-based detection of magnetic field effects in photoreactions of flavins, Phys. Chem. Chem. Phys. 17, 18456 (2015).
- [39] D. R. Kattnig, E. W. Evans, V. Déjean, C. A. Dodson, M. I. Wallace, S. R. Mackenzie, C. R. Timmel, and P. J. Hore, Chemical amplification of magnetic field effects relevant to avian magnetoreception, Nat. Chem. 8, 384 (2016).
- [40] V. Déjean, M. Konowalczyk, J. Gravell, M. J. Golesworthy, C. Gunn, N. Pompe, O. F. V. Elst, K.-J. Tan, M. Oxborrow, D. G. A. L. Aarts *et al.*, Detection of magnetic field effects by confocal microscopy, Chem. Sci. 11, 7772 (2020).
- [41] N. Ikeya and J. R. Woodward, Cellular autofluorescence is magnetic field sensitive, Proc. Natl. Acad. Sci. USA 118, e2018043118 (2021).
- [42] N. Ikeya, E. A. Nasibulov, K. L. Ivanov, K. Maeda, and J. R. Woodward, Single-molecule spectroscopy of radical pairs, a theoretical treatment and experimental considerations, Mol. Phys. 117, 2604 (2019).
- [43] H. Liu, M. B. Plenio, and J. Cai, Scheme for detection of single-molecule radical pair reaction using spin in diamond, Phys. Rev. Lett. 118, 200402 (2017).
- [44] A. Finkler and D. Dasari, Quantum sensing and control of spinstate dynamics in the radical-pair mechanism, Phys. Rev. Appl. 15, 034066 (2021).
- [45] M. W. Doherty, N. B. Manson, P. Delaney, F. Jelezko, J. Wrachtrup, and L. C. L. Hollenberg, The nitrogen-vacancy colour centre in diamond, Phys. Rep. 528, 1 (2013).
- [46] R. Schirhagl, K. Chang, M. Loretz, and C. L. Degen, Nitrogen-vacancy centers in diamond: nanoscale sensors for physics and biology, Annu. Rev. Phys. Chem. **65**, 83 (2014).
- [47] D. A. Simpson, Quantum probes for biology: Unlocking single molecule dynamics, Nano Today 24, 7 (2019).
- [48] G. Balasubramanian, I. Y. Chan, R. Kolesov, M. Al-Hmoud, J. Tisler, C. Shin, C. Kim, A. Wojcik, P. R. Hemmer, A. Krueger *et al.*, Nanoscale imaging magnetometry with diamond spins under ambient conditions, Nature (London) 455, 648 (2008).
- [49] J. R. Maze, P. L. Stanwix, J. S. Hodges, S. Hong, J. M. Taylor, P. Cappellaro, L. Jiang, M. V. Dutt, E. Togan, A. S. Zibrov *et al.*, Nanoscale magnetic sensing with an individual electronic spin in diamond, Nature (London) 455, 644 (2008).
- [50] J. M. Taylor, P. Cappellaro, L. Childress, L. Jiang, D. Budker, P. R. Hemmer, A. Yacoby, R. Walsworth, and M. D. Lukin, High-sensitivity diamond magnetometer with nanoscale resolution, Nat. Phys. 4, 810 (2008).
- [51] F. Dolde, H. Fedder, M. W. Doherty, T. Nöbauer, F. Rempp, G. Balasubramanian, T. Wolf, F. Reinhard, L. C. L. Hollenberg, F. Jelezko *et al.* Electric-field sensing using single diamond spins, Nat. Phys. 7, 459 (2011).
- [52] F. Dolde, M. W. Doherty, J. Michl, I. Jakobi, B. Naydenov, S. Pezzagna, J. Meijer, P. Neumann, F. Jelezko, N. B. Manson *et al.*, Nanoscale detection of a single fundamental charge in ambient conditions using the NV⁻ center in diamond, Phys. Rev. Lett. 112, 097603 (2014).
- [53] D. Suter and F. Jelezko, Single-spin magnetic resonance in the nitrogen-vacancy center of diamond, Prog. Nucl. Magn. Reson. Spectrosc. **98**, 50 (2017).
- [54] L. M. Pham, S. J. DeVience, F. Casola, I. Lovchinsky, Alexander O. Sushkov, E. Bersin, J. Lee, E. Urbach, P. Cappellaro, H. Park, A. Yacoby, M. Lukin, and Ronald L.

- Walsworth, Nmr technique for determining the depth of shallow nitrogen-vacancy centers in diamond, Phys. Rev. B **93**, 045425 (2016).
- [55] M. L. Goldman, M. W. Doherty, A. Sipahigil, N. Y. Yao, S. D. Bennett, N. B. Manson, A. Kubanek, and M. D. Lukin, State-selective intersystem crossing in nitrogen-vacancy centers, Phys. Rev. B 91, 165201 (2015).
- [56] S. Felton, A. M. Edmonds, M. E. Newton, P. M. Martineau, D. Fisher, D. J. Twitchen, and J. M. Baker, Hyperfine interaction in the ground state of the negatively charged nitrogen vacancy center in diamond, Phys. Rev. B 79, 075203 (2009).
- [57] K. Schwinn, N. Ferré, and M. Huix-Rotllant, Uv-visible absorption spectrum of fad and its reduced forms embedded in a cryptochrome protein, Phys. Chem. Chem. Phys. 22, 12447 (2020).
- [58] M. Kruszewski, Labile iron pool: The main determinant of cellular response to oxidative stress, Mutat. Res. - Fundam. Mol. Mec. 531, 81 (2003).
- [59] T. P. Fay, L. P. Lindoy, D. E. Manolopoulos, and P. J. Hore, How quantum is radical pair magnetoreception? Faraday Discuss. 221, 77 (2020).
- [60] H. G. Hiscock, S. Worster, D. R. Kattnig, C. Steers, Y. Jin, D. E. Manolopoulos, H. Mouritsen, and P. J. Hore, The quantum needle of the avian magnetic compass, Proc. Natl. Acad. Sci. USA 113, 4634 (2016).
- [61] O. Efimova and P. J. Hore, Role of exchange and dipolar interactions in the radical pair model of the avian magnetic compass, Biophys. J. 94, 1565 (2008).
- [62] N. Zhao, J. Honert, B. Schmid, M. Klas, J. Isoya, M. Markham, D. Twitchen, F. Jelezko, R.-B. Liu, H. Fedder *et al.*, Sensing single remote nuclear spins, Nat. Nanotechnol. 7, 657 (2012).
- [63] J. Zopes, K. S. Cujia, K. Sasaki, J. M. Boss, K. M. Itoh, and C. L. Degen, Three-dimensional localization spectroscopy of individual nuclear spins with sub-angstrom resolution, Nat. Commun. 9, 4678 (2018).
- [64] J. Zopes, K. Herb, K. S. Cujia, and C. L. Degen, Threedimensional nuclear spin positioning using coherent radiofrequency control, Phys. Rev. Lett. 121, 170801 (2018).
- [65] A. Laraoui, D. Pagliero, and C. A. Meriles, Imaging nuclear spins weakly coupled to a probe paramagnetic center, Phys. Rev. B 91, 205410 (2015).
- [66] F. Shi, Q. Zhang, P. Wang, H. Sun, J. Wang, X. Rong, M. Chen, C. Ju, F. Reinhard, H. Chen *et al.*, Single-protein spin resonance spectroscopy under ambient conditions, Science 347, 1135 (2015).
- [67] C. L. Degen, F. Reinhard, and P. Cappellaro, Quantum sensing, Rev. Mod. Phys. 89, 035002 (2017).
- [68] N. Mizuochi, P. Neumann, F. Rempp, J. Beck, V. Jacques, P. Siyushev, K. Nakamura, D.J. Twitchen, H. Watanabe, S. Yamasaki, F. Jelezko, and J. Wrachtrup, Coherence of single spins coupled to a nuclear spin bath of varying density, Phys. Rev. B 80, 041201(R) (2009).
- [69] S. Sangtawesin, B.L. Dwyer, S. Srinivasan, J. J. Allred, L. V. H. Rodgers, K. DeGreve, A. Stacey, N. Dontschuk, K. M. O'Donnell, D. Hu, D. A. Evans, C. Jaye, D. A. Fischer, M. L. Markham, D. J. Twitchen, H. Park, M. D. Lukin, N. P. deLeon, Origins of diamond surface noise probed by correlating single-spin measurements with surface spectroscopy, Phys. Rev. X 9, 031052 (2019).

- [70] H.-J. Werner, Z. Schulten, and K. Schulten, Theory of the magnetic field modulated geminate recombination of radical ion pairs in polar solvents: Application to the pyrene–N, N-dimethylaniline system, J. Chem. Phys. **67**, 646 (1977).
- [71] C. T. Rodgers, S. A. Norman, K. B. Henbest, C. R. Timmel, and P. J. Hore, Determination of radical re-encounter probability distributions from magnetic field effects on reaction yields, J. Am. Chem. Soc. 129, 6746 (2007).
- [72] H. P. Erickson, Size and shape of protein molecules at the nanometer level determined by sedimentation, gel filtration, and electron microscopy, Biol. Proced. Online 11, 32 (2009).
- [73] D. Kobylkov, J. Wynn, M. Winklhofer, R. Chetverikova, J. Xu, H. Hiscock, P. J. Hore, and H. Mouritsen, Electromagnetic 0.1–100 khz noise does not disrupt orientation in a night-migrating songbird implying a spin coherence lifetime of less than 10 μs, J. R. Soc. Int. 16, 20190716 (2019).
- [74] M. S. Grinolds, S. Hong, P. Maletinsky, L. Luan, M. D. Lukin, R. L. Walsworth, and A. Yacoby, Nanoscale magnetic imaging of a single electron spin under ambient conditions, Nat. Phys. 9, 215 (2013).
- [75] B. Brocklehurst, Spin correlation in the geminate recombination of radical ions in hydrocarbons. Part 1.-Theory of the magnetic field effect, J. Chem. Soc. Faraday 72, 1869 (1976).
- [76] B. Brocklehurst and K. A. McLauchlan, Free radical mechanism for the effects of environmental electromagnetic fields on biological systems, Inter. J. Rad. Biology **69**, 3 (1996).
- [77] H. G. Hiscock, H. Mouritsen, D. E. Manolopoulos, and P. J. Hore, Disruption of magnetic compass orientation in migratory birds by radiofrequency electromagnetic fields, Biophys. J. 113, 1475 (2017).
- [78] G. de Lange, T. V. D. Sar, M. Blok, Z.-H. Wang, V. Dobrovitski, and R. Hanson, Controlling the quantum dynamics of a mesoscopic spin bath in diamond, Sci. Rep. 2, 382 (2012).

- [79] E. Bauch, C. A. Hart, J. M. Schloss, M. J. Turner, J. F. Barry, P. Kehayias, S. Singh, and R. L. Walsworth, Ultralong dephasing times in solid-state spin ensembles via quantum control, Phys. Rev. X 8, 031025 (2018).
- [80] M. Steiner, P. Neumann, J. Beck, F. Jelezko, and J. Wrachtrup, Universal enhancement of the optical readout fidelity of single electron spins at nitrogen-vacancy centers in diamond, Phys. Rev. B 81, 035205 (2010).
- [81] A. M. Lewis, T. P. Fay, and D. E. Manolopoulos, An efficient quantum mechanical method for radical pair recombination reactions, J. Chem. Phys. 145, 244101 (2016).
- [82] T. P. Fay, A. M. Lewis, and D. E. Manolopoulos, Spindependent charge recombination along para-phenylene molecular wires, J. Chem. Phys. 147, 064107 (2017).
- [83] I. Lovchinsky, A. O. Sushkov, E. Urbach, N. P. de Leon, S. Choi, K. D. Greve, R. Evans, R. Gertner, E. Bersin, C. Müller *et al.*, Nuclear magnetic resonance detection and spectroscopy of single proteins using quantum logic, Science 351, 836 (2016).
- [84] A. O. Sushkov, N. Chisholm, I. Lovchinsky, M. Kubo, P. K. Lo, S. D. Bennett, D. Hunger, A. Akimov, R. L. Walsworth, H. Park *et al.* All-optical sensing of a single-molecule electron spin, Nano Lett. 14, 6443 (2014).
- [85] P. Wang, S. Chen, M. Guo, S. Peng, M. Wang, M. Chen, W. Ma, R. Zhang, J. Su, X. Rong *et al.* Nanoscale magnetic imaging of ferritins in a single cell, Science advances 5, eaau8038 (2019).
- [86] C. R. Timmel, U. Till, B. Brocklehurst, K. A. Mclauchlan, and P. J. Hore, Effects of weak magnetic fields on free radical recombination reactions, Mol. Phys. 95, 71 (1998).
- [87] D. A. Hopper, H. J. Shulevitz, and L. C. Bassett, Spin readout techniques of the nitrogen-vacancy center in diamond, Micromachines 9, 437 (2018).