

## Nontopological Saddle-Splay and Curvature Instabilities from Anisotropic Membrane Inclusions

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Anisotropic inclusions are shown to induce spontaneous deviatoric bendings in lipidic membranes, by orienting at right angles across the bilayer. In the limit of strong membrane curvatures, a nonanalytical bending energy term is generated that favors saddlelike and cylindrical shapes, without penalizing spherical ones. An “egg-carton” instability results in flat membranes as well as a wormlike shape instability for vesicles. [S0031-9007(96)00321-3]

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Fluid lipid bilayers are model systems of biological membranes [1]. Pure surfactant bilayers are rather well understood in terms of local hydrophilic and hydrophobic interactions, stress profiles, and geometrical considerations [1–3]. Recent studies focus on *heterogeneities* as means of controlling membrane elastic moduli. The effect of cosurfactants [4,5] and anchored polymers [6–8], the behavior of two component membranes [9,10] and polymerized bilayers [11–13] have been theoretically and experimentally studied. Besides, models involving “hat” and “saddle” defects [14] or antagonistic membrane components [10] have been proposed to tentatively explain an apparent submicroscopic roughness in some lipid bilayers [15]. Isotropic membrane inclusions can yield spontaneous curvature effects [16] and corrugations [17], however, only when present with different densities on both sides of the bilayer. In this Letter, we consider the effect of *anisotropic* membrane inclusions. Possible candidates are the dimeric surfactants called “gemini” [18,19] whose polar heads are linked by a spacer. We show that their orientation should couple with the membrane principal curvatures, reducing the bending rigidity  $\kappa$  and increasing  $\bar{\kappa}$ . Stronger effects might be expected from dimers of detergents with large polar heads, or from asymmetric integral proteins [20]. In the limit of large curvatures, we show that anisotropic inclusions yield a *nonanalytical* saddle-splay contribution proportional to the modulus of the deviatoric bending. Above an inclusion concentration threshold, this term induces an “egg-carton” instability in flat membranes and a vesicle instability yielding long wormlike shapes.

Let us consider a fluid lipidic bilayer containing a density  $n$  of anisotropic inclusions. For the sake of simplicity, we assume that  $n$  is constant throughout the membrane and that the inclusions are equally distributed on both sides of the bilayer. In this case, isotropic inclusions would yield no effect by exact compensation [17]. We consider a smoothly deformed membrane given by the equation  $u(x, y)$  of its midsurface. The orientation of a single inclusion can be described by a unit vector  $\vec{a}$  lying in the plane  $(x, y)$ . Its energy depends on the

local curvature of the bilayer. To lowest order in the curvature tensor  $u_{ij} = \partial^2 u / \partial x_i \partial x_j$ , the Hamiltonian of the inclusion can be written as

$$h(u_{ij}; a_k) = h_0 + T_{ij}(a_k)u_{ij} + \mathcal{O}(u_{ij}^2), \quad (1)$$

with summation over repeated indices implied. This equation describes the inclusion contribution to the spontaneous curvature of the membrane. The tensor  $T_{ij}$  can be decomposed in a term proportional to the identity tensor  $\delta_{ij}$  and a term proportional to  $a_i a_j$ . Calling  $c_1$  and  $c_2$  the eigenvalues of  $u_{ij}$  (i.e., the principal curvatures) and calling  $\vartheta$  the orientation of  $\vec{a}$  with respect to, say, the axis of curvature  $c_1$ , we are led to

$$h(c_1, c_2, \vartheta) = h_0 + \alpha(c_1 + c_2) + \omega(c_1 - c_2) \cos 2\vartheta, \quad (2)$$

in which we can choose  $\omega > 0$  without loss of generality. Hence, anisotropic inclusions behave as quadrupoles in the membrane curvature field. They will be disoriented by thermal fluctuations when  $|c_1 - c_2| \lesssim c^*(T)$ , with

$$c^*(T) \sim \frac{T}{\omega}, \quad (3)$$

otherwise, they will tend to orient along the most favorable axis of principal curvature. Let us estimate  $c^*$  for various types of inclusions. For small geminated surfactants of size comparable to the monolayer thickness, we expect  $1/c^*$  to be a molecular size. Larger inclusions might yield larger effects. Let us imagine for instance an integral protein with a conical section of angle  $+\theta_0$  in the  $x$ - $z$  plane and angle  $-\theta_0$  in the  $y$ - $z$  plane. The lipids are tilted around it over some relaxation length  $\xi$ . This tilt and the corresponding energy can be relaxed by curving the membrane. To estimate the corresponding  $\omega$ , we shall consider curving the membrane cylindrically in the  $x$ - $z$  plane (Fig. 1). Calling  $\vec{m}$  the projection of the lipid orientation onto the membrane plane, the distortion energy density reads [21]

$$f_d = \frac{1}{2} B_{\perp} \vec{m}^2 + \frac{1}{2} K (\nabla_{\perp} \cdot \vec{m} - c)^2. \quad (4)$$

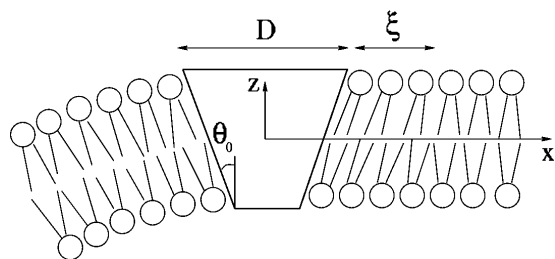


FIG. 1. Lipid tilt distortion around a conical inclusion in a flat (right) or curved (left) membrane.

The second term arises from the splay of the lipids, which is coupled with the membrane bending elasticity. The solution of the Euler-Lagrange equation associated with (4) is  $m(s) = \theta \exp(-s/\xi)$ , with  $s$  the curvilinear coordinate along the surface,  $\theta$  the boundary tilt, and  $\xi = \sqrt{K/B_{\perp}}$ . The resulting energy is  $W = \frac{1}{2}\kappa\xi^{-1}\theta^2 - \kappa\theta c$  per unit length. With the boundary tilt given by  $\theta(c) = \theta_0 - \frac{1}{2}Dc$  (the inclusion would fit perfectly in the membrane for  $c = 2\theta_0/D$ ), we obtain  $\omega = -D(\partial W/\partial c)|_{c=0}$ , yielding after dividing by  $T$

$$1/c^* = \frac{K}{T} \theta_0 D \left(1 + \frac{D}{2\xi}\right). \quad (5)$$

With  $K \sim \kappa \sim 20T$  for a phospholipid membrane,  $D \sim 60 \text{ \AA}$ ,  $\xi \sim 30 \text{ \AA}$  (a few lipid head sizes), and  $\theta_0 \sim 45^\circ$ , we obtain  $1/c^* \sim 2000 \text{ \AA}$ . For large head detergent dimers, we might expect  $1/c^* \sim 300 \text{ \AA}$ .

According to statistical mechanics, the free energy of a single inclusion derived from (2) is

$$\mu(c_1, c_2) = h_0 + \alpha(c_1 + c_2) - T \ln \int_0^{2\pi} \frac{d\vartheta}{\vartheta_0} e^{-(\omega/T)(c_1 - c_2) \cos 2\vartheta}, \quad (6)$$

where  $\vartheta_0$  is an irrelevant angle quantum. The integral in (6) is a Bessel function  $2\pi\vartheta_0^{-1}I_0[\omega(c_1 - c_2)/T]$ . The total contribution of the inclusions to the membrane energy,  $g(c_1, c_2)$ , is given by  $\frac{n}{2}\mu(c_1, c_2) + \frac{n}{2}\mu(-c_1, -c_2)$ , since inclusions in opposite sides of the bilayer experience opposite curvatures. We obtain

$$g(c_1, c_2) = -nT \ln I_0 \left[ \frac{c_1 - c_2}{c^*(T)} \right], \quad (7)$$

in which we eliminated a constant contribution, namely,  $nh_0 - nT \ln(2\pi/\vartheta_0)$ , that includes the rotational entropy of the inclusions. As expected in a symmetric bilayer, the term proportional to  $c_1 + c_2$  disappears; however, the contribution resulting from the anisotropy remains. A graph of  $g$  is shown in Fig. 2. The membrane bending energy is obtained by adding the contribution  $g(c_1, c_2)$  to the Helfrich energy [22]

$$f(c_1, c_2) = \frac{1}{2} \kappa (c_1 + c_2)^2 + \bar{\kappa} c_1 c_2, \quad (8)$$

where  $\kappa$  and  $\bar{\kappa}$  are the bending modulus and the Gaussian modulus, respectively. We consider two limits.

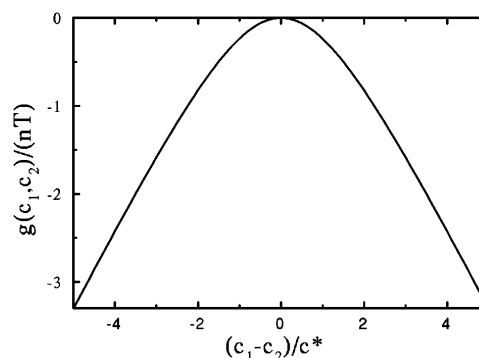


FIG. 2. Contribution of anisotropic inclusions to the bending energy of a membrane. For  $|c_1 - c_2| \geq c^*$  the energy gain is almost linear.

(a)  $|c_1 - c_2| \leq c^*$ .—If the deviatoric bending is smaller than the threshold  $c^*$ , the inclusions contribution simply renormalizes the membrane bending moduli. From  $g(c_1, c_2) \sim -\frac{1}{4}nT^{-1}\omega^2(c_1 - c_2)^2 + \mathcal{O}(4)$ , we derive

$$\Delta\kappa = -\frac{n\omega^2}{2T}, \quad (9a)$$

$$\Delta\bar{\kappa} = \frac{n\omega^2}{T}. \quad (9b)$$

Hence, the bending modulus  $\kappa$  is reduced, as is usually the case for cosurfactants [1]. Furthermore, anisotropic inclusions favor *negative* Gaussian curvature (i.e., saddle shapes), as will be made clearer in the following.

(b)  $|c_1 - c_2| \geq c^*$ .—If the deviatoric bending is larger than  $c^*$  [23], which is possible for large inclusions,  $g$  becomes approximately

$$g(c_1, c_2) \sim nA - n\omega|c_1 - c_2|, \quad (10)$$

where  $A = \frac{1}{2}T \ln[2\pi|c_1 - c_2|/c^*]$  can be treated as a constant thanks to its weak logarithmic dependence. This linear behavior can be easily understood from a mean field picture. According to Eq. (2), the orientation of minimum energy for a single inclusion corresponds to  $\vartheta = 0$  if  $c_1 - c_2 < 0$  and to  $\vartheta = \pi/2$  if  $c_1 - c_2 > 0$ . In both cases, the anisotropic energy contribution is exactly  $\omega|c_1 - c_2|$ . Since  $c_1 - c_2$  changes sign from one monolayer of the membrane to the other, anisotropic inclusions will orient at right angles when the membrane is shaped as a saddle. This corresponds to the most favorable situation (Fig. 3). Conversely, if the membrane is shaped as a sphere, there is exact compensation on the two sides of the bilayer. Thus, in contrast with the usual Gaussian curvature term  $\bar{\kappa}c_1c_2$ , the presence of anisotropic inclusions provides an energy gain within the saddles that is *not* compensated by a corresponding energy loss in surrounding hats. In other words, the Gauss-Bonnet theorem does not apply to

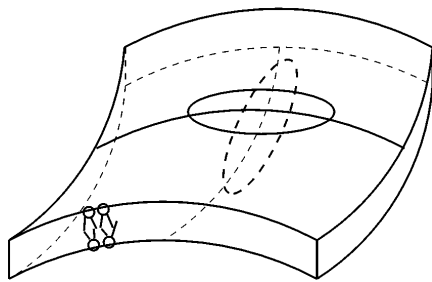


FIG. 3. Anisotropic inclusions (arbitrary scale) in a membrane with deviatoric bending. The inclusions depicted are supposed to “prefer,” besides a mean curvature of whatever sign, an excess of curvature around their short axis and a default of curvature around their long axis. By orienting at right angles across the bilayer, both inclusions can satisfy this differential preference. An additional mean curvature does not change the tendency of the inclusions to align.

Eq. (10). This property was first understood by Fischer [24,25], who postulated by analogy with the spontaneous mean curvature a term proportional to  $|c_1 - c_2|$  for pure membranes, but estimated it to be negligible.

*Egg-carton instability.*—The saddle splay favored by anisotropic inclusions yields an instability. Let us compare the energy of a corrugated membrane,

$$u(x, y) = U \sin qx \sin qy, \quad (11)$$

with that of a flat one. Since the integral over the membrane of the Gaussian term  $\bar{\kappa}c_1c_2$  is a topological invariant, the instability must arise from the nonanalytical form (10). From the standard relations  $c_1 + c_2 = \text{Tr}u_{ij}$  and  $c_1c_2 = \text{Det}u_{ij}$ , we infer  $c_1 + c_2 = u_{xx} + u_{yy}$  and  $|c_1 - c_2| = [(u_{xx} - u_{yy})^2 + 4u_{xy}^2]^{1/2}$ . Hence, for the egg carton (11),  $c_1 + c_2 = -2q^2U \sin qx \sin qy$  and  $|c_1 - c_2| = 2q^2U |\cos qx \cos qy|$ . Balancing the inclusion gain (10) in the saddles, of order  $n\omega q^2U$ , with the average bending cost  $E_b = \frac{1}{2}\kappa(c_1 + c_2)^2$  of order  $\kappa q^4U^2$ , we arrive at the instability criterion

$$\frac{n}{q^2U} \geq \frac{\kappa}{\omega}. \quad (12)$$

Since the smallest acceptable value of  $q^2U$  that allows us to use the linear form (10) is precisely  $c^*$ , the egg-carton instability occurs when the inclusion density overcomes  $\kappa c^*/\omega$ , i.e., above the threshold

$$n_c \sim \frac{\kappa}{T} c^{*2}. \quad (13)$$

The curvature modulation is then of order  $c^*$ ; however, a more complete calculation would be required to determine the corrugation's amplitude and wave vector. The inclusion coverage at threshold,  $d^2n_c \sim (T/\kappa)\theta_0^{-2}[1 + D/(2\xi)]^{-2}$  according to our estimation of  $c^*$  for conical objects, can be very small for large inclusions with  $\theta_0 \sim 1$ .

*Wormlike vesicle instability.*—The energy term (10) favors not only saddles but also *cylinders*, for which

$|c_1 - c_2|$  is nonzero as well. This might cause an instability in the shape of vesicles, such as that recently reported in the presence of geraniol, a biological cosurfactant [26]. Let us compare the energy of a spherical vesicle  $S$ , of radius  $R$ , with that of a cylindrical one  $C$ , of radius  $r$ , and length  $L$ , terminated by spherical end caps. The constraint of constant membrane area implies  $2\pi rL = 4\pi(R^2 - r^2)$ . The two vesicles possess the same Gaussian energy and furthermore the bending energy  $8\pi\kappa$  of  $S$  equals that of the end caps of  $C$ . Thus the energy excess  $\Delta E$  of  $C$  with respect to  $S$  consists only in the energy of the cylindrical region of  $C$ , namely,

$$\Delta E(r) = 4\pi(R^2 - r^2) \left[ \frac{\kappa}{2r^2} - nT \ln I_0 \left( \frac{1}{rc^*} \right) \right]. \quad (14)$$

Typical graphs of  $\Delta E(r)$  are shown in Fig. 4. The instability condition is  $\Delta E < 0$  for  $r \leq R$ . In the limit of large vesicles  $Rc^* \gg 1$ , expanding the Bessel function in power series yields  $\Delta E \sim \pi(R^2 - r^2)[2\kappa - nT/c^{*2}](1/r^2)$ . We thus obtain the same threshold as previously

$$n_c = 2 \frac{\kappa}{T} c^{*2}. \quad (15)$$

For  $n \leq n_c$ , vesicles should exhibit strong shape fluctuations. As soon as  $n \geq n_c$ , they turn to cylinders with a small equilibrium radius  $r_e$  of order  $1/c^*$  (Fig. 4). To estimate it, we can set  $r_e \ll R$  and use the asymptotic form (10) for the inclusions contribution. This gives  $\Delta E \sim 4\pi R^2 [\frac{1}{2}\kappa/r^2 - nTc^{*-1}/r]$ . Hence  $r_e \sim \kappa c^*/nT$ , i.e.,

$$\frac{1}{r_e} \sim 2 \frac{n}{n_c} c^*. \quad (16)$$

This value is independent of  $R$  in the large vesicle limit. Long wormlike vesicles (Fig. 5) should behave as flexible polymers [26] and could undergo a “pearling” instability. Preliminary calculations show that such an instability is possible although not as favorable as the egg-carton one in flat membranes. It would also be interesting to

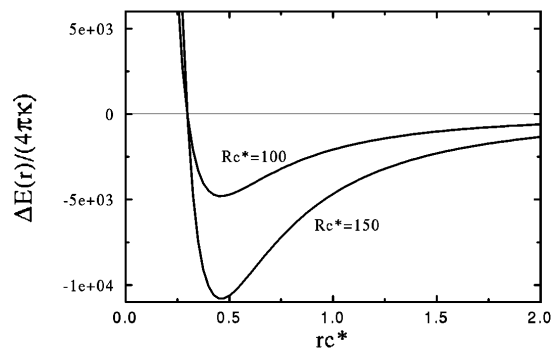


FIG. 4. Energy excess of cylindrical vesicles of radius  $r$  with respect to spherical ones of radius  $R$ , for  $n \sim 1.5n_c$ . The minimum is obtained for  $r_e \sim 1/c^*$ , independently of the vesicle total area.

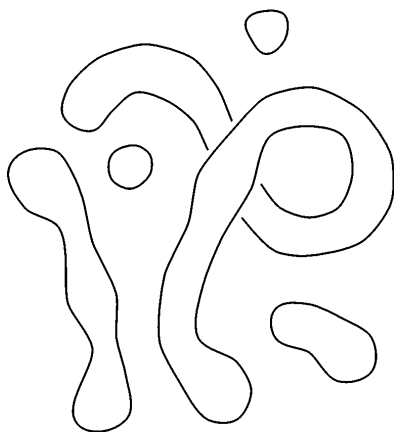


FIG. 5. Wormlike vesicles induced by anisotropic inclusions.

study whether an instability yielding *spikes* on the vesicle surface (like the blood cell echinocytosis) would not compete with the transformation to wormlike vesicles.

In summary, inclusions with an *anisotropic* spontaneous curvature could play an important role in the morphology of membranes. Whereas isotropic inclusions yield no spontaneous mean curvature when symmetrically absorbed in a bilayer, anisotropic inclusions yield a spontaneous deviatoric bending  $c_1 - c_2 \neq 0$  by orienting at right angles across the bilayer. This favors saddlelike or cylindrical membrane shapes. The effect of small anisotropic inclusions is mainly to reduce the bending modulus  $\kappa$  and to increase the Gaussian modulus  $\bar{\kappa}$ . Large inclusions, such as big dimeric detergents or proteins, yield in the limit of strong mesoscopic curvatures an energy term proportional to the *modulus* of the deviatoric bending. Such a nonanalytical term favors saddles without penalizing spheres, contrary to the usual Gaussian curvature term. We showed that above an inclusion concentration threshold  $n_c$  (as small as the inclusions are large), flat membranes undergo an egg-carton instability with a mesoscopic roughness. This might be an alternative to the “hat model” to explain the roughness of real membranes [14]. Above  $n_c$ , spherical vesicles should transform to *wormlike* vesicles with a diameter also probably in the mesoscopic range. This might explain the recently observed “entangled tubular vesicle phase” [26], obtained in the presence of geraniol, a branched co-surfactant that could possibly yield anisotropic associations with the membrane lipids [27]. Extension of the present model should include inhomogeneities in the distribution of the inclusions and the investigation of a spiked vesicle regime.

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- [1] J.N. Israelachvili, *Intermolecular and Surface Forces* (Academic Press, London, 1992).
- [2] A. G. Petrov and J. Bivas, *Prog. Surf. Sci.* **18**, 389 (1984).
- [3] W. Helfrich, in *Physics of Defects*, Proceedings of the Les Houches Summer School, Session XXVR, edited by R. Balian *et al.* (North-Holland, Amsterdam, 1981), p. 716.
- [4] P. G. de Gennes and C. Taupin, *J. Phys. Chem.* **86**, 2294 (1982).
- [5] M. M. Kozlov and W. Helfrich, *Langmuir* **8**, 2792 (1992).
- [6] G. Blume and G. Cevc, *Biochim. Biophys. Acta* **1029**, 91 (1990).
- [7] C. R. Safinya *et al.* (to be published).
- [8] R. Lipowsky, *Europhys. Lett.* **30**, 197 (1995).
- [9] S. A. Safran, P. Pincus, and D. Andelman, *Science* **248**, 354 (1990).
- [10] M. M. Kozlov and W. Helfrich, *J. Phys. II (France)* **4**, 1427 (1994).
- [11] E. Sackmann, P. Eggl, C. Fahn, H. Bader, H. Ringsdorf, and M. Schollmeier, *Ber. Bunsen-Ges. Phys. Chem.* **89**, 1198 (1985).
- [12] M. M. Kozlov and W. Helfrich, *Langmuir* **10**, 4219 (1994).
- [13] M. M. Kozlov and W. Helfrich, *Phys. Rev. E* **51**, 3324 (1995).
- [14] W. Helfrich, *Liq. Cryst.* **5**, 1647 (1989).
- [15] W. Helfrich and M. Mutz, *Fluctuations and Pattern Growth: Experiments and Theory*, edited by H. E. Stanley and N. Ostrowsky (Kluwer, Dordrecht, The Netherlands, 1988), p. 222.
- [16] H. Gruler, *Z. Naturforsch.* **30c**, 608 (1975).
- [17] S. Leibler, *J. Phys. (Paris)* **47**, 507 (1986).
- [18] F. M. Menger and C. A. Littau, *J. Am. Chem. Soc.* **113**, 1451 (1991).
- [19] R. Zana and Y. Talmon, *Nature (London)* **362**, 228 (1993).
- [20] H. Lodish, D. Baltimore, A. Berk, S. L. Zipursky, P. Matsudaira, and J. Darnel, *Molecular Cell Biology* (Scientific American Books, New York, 1995).
- [21] T. C. Lubensky and F. C. MacKintosh, *Phys. Rev. Lett.* **71**, 1565 (1993).
- [22] W. Helfrich, *Z. Naturforsch.* **28c**, 693 (1973).
- [23] But still smaller than a microscopic spontaneous curvature  $c_s$ , above which second order terms (2) need to be considered.
- [24] T. M. Fischer, *J. Phys. II (France)* **2**, 337 (1992).
- [25] T. M. Fischer, *J. Phys. II (France)* **3**, 1795 (1993).
- [26] S. Chiruvolu, H. E. Warriner, E. Naranjo, S. H. J. Idziak, J. O. Rädler, R. J. Plano, J. A. Zasadzinski, and C. R. Safinya, *Science* **266**, 1222 (1994).
- [27] C. R. Safinya (private communication).