Confinement induced three-dimensional trajectories of microswimmers in rectangular channels

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We study the trajectories of a model microorganism inside three-dimensional channels with square and rectangular cross sections. Using (1) numerical simulations based on the lattice-Boltzmann method and (2) analytical expressions using far-field hydrodynamic approximations and the method of images we systematically investigate the role of the strength and finite-size of the squirmer, confinement dimensions, and initial conditions in determining the three-dimensional trajectories of microswimmers. Our results indicate that the hydrodynamic interactions with the confining walls of the channel significantly affect the swimming speed and trajectory of the model microswimmer. Specifically, pullers always display sliding motion inside the channel: weak pullers slide through the channel center line, while strong pullers slide through a path close to any of the walls. Pushers generally follow helical motion in a square channel. Unlike pullers and pushers, the trajectories of neutral swimmers are not easy to generalize and are sensitive to the initial conditions. Despite this diversity in the trajectories, the far-field expressions capture the essential features of channel-confined swimmers. Finally, we propose a method based on the principle of superposition to understand the origin of the three-dimensional trajectories of channel confined swimmers. Such construction allows us to predict and justify the origin of apparently complex three-dimensional trajectories generated by different types of swimmers in channels with square and rectangular cross sections.

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I. INTRODUCTION

Microorganisms swim in complex natural and biological environments overwhelmingly constituted by confined spaces [1,2]. A few examples are sperm cells swimming inside the female reproductive tract [3,4], and bacteria swimming in the digestive gut and in pores in the extracellular matrix [5,6]. It is interesting to note that the swimming dynamics of microorganisms is determined by the stimuli or cues that they receive from the local environment, which may emanate from confining surfaces in immediate proximity and from other active or passive entities in the vicinity [7–9]. In addition to the steric [10], chemical [11], electrical [12,13], and thermal [14] interactions between a microswimmer and a confining surface, hydrodynamic interactions with the confining walls can play a dominant role in dictating the microswimmer dynamics.

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In one of the earliest experiments, it was shown that wall-induced hydrodynamic interaction led to the preferential accumulation of bull sperm cells near the confining walls [15]. The hydrodynamic interaction of a microswimmer that swims using flagella or a flagellum at the posterior, known as pushers (e.g., sperm cells and flagellated bacteria), with a rigid wall results in reorientation of the swimmer parallel to the wall and in a simultaneous attraction towards the closest wall [15,16]. Subsequently, several studies focused on this hydrodynamic origin of the accumulation of swimming cells near a confining wall using both sperm cells [17,18] and bacterial cells [19,20]. Interestingly, it was also shown that the swimming trajectory of a pusher-type microswimmer turns from straight, with intermittent tumbling, in the bulk to circular close to a wall due to the involved hydrodynamic interactions [16,21]. The directionality of the circular trajectory, i.e., clockwise or anticlockwise, depends on whether the confining surface is a rigid wall or an air-water interface [16,21,22]. Furthermore, microswimmers exhibit other surprising behaviors like orbiting/sliding, hovering, scattering, bouncing and oscillation near solid boundaries, specifically curved ones, and in confinements due to the involved hydrodynamic interactions with walls [23–27]. Instead of smooth and rigid walls, hydrodynamic interactions of microswimmers were also studied near corrugated periodic surfaces [28,29], elastic boundaries [30,31], and in thin films [32]. Further, the changes in the trajectories of microswimmers in confinements were studied by varying the channel geometry [29,33] and the channel size [34–36], and by including the higher-order hydrodynamic moments describing the microswimmer [37]. Most of these studies indicate two things: (1) confinement enhances swimming velocity and (2) oscillatory or helical trajectories are commonly observed when the microswimmers are confined in narrow channels or capillary tubes. A detailed numerical study [34] of the trajectories of a single spherical squirmer in a circular tube revealed that in the cylindrical confinement neutral microswimmers adopt helical trajectories, while weak and strong pullers exhibit sliding motion through the center of the tube and sliding near the wall respectively. Pusher-type swimmers end up crashing against the walls [34]. Helical motion of rodlike swimmers, both pushers and pullers, in a circular tube and in a channel of square cross section were also briefly reported in [37]. Despite the importance of hydrodynamic interaction with the confining walls, studies detailing the role of geometry of the channel, in particular the role of anisotropy of the channel cross section, on the dynamics of microswimmers remain scarce.

Recently, understanding the motility and collective behavior of biological microswimmers like bacteria in a microfluidic environment has gained importance for its significance in ecology, biological processes, diagnostics, and biocomputation [38]. Additionally, synthetic microswimmers that are engineered to mimic the behavior of their biological counterparts also find applications in microfluidic devices and confinements [39]. Although microchannels with rectangular cross sections are most common in all microfluidic applications, the role of the anisotropic channel cross section on microswimmer dynamics and the resulting three-dimensional (3D) complex trajectories of the microswimmers remain largely unaddressed in the literature. The present work aims to address this lack of understanding by analyzing the trajectories of squirmers of varying strength in a range of microchannels of rectangular cross section.

We use a spherical squirmer model for the microswimmer which significantly reduces the complexity associated with the shape and the swimming strokes of the microswimmer. We also restrict our analysis to flat-walled microchannels in order to avoid the complexity of microswimmer dynamics near curved surfaces [25,27]. We study the dynamics of squirmers in these rectangular microchannels using numerical simulations which employ the lattice-Boltzmann method (LBM). Further, we develop an approximate analytical approach for calculating the 3D trajectories of the squirmers based on the far-field hydrodynamic approximation [37,40–43] of the microswimmer flow field along with method of images [42,44]. The results from this analytical approach are compared with those from full numerical simulations in order to test the efficacy of the former. Our results illustrate the consequences of a rectangular confinement on the trajectory of a microswimmer in a microchannel. Importantly, our analyses also help in disentangling the complexity of the 3D trajectories of channel confined swimmers by constructing them as superposition of trajectories from simpler configurations.

The rest of the paper is structured as follows: in Sec. II the squirmer model and the geometry of the channel are described. Section III is dedicated to explaining the solution methodology wherein we discuss the method of far-field approximation along with the method of images and the LBM. We then present the results in Sec. IV by first finding the instantaneous swimming velocity of the squirmer and then validating the accuracy of the far-field approximation. Then, in Sec. IV B, we move on to explore the two-dimensional (2D) trajectories of squirmers in the square channels. We will show that swimmers either swim parallel to the channel axis or perform a wavelike motion depending upon the strength of the force dipole. The results are compared with far-field approximations wherever possible. The analysis is then generalized for the case of 3D motion in both square (Sec. IV C) and rectangular (Sec. IV D) channels, and the origin of the complexity of the trajectories is explained using a simple superposition method.

II. MATHEMATICAL MODEL

A. Squirmer model

In this study, we use the squirmer model to simulate microswimmers [45–47]. We consider an axisymmetric, nondeformable, spherical squirmer [47] with a tangential surface slip velocity $U_s(\phi)$ given by

$$U_s(\phi) = \sum_{n=1}^{\infty} B_n \frac{2}{n(n+1)} \sin(\phi) P'_n(\cos\phi), \tag{1}$$

where B_n is the strength of the *n*th mode of tangential slip velocity, and P'_n is the associated Legendre polynomial. The polar angle between the position vector on the surface of the squirmer \hat{r} and the swimming direction \hat{e} is represented as $\phi = \cos^{-1}(\hat{e} \cdot \hat{r}/a)$. The swimming speed of the squirmer in an unconfined environment, $\hat{U}_f = (2/3)B_1\hat{e}$, is dependent on the first swimming mode alone.

In this work, we have $B_n = 0$, for n > 2. The relative strengths of the first two modes, $\beta = B_2/B_1$, determine the hydrodynamic nature of the swimming motion. On one hand, the squirmers with $\beta < 0$ are called pushers (e.g., *Escherichia coli*). They swim by pushing the fluid at the front and back, and pulling the fluid from the sides. On the other hand, pullers, $\beta > 0$ (e.g., *Chlamydomonas*), swim by pulling the fluid from front and back. The squirming motion with $\beta = 0$ corresponds to neutral swimmers (e.g., *Paramecium*). These microswimmers move the fluid from front to back by generating flow field corresponding to a source dipole [47].

B. Swimming through a rectangular channel

We consider the spherical squirmer of radius a swimming in the rectangular channel as illustrated in Fig. 1. The x axis is along the channel center line, and the y and z axes are considered along the height and width of the channel respectively. From the channel center line (x axis), the squirmer is located at a distance b in the y direction and at a distance c in the z direction. The dimensionless positions x^* , y^* , and z^* are defined as

$$x^* = x/a, (2)$$

$$y^* = b/(H - a),\tag{3}$$

$$z^* = c/(W - a). \tag{4}$$

In other words, $y^* = 0$ represents a plane passing through the channel center and perpendicular to the y axis. $y^* = \pm 1$ represents the location of the squirmer when it touches the walls perpendicular to the y axis (henceforth referred to as top and bottom walls). Similarly, $z^* = 0$ is the plane passing through the channel center and perpendicular to the z axis. $z^* = \pm 1$ represents the location of the

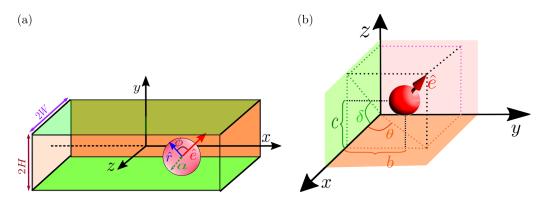


FIG. 1. (a) Schematic representation of a spherical squirmer of radius a swimming in a rectangular channel. The unit vector \hat{e} represents the orientation of the squirmer, and \hat{r} is a position vector that describes the surface of the squirmer with respect to its center. The channel is of height 2H and width 2W. The origin of the Cartesian coordinates coincides with the center of the channel. The x direction is periodic, and it aligns with the channel center line, which is shown as the dashed line. The y and z axes are oriented along the height and width of the channel respectively. (b) Symbols used to describe the location and orientation of the squirmer. From the channel center line, the squirmer is located at a distance b in the y direction and c in the c direction. The angles c0 and c0 describe the orientation of the microswimmer and are measured from c1 axis to the projection of the orientation vector c2 on c3 and c4 planes respectively.

squirmer when it touches the walls perpendicular to z axis (henceforth referred to as left and right walls).

The strength or the extent of the confinement is quantified by the ratio of the squirmer radius to the half channel height, a/H. The aspect ratio of the channel cross section, AR, is defined as

$$AR = (W - H)/W \tag{5}$$

such that AR = 0 represents a square channel. The rectangular channels have $AR \neq 0$. $AR \rightarrow 1$ represents a rectangular channel with a very large width.

C. Governing equations for the squirmer velocity field

The squirmer is suspended in an incompressible, Newtonian fluid, and the velocity field generated by the squirmer is described by the Navier-Stokes equations,

$$\nabla \cdot \hat{\boldsymbol{u}} = 0, \tag{6}$$

$$\rho \left(\frac{\partial \hat{\boldsymbol{u}}}{\partial t} + \hat{\boldsymbol{u}} \cdot \nabla \hat{\boldsymbol{u}} \right) = -\nabla p + \mu \nabla^2 \hat{\boldsymbol{u}}, \tag{7}$$

where \hat{u} is the velocity field, p is the pressure, and μ and ρ are the viscosity and density of the surrounding liquid respectively. Note that the Reynolds number, which gives a relative measure of the inertial effects to the viscous effects, associated with the microswimmer dynamics in the viscous liquid is negligibly small. In that limit, Eq. (7) reduces to the Stokes equation

$$\nabla p = \mu \nabla^2 \hat{\boldsymbol{u}} \tag{8}$$

The periodic boundary condition is applied at the two open boundaries (along the *x* axis) of the rectangular channel, and the no-slip boundary condition is specified at the four confining walls.

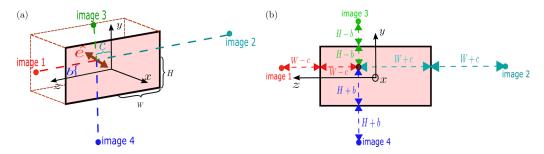


FIG. 2. (a) Schematic representation of the hydrodynamic image systems in a rectangular channel having a half height, H, and half width, W. A combination of force dipole of strength α and source dipole of strength q, oriented along $\hat{\mathbf{e}}$, is placed at (y, z) = (b, c). To the leading order, the hydrodynamic interaction of the microswimmer with each wall is approximated by a corresponding image system for this combination of force and source dipoles. The positions of the four image systems corresponding to the four walls are shown in the figure. (b) The y-z plane of the channel and the distances of the squirmer and the image systems from the confining walls of the channel are shown here.

III. SOLUTION METHODOLOGY

We use two different approaches to determine the velocities and trajectories of the microswimmer. The first method is an approximate analytical approach, where we use the far-field hydrodynamic approximation and the method of images to account for the hydrodynamic interactions between the microswimmer and all the channel walls. In the second method, which is a numerical method, we perform the full-scale simulation of the squirmer in a rectangular channel using LBM. These approaches are discussed below.

A. Method 1: Far-field hydrodynamic approximation

In the multipole description of a force-free and torque-free microswimmer, the flow field generated by the squirmer is that of a stresslet in the leading order [42]. The stresslet corresponds to two point forces acting at a sufficiently small separation distance. The point forces and their separation vector are aligned in the same direction. The strength of the stresslet, α , is related to the second squirming mode as [48],

$$\alpha = -B_2(4\pi\,\mu a^2). \tag{9}$$

The sign of the stresslet strength determines whether the squirmer is a puller ($\alpha < 0$) or a pusher ($\alpha > 0$). Of course, $\alpha = 0$ corresponds to a neutral swimmer. It is now known that approximating the squirmer solely by a force dipole, or a stresslet, is insufficient to address the wall-confined dynamics for a finite-sized microswimmer especially in a confinement [37,49–51]. Hence, in order to consider the effects of the finite size of the spherical microswimmer inside a rectangular channel, we approximate the squirmer as a superposition of force and source dipoles. The strength of the source dipole q is related to the first squirming mode as [48]

$$q = B_1(8\pi \mu a^3/3). \tag{10}$$

The aforementioned description of a squirmer works well when it is isolated, and located far from the confining boundaries of the channel. In the present work, the presence of a neighboring channel wall is taken into account by considering the associated hydrodynamic interactions between the microswimmer and the wall using the method of images [42,44]. We consider a squirmer, represented by the combination of stresslet of strength α and source dipole of strength q, located at (y, z) = (b, c), and oriented along $\hat{\mathbf{e}}$. Figure 2 shows the schematic representing the y-z cross section of the channel, the location of the squirmer under consideration, and the corresponding positions

of the total image systems (i.e., the image systems for stresslet and source dipole singularities) for each of the four confining walls of the rectangular channel.

We first calculate the total velocity field due to the image systems for the squirmer for each pair of walls, i.e., the left and right walls (image systems 1 and 2 in Fig. 2) and the top and bottom walls (image systems 3 and 4 in Fig. 2), following the methodology outlined in [42,43]. Next, we use Faxén's laws [52] to calculate the wall-induced translational and angular velocities of the microswimmer corresponding to the velocity fields generated by the image systems for each pair of walls. Applying Faxén's laws, the induced translational velocity, $\bar{U}_{\text{left/right}}$, and angular velocity, $\bar{\Omega}_{\text{left/right}}$, of the microswimmer due to the presence of the left and right walls can be calculated as [42,43]

$$\bar{\boldsymbol{U}}_{\text{right/left}} = \left\{ \bar{\alpha} \left[\mp \frac{3(1 - 3\cos^{2}(\theta_{z}))}{8(\bar{W} \pm \bar{c})^{2}} \pm \frac{(7 - 11\cos^{2}(\theta_{z}))}{64(\bar{W} \pm \bar{c})^{4}} \right] + \bar{q} \left[\mp \frac{1}{(\bar{W} \pm \bar{c})^{3}} \pm \frac{1}{8(\bar{W} \pm \bar{c})^{5}} \right] \cos(\theta_{z}) \right\} \hat{\boldsymbol{z}} + \left\{ \bar{\alpha} \left[\pm \frac{3}{8(\bar{W} \pm \bar{c})^{2}} \pm \frac{0.078125}{(\bar{W} \pm \bar{c})^{4}} \right] \sin 2(\theta_{z}) + \bar{q} \left[\mp \frac{1}{4(\bar{W} \pm \bar{c})^{3}} \pm \frac{3}{16(\bar{W} \pm \bar{c})^{5}} \right] \sin(\theta_{z}) \right\} \left[\frac{(\hat{\boldsymbol{e}} \cdot (\hat{\mathbf{x}}\hat{\mathbf{x}} + \hat{\mathbf{y}}\hat{\mathbf{y}}))}{|\hat{\boldsymbol{e}} \cdot (\hat{\mathbf{x}}\hat{\mathbf{x}} + \hat{\mathbf{y}}\hat{\mathbf{y}})|} \right], \tag{11}$$

$$\bar{\boldsymbol{\Omega}}_{\text{right/left}} = \left[\mp \frac{3\bar{\alpha}}{16} \frac{\sin 2(\theta_{z})}{(\bar{W} \pm \bar{c})^{3}} \pm \frac{3\bar{q}}{8} \frac{\sin(\theta_{z})}{(\bar{W} \pm \bar{c})^{4}} \right] \left[\frac{\hat{\boldsymbol{e}} \cdot (\hat{\mathbf{x}}\hat{\mathbf{x}} + \hat{\mathbf{y}}\hat{\mathbf{y}})}{|\hat{\boldsymbol{e}} \cdot (\hat{\mathbf{x}}\hat{\mathbf{x}} + \hat{\mathbf{y}}\hat{\mathbf{y}})|} \times (-\hat{\boldsymbol{z}}) \right], \tag{12}$$

where the \bar{U} and $\bar{\Omega}$ represent the dimensionless translational and angular velocities. Here each of the terms with \pm/\mp represent two terms due to the image systems at the right and left walls. Specifically, the first and second signs represent the contributions from the right and left walls respectively. Strengths of force and source dipoles are nondimensionalized using a as the length scale and U_f as the velocity scale to get $\bar{\alpha}$ and \bar{q} respectively. θ_z is defined as $\theta_z = \arccos(\hat{\mathbf{e}} \cdot \hat{\mathbf{z}})$. Note that Eq. (11) includes not only the lowest order terms obtained after applying Faxén's first law to the velocity field for the image systems, but also the higher-order terms due to the Laplacian operator, i.e., terms proportional to $\frac{1}{(\bar{W}\pm\bar{c})^4}$ for the force-dipole image system and terms proportional to $\frac{1}{(\bar{W}\pm\bar{c})^5}$ for the source dipole image system.

The translational and rotational velocities of the microswimmer induced by the top and bottom walls, $\bar{U}_{\text{top/bottom}}$, can be also calculated using equations similar to Eqs. (11) and (12). Specifically, these can be obtained by changing $\hat{\mathbf{y}} \to \hat{\mathbf{z}}$, $\hat{\mathbf{z}} \to \hat{\mathbf{y}}$, $\bar{W} \to \bar{H}$, $\bar{c} \to \bar{b}$, and $\theta_z \to \theta_y$. Finally, the total translational velocity \bar{U}_{channel} and angular velocity $\bar{\Omega}_{\text{channel}}$ of the microswimmer due to the rectangular confinement can be calculated by adding the induced translational and angular velocities due to each wall (image system) as follows:

$$\bar{\boldsymbol{U}}_{\text{channel}} = \hat{\boldsymbol{e}} \cdot (\hat{\boldsymbol{x}}\hat{\boldsymbol{x}} + \hat{\boldsymbol{y}}\hat{\boldsymbol{y}} + \hat{\boldsymbol{z}}\hat{\boldsymbol{z}}) + \sum_{\text{all walls}} \bar{\boldsymbol{U}}_{\text{wall}},$$

$$\bar{\boldsymbol{\Omega}}_{\text{channel}} = \sum_{\text{all walls}} \bar{\boldsymbol{\Omega}}_{\text{wall}},$$
(13)

where the first term in the expression of $\bar{U}_{channel}$ is the self-propulsion velocity of the squirmer. It must be noted that the above formulation neglects the hydrodynamic interactions between the image systems, and the consequent effect on the dynamics of the microswimmer. Ideally, an infinite series of image systems needs to be considered in order to satisfy the boundary conditions at all the walls simultaneously. Therefore, Eq. (13) gives only approximate expressions for calculating the dynamics of a finite-sized microswimmer in the rectangular channel. The efficacy of this

methodology cannot be commented upon *a priori*, and is ascertained here by comparison with a full-scale numerical simulation.

B. Method 2: Numerical method

To obtain a rigorous description of the microswimmer dynamics in the rectangular channel, we use lattice-Boltzmann based numerical method and solve Eqs. (6)–(7). We follow the method discussed in Ahana and Thampi [9] to calculate the spatiotemporal evolution of the fluid flow driven by the squirmer. The fluid motion and the dynamics of the microswimmer are coupled to each other as follows. If the microswimmer has a translational velocity \hat{U} , a rotational velocity $\hat{\Omega}$, and a surface slip velocity \hat{U}_s [given by Eq. (1)] then the velocity on the surface of the squirmer is given by

$$\hat{\boldsymbol{u}}^{s}(\hat{\boldsymbol{r}}) = \hat{\boldsymbol{U}}_{s}(\hat{\boldsymbol{r}}) + \hat{\boldsymbol{U}} + \hat{\boldsymbol{\Omega}} \times \hat{\boldsymbol{r}}. \tag{14}$$

The boundary condition given by Eq. (14) is incorporated in the lattice-Boltzmann frame work by implementing a midgrid bounce-back scheme [53]. At the same time, the dynamics of the microswimmer is dictated by the stresses exerted by the fluid on the microswimmer. The total force \hat{F} and torque \hat{L} on the microswimmer can be computed [26] and are used to update the translational and angular velocity of the microswimmer respectively, as described in Nguyen and Ladd [54],

$$m_s \hat{U}(t + \Delta t) = m_s \hat{U}(t) + \Delta t \hat{F}(t)$$
(15)

$$I_{s}\hat{\mathbf{\Omega}}(t+\Delta t) = I_{s}\hat{\mathbf{\Omega}}(t) + \Delta t\hat{\mathbf{L}}(t)$$
(16)

where $m_s = (4/3)\pi \rho_s a^3$ is the mass and $I_s = (2/5)m_s a^2$ is the moment of inertia of the squirmer.

A D3Q19 model of LBM is used in our simulations, which solves fluid motion in three dimensions with 19 discrete velocity directions. The spatial and temporal resolutions in LBM are chosen as unity, and a single relaxation BGK collision operator was used. Simulations are performed in a domain of dimensions $200\times32\times32$ (200 points in the x direction) for the 2D confinement. The domain is typically chosen as $200\times64\times64$ to simulate 3D confinement, and wall boundary conditions are applied using a midgrid bounce-back scheme [55] on the four boundaries in the y and z directions.

IV. RESULTS AND DISCUSSION

Our results, presented in this section, are organized as follows. We start by computing the instantaneous velocity of the squirmer located at various positions inside the channel while its orientation is parallel to the channel axis. These results will enable us to understand the origin of the 2D trajectories that squirmers follow inside the channel, which are discussed subsequently. We will then examine the 3D trajectories of squirmers in square channels (AR = 0) and in rectangular channels (AR > 0). The principle of superposition is described afterwards to understand the apparently complex 3D trajectories.

A. Instantaneous velocity of squirmers

The instantaneous swimming velocity of a squirmer in the square channel, oriented parallel to the channel center line $(\theta = 0)$, is calculated as a function of distance from the center line y^* and for different confinement ratios a/H. The results are plotted as follows: the instantaneous velocity parallel to the channel axis U_x and perpendicular to the channel axis U_y is plotted in Figs. 3(a) and 3(b) respectively, and in Fig. 3(c) the angular velocity in the z direction (Ω_z) is depicted.

The swimming velocity parallel to the channel axis, U_x , is independent of the second mode of squirming B_2 . Hence, Fig. 3(a) is the same for pushers, pullers, and neutral swimmers. This happens due to the fore-aft symmetry of the second squirming mode, $B_2 \sin(2\theta)$, which does not generate any wall-induced velocity along the channel length for a microswimmer oriented parallel to the channel axis [20]. As seen in Fig. 3(a), U_x decreases with increase in confinement, a/H, and with increasing

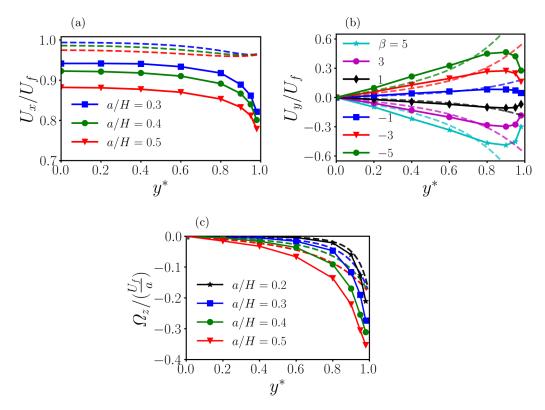


FIG. 3. Instantaneous swimming velocity of a squirmer in a square channel when it is initially located at $(0, y^*, 0)$ with its orientation parallel to the channel axis $(\theta = 0)$. (a) Swimming velocity in the x direction, U_x , scaled by the swimming speed in unconfined fluid U_f , for various values of a/H. (b) Swimming speed U_y in the y direction, scaled by U_f , for different values of β when a/H = 0.3. (c) The instantaneous angular velocity of the swimmer in the direction normal to the plane of locomotion Ω_z , scaled by the quantity U_f/a , for different values of a/H. Plot (c) is same for all types of squirmers, since the induced angular velocity due to B_2 mode will vanish for $\theta = 0$. Dashed lines here (and throughout this study) represent the results from the far-field expressions. The far-field approximation provides qualitatively accurate predictions for U_x , U_y , and Ω_z .

distance from the center line, y^* . The sharp decrease in U_x for $y^* > 0.8$ is an indication of strong interaction of the squirmer with the nearby wall. The far-field approximation predicts qualitatively similar behavior for $y^* < 0.9$, which is shown as dashed lines in Fig. 3(a). Far-field approximation fails for $y^* > 0.9$ since these approximations are highly unstable when the microswimmer comes in the immediate vicinity of a wall. Unlike U_x , the instantaneous velocity in the y direction U_y is dependent on β . Therefore, U_y is plotted against the squirmer location, y^* , by varying β for a fixed confinement ratio of a/H = 0.3. The magnitude of U_y is equal, but the signs are opposite for pushers and pullers of same strength. Pullers ($\beta > 0$) have a negative velocity along the y direction ($U_y < 0$), and accordingly, they are repelled from the nearest wall. In contrast, pushers ($\beta < 0$) are attracted towards the nearest boundary wall ($U_y > 0$). Similar results are also obtained from the far-field calculations [Eq. (13)] for $y^* < 0.8$. As the squirmer gets very close to the wall ($y^* > 0.8$), the resulting hydrodynamic interaction reduces the strength of U_y substantially as shown in Fig. 3(b). It may also be seen that $|U_y|$ increases as the stresslet strength of the pushers and pullers increases.

As shown in Fig. 3(c) hydrodynamic interaction of the squirmer with the channel boundaries also results in a wall-induced angular velocity which changes the orientation of squirmers. The angular velocity Ω_z increases with increase in both y^* and a/H, a manifestation of strong hydrodynamic interaction between the nearest wall and the finite-sized squirmer. Positive (negative) value of

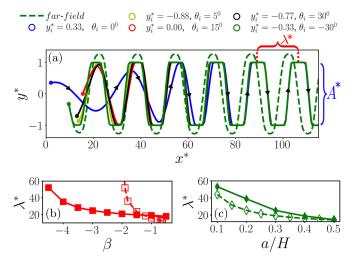


FIG. 4. (a) Oscillatory motion of pushers in the x-y plane. Simulations with different initial conditions (y_i^*, θ_i) give trajectories of the same wavelength (λ^*) and amplitude (A^*) . Variation of λ^* with (b) the strength of the second mode of squirming, β , and (c) the confinement ratio a/H.

angular velocity corresponds to anticlockwise (clockwise) rotation of the squirmer. Far-field expressions predict similar variations in Ω_z with increase in y^* and confinement. The results indicate that all squirmers rotate away from the closest wall. This is a consequence of the hydrodynamic interaction due to the finite-size of the microswimmer, as captured by the first squirming mode or the source dipole in the far-field calculation. The angular velocity due to the source dipole has a $\cos\theta$ dependence, which generates a nonzero angular velocity even when the squirmer is oriented parallel to the channel axis.

B. 2D motion in square channels

In this section, we focus on the 2D trajectories of the squirmers inside a square channel. The initial location and orientation of the squirmer are identified by specifying a subscript i to the relevant variable. In order to obtain 2D trajectories, the squirmer is initially placed in the x-y plane with its orientation vector lying in this plane, i.e., $z_i^* = 0$, $\delta_i = 0$. The symmetry of the system ensures that the squirmer remains in the x-y plane as it propels and generates a 2D trajectory.

1. Oscillatory trajectories of pushers

Our simulations show that pushers make periodic, oscillatory trajectories in the center plane of the channel as plotted in Fig. 4(a). As the pusher approaches the wall, angular velocity Ω_z induced by the hydrodynamic interaction with the wall [Fig. 3(c)] rotates the squirmer away from the nearest wall. This is the key reason for the existence of oscillatory trajectory of pushers. During such motion, pushers swim across the entire channel width, i.e., the channel width determines the amplitude, A^* , of the oscillatory trajectory. At the same time, the wavelength (λ^*) of the trajectory is set by the channel width, confinement, and β . Both λ^* and A^* of the trajectories are found to be independent of initial conditions. The oscillatory trajectories are also qualitatively similar to those generated by circular squirmers in 2D channels [26].

The prediction from the far-field calculation is plotted in Fig. 4(a) as a dashed line. The far-field analysis also gives an oscillatory trajectory, but with a slightly different amplitude and wavelength. The trajectories from the far-field calculations, for a given β and a/H, are also independent of initial conditions. It is important to note here that we have to consider both the force and source dipoles in the far-field calculations to reproduce the oscillatory trajectory of a confined microswimmer,

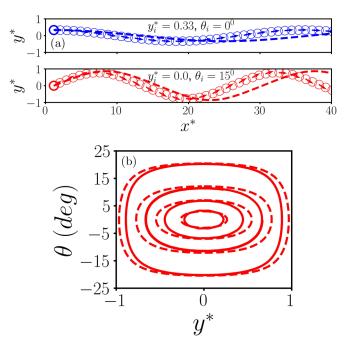


FIG. 5. Periodic oscillations of channel confined neutral swimmers: (a) Oscillatory trajectories in x-y plane corresponding to two different initial conditions. The arrows show the instantaneous orientation of the squirmer. (b) Phase diagram in the (y^*, θ) plane showing the periodic oscillation of neutral swimmers as stable limit cycles. Each closed curve represents oscillation corresponding to a particular initial position and orientation.

as predicted by the numerical simulation. Considering solely the force dipole singularity results in a sliding trajectory along the wall for a pusher, which will be an erroneous prediction for a microswimmer in a confinement. Also, the higher-order terms, i.e., the terms $\sim 1/r^4$ and $\sim 1/r^5$ in Eq. (13), are essential to reproduce the oscillations of pushers. The contributions to the velocity from these higher-order terms are opposite to the first-order terms and become significant when the pusher approaches the wall. These effects, along with the finite angular velocity induced by the source dipole singularity, prevent the unphysical crossing of the rigid wall by the microswimmer.

On analyzing the trajectories further, we find that λ^* exhibits a strong dependence on the dipole strength of pushers and the confinement ratio. This dependence, for both the results from the numerical simulations and the far-field calculations, is illustrated in Figs. 4(b) and 4(c). Strong pushers spend more time closer to the wall due to their stronger velocity component towards the wall, and make larger escape angles. Accordingly, the wavelength of the trajectory increases with the increase in the strength of pushers [Fig. 4(b)]. In contrast, λ^* decreases with the increase in confinement ratio, a/H [Fig. 4(c)]. Strong confinement results in stronger hydrodynamic interaction of the finite-sized microswimmer with the walls, which manifests in greater angular velocity and stronger repulsion from the wall; accordingly, pushers escape quickly.

2. Periodic oscillation of neutral swimmers

Next, we move on to the trajectory analysis of a neutral swimmer. We find that a neutral swimmer always follows a periodic oscillatory trajectory in the channel with λ^* and A^* that are *dependent* on initial conditions. Figure 5(a) illustrates an example, where trajectories of a neutral swimmer for two different initial conditions $y_i^* = 0.33$, $\theta_i^* = 0^\circ$ (top) and $y_i^* = 0.0$, $\theta_i^* = 15^\circ$ (bottom) are shown.

Unlike pushers, neutral swimmers immediately escape as they approach the walls. Again, this is a consequence of the nonzero angular velocity Ω_z , which changes the orientation of the squirmer

as it approaches the wall. The amplitude of the trajectory is constant over time and is two times the initial distance from the channel center line $(A^* = 2y_i^*)$ if the squirmer is initially oriented parallel to the wall $(\theta_i = 0)$. These results are qualitatively consistent with the behavior of neutral swimmers inside a capillary tube [34] and circular neutral swimmers in 2D channels [26]. In this case also, the far-field predictions are in good agreement with the simulation data [Fig. 5(a)]. It is to be noted that the strength of the force dipole is zero for neutral swimmers. The far-field velocities have contributions only from the source dipole, and are capable of reproducing oscillatory trajectories with wavelength and amplitude chosen by the initial conditions.

The oscillatory trajectories of neutral swimmers can be characterized by two parameters: the distance from the center line (y^*) and the orientation of the squirmer (θ) . Therefore, the oscillatory trajectories corresponding to different initial conditions can be easily represented in a phase portrait in the (y^*, θ) plane as in Fig. 5(b). For every initial condition except $(y_i^* = 0, \theta_i = 0)$, the neutral swimmer exhibits a periodic oscillation which represents a stable limit cycle in the phase portrait. The initial condition $(y_i^* = 0, \theta_i = 0)$ is an unstable fixed point that corresponds to the translation of the squirmer through the center line of the channel without any deflection. However, a slight perturbation will produce an oscillatory trajectory. Even though both pushers and neutral swimmers follow periodic trajectories, as we see in the later sections, the 3D motion of neutral swimmers are more challenging to understand due to the strong dependence of their trajectories on the initial conditions.

3. Sliding motion of pullers

In contrast to pushers and neutral swimmers, pullers exhibit a sliding motion in the channel. This sliding can be either through the channel center line or along a straight path close to the walls depending on the strength of the puller (β) and the initial conditions.

Figure 6(a) illustrates sliding of a puller ($\beta = 1$) through the channel center line. Before reaching this steady state, pullers exhibit transient oscillations, the amplitude of which gradually decreases, and finally they swim in a straight-line trajectory. During this straight line motion through the channel center line, they maintain an orientation towards the +ve x axis. Interestingly, stronger pullers, say, $\beta = 5$ as shown in Fig. 6(b), after exhibiting damped oscillations end up in a path parallel to the channel axis, but not along the center line. This trajectory is nearer to one of the walls depending on the initial conditions, and the squirmer will be oriented slightly towards the nearest wall. In this configuration both the wall-induced translational velocity normal to the wall and the angular velocity are simultaneously zero, which sustains the straight line motion.

These results imply the existance of a critical strength of pullers, $\beta_c \approx 3.55$, below which pullers slide through the channel center line and above which they slide near to one of the walls. A similar observation was also made for microswimmers in a capillary [34]. Our simulations show that a strong puller converges to its equilibrium trajectory close to a wall quickly [Fig. 6(b)], while a weak puller ($\beta < 3$) takes a longer time to attain the sliding trajectory along the channel center line [Fig. 6(a)]. Note that the initial condition ($y_i^* = 0$, $\theta_i = 0$) is a special condition which results in the straight line translation of the puller through the channel center line.

As in the previous sections, the far-field analysis successfully predicts the trajectory of weak pullers through the channel center line. However, it fails to predict the sliding motion near one of the walls for a strong puller [Fig. 6(b)] illustrating the role of source dipole in determining the behavior of a puller when it is close to the confining surface. As we increase the strength of the force dipole, we find a transition (not shown) from sliding through the center line to a trajectory where pullers orient perpendicular to the nearest wall and get trapped by that wall (hovering behavior).

4. Phase diagram and the role of initial conditions

In this section, we construct a phase diagram showing the behavior of a squirmer for different strength, β , and confinement ratio, a/H. We also report that the phase diagram depends on the

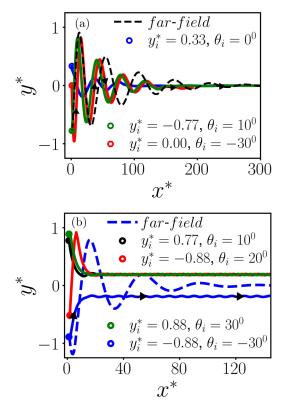


FIG. 6. Sliding motion of pullers in x-y plane for various initial conditions for (a) $\beta = 1$ and (b) $\beta = 5$. The swimmers exhibit initial, transient oscillations, and finally they slide along the channel center line as in (a) or closer to one of the channel walls as in (b) depending on their strength.

initial conditions by comparing it for two different initial conditions: initial orientations $\theta_i = 60^\circ$ and $\theta_i = 30^\circ$ for fixed initial position $y_i^* = 0$.

Figures 7(a) and 7(b) show that five kinds of swimming motions can be observed: sliding along the wall, near the wall, along the channel center line, and then hovering and periodic oscillations. Neutral swimmers mainly perform oscillatory motion except in very weak confinements $(a/H \lesssim 0.1)$. Pushers perform both sliding along a wall and oscillatory motion depending on the values of β and a/H. The stronger the β , the larger the a/H at which oscillatory behavior emerges. At the same time, pullers exhibit hovering behavior and sliding along the channel center line or near a wall depending upon the confinement ratio a/H and the initial condition. Overall, on comparing the two phase diagrams in Fig. 7, it is evident that initial conditions play an important role in deciding the trajectories of channel-confined squirmers.

In fact, it is clear from these phase diagrams that the sliding motion of pushers and hovering behavior of pullers occur for a larger range of confinement ratios as the strength of the squirmer (β) increases. This behavior is also consistent with the predictions of far-field approximations, as we can get the sliding motion of pushers and the hovering behavior of pullers as we increase the strength of force dipole in the far-field calculations.

C. 3D trajectories in square channels

In this section, we will analyze the 3D trajectories of squirmers in a square channel (AR = 0). Similar to the findings in Fig. 7, we see that the 3D trajectories are also dependent on the initial conditions. Therefore, in general, it is hard to categorize the trajectories, and we look for common

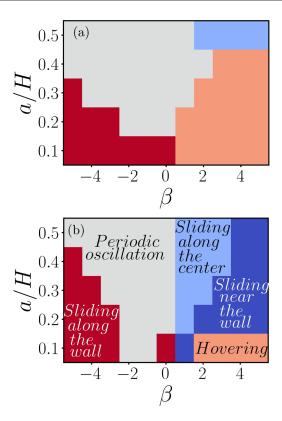


FIG. 7. Phase diagram in the β -a/H plane showing different behavior of squirmers in the midplane of a square channel for two different initial conditions: (a) $y_i^* = 0$, $\theta_i = 60^\circ$ and (b) $y_i^* = 0$, $\theta_i = 30^\circ$ /s. The legends are same for both plots.

and contrasting features of the trajectories of various swimmers as we discuss below. Such a discussion will give us interesting insights into the behavior of channel confined microswimmers. In the simulations reported here, the squirmers are initially placed in the *x-y* plane, away from the channel center in the *z* direction ($z_i^* = \pm 0.77$) with orientation θ_i ($-30^0 \le \theta_i \le 30^\circ$), which gives the squirmer a 3D trajectory inside the channel.

1. Helical trajectories of pushers

We will first study the case of pushers. We have already seen that pushers make an oscillatory motion (Sec. IV B 1) in the x-y plane. Similarly, in three dimensions, pushers exhibit a helical trajectory as illustrated in Fig. 8(a). After the initial transience, pushers execute helical trajectories which span the entire channel. As in two dimensions, here we find that the amplitude of these helical trajectories is independent of initial conditions. The 2D projections of these helical trajectories on y^* - z^* plane is a closed curve as shown in the inset. The start and end points and the direction of motion are also indicated in these figures. It can be seen that the helices conform to the inside surface of the channel, and hence the amplitude of the helix is determined by the channel size.

An interesting variation of the aforementioned helical trajectories happens when the pusher is initially placed at a point that is equidistant in the y and z directions from the center line $(y_i^* = z_i^*)$ and with an orientation parallel to the x axis $(\theta_i = 0)$. In this position, the hydrodynamic interactions and the wall-induced velocities in the y and z directions will be equal in magnitude, and the pusher will exhibit an oscillation through a line connecting opposite edges, which we refer to as an edge-to-edge oscillation [Fig. 8(d)].

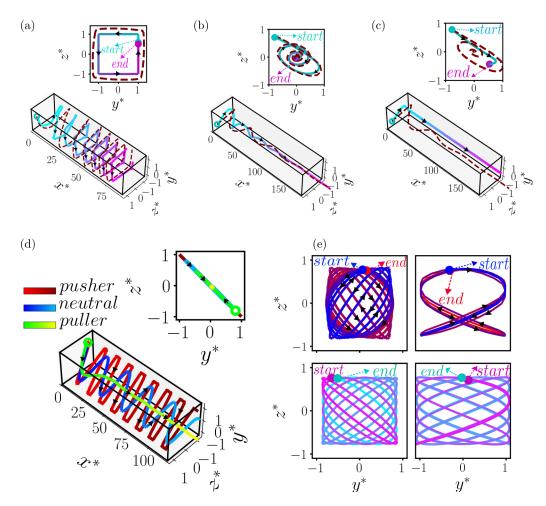


FIG. 8. The 3D trajectory of microswimmers in a square channel (a) of the pusher with initial conditions $y_i^* = 0.0$, $z_i^* = 0.77$, and $\theta_i = 15^\circ$, (b) of a weak puller ($\beta = 1$) with initial conditions $y_i^* = -0.77$, $z_i^* = 0.77$, and $\theta_i = 10^\circ$, and (c) of a strong puller ($\beta = 5$) with same initial conditions as in (b). The projected trajectory in the y^* - z^* cross section is also shown (excluding the initial transients) on the right top of each figure. (d) The 3D trajectory of all three type of microswimmers with an initial condition $y_i^* = z_i^* = 0.77$ and $\theta_i = 0^\circ$, and they move in a plane that connect opposite edges of the channel. (e) The 3D trajectory of a neutral swimmer projected to the y^* - z^* plane (excluding transients) for different initial conditions (top two figures) compared with corresponding far-field trajectories (bottom figure). The start and end points are also marked in the trajectory plots.

The trajectories calculated using the far-field approximation are also shown (dashed lines) in Fig. 8(a). These calculations are in good agreement with the simulation data. The y^*-z^* plane of the trajectory is shown in the inset. Note that the far-field calculations show two different steady-state behaviors for channel-confined pushers. The first one is the helical trajectory, as shown in Fig. 8(a). The second one is the case wherein the pusher does not translate the x direction, but it orbits the y-z plane spanning the entire channel [dashed line in Fig. 9(a)]. These steady-state behaviors are dependent on the strength of the source dipole, \bar{q} , for a given value of the force dipole. The transition between these two steady-state behaviors for increasing values of \bar{q} are shown in the phase plots in Fig. 9(b). The angle that the pusher makes with the y axis is plotted against y^* , and the trajectories (stable limit cycles) are color coded with the displacement in the x direction. This displacement is

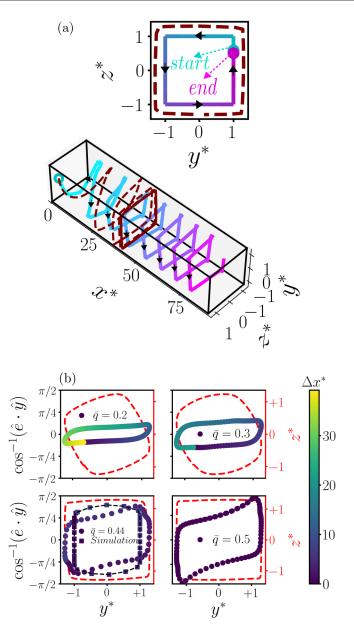


FIG. 9. (a) Comparison of the 3D trajectory of pusher from simulation and far-field calculation (dotted line) for $\bar{q}=0.5$. The far-field calculations predict a steady state where pusher continuously orbits in the *y-z* plane without moving along the *x* axis. (b) The phase plots, from far-field calculations, showing the angle of orientation (angle between \hat{e} and the *y* axis) variation with y^* for various values of \bar{q} . The color bar represents the displacement in the *x* direction while completing one cycle. The red dashed lines are the y^* - z^* cross section.

zero for $\bar{q}=0.5$, and the phase plot corresponds to the steady state where the pusher continuously orbits in the y-z plane without moving along the x axis. When \bar{q} is reduced to 0.44, the pusher exhibits finite displacement along the x direction, while swimming in a helical trajectory. Note that the results from the numerical simulation and the far-field calculations match quite well for $\bar{q}=0.44$. Hence, we have considered $\bar{q}=0.44$ in our far-field calculations reported in Fig. 8(a).

With further reduction in \bar{q} , the pusher continues to execute the helical motion, but does not adhere to the contours of the channel cross section [red dashed lines in Fig. 9(b)]. In summary, far-field calculations for pushers predict that for a particular force dipole strength, there is a critical source dipole strength above which the pusher does not move along the channel axis, and below which the pusher executes helical motion. However, it must be noted that numerical simulations did not show such a change in behavior for pushers.

2. 3D motion of neutral swimmers

We have already seen that neutral swimmers execute oscillatory trajectories in two dimensions, and the wavelength and amplitude of these trajectories are dependent on initial conditions. Similarly, in three dimensions, neutral swimmers exhibit initial-condition-dependent, complex trajectories. Examples of such trajectories, projected onto the y^* - z^* plane, are shown in Fig. 8(e). In the top-left figure shown in Fig. 8(e), the initial conditions are $y_i^* = 0.11$, $z_i^* = -0.77$, and $\theta_i = 15^\circ$. In this case, the neutral squirmer follows a clockwise trajectory followed by an anticlockwise trajectory while translating along the channel length thus making an aperiodic helical trajectory. In the top-right example of Fig. 8(e), the initial conditions are $(y_i^* = 0, z_i^* = 0.77, \text{ and } \theta_i = -30^\circ)$. In this case, the neutral swimmer exhibits a ribbon-like pattern in the y^* - z^* plane. Such initial condition dependency, and the variety of the trajectories, make it hard to characterize the trajectories of channel-confined neutral swimmers. Nevertheless, the origin of this behavior will be discussed later in Sec. IV C 4. Note that the initial conditions $y_i^* = z_i^* = 0.77$ and $\theta_i = 0$ would give rise to a edge-to-edge oscillating trajectory [Fig. 8(d)] as we saw in the case of pushers.

The bottom panel of Fig. 8(e) shows the far-field trajectory of the corresponding simulation in the top panel. In this case, the far-field trajectories (dashed lines) may look different from the results from the numerical simulations. As we will see, the superposition principle described in Sec. IV C 4 will explain this difference observed from the results of two methods.

3. Sliding motion of pullers along or parallel to the channel center line

Figures 8(b) and 8(c) show the 3D trajectories of pullers and their projections onto the y^*-z^* plane. Trajectories in both figures correspond to same initial conditions $y_i^* = -0.77$, $z_i^* = 0.77$, and $\theta_i = 10$, but with increasing strength of the puller. For a weak puller [Fig. 8(b)], the trajectory exhibits a spiralling motion initially, but the amplitude of the spiral decreases eventually and the puller slides along the channel center line. However, for a stronger puller [Fig. 8(c)], the sliding motion is closer and parallel to one of the walls. These results are also consistent with the sliding motion of pullers observed in two dimensions (discussed in Sec. IV B 3).

The far-field calculations (dashed lines) predict a comparable trajectory for a weak puller, $\beta = 1$ [Fig. 8(b)]. But, as in the 2D case, the far-field calculations fail to predict the sliding motion near a wall for strong pullers in three dimensions.

4. Understanding 3D trajectories as a superposition of 2D trajectories

In this section, we introduce a simple technique to understand and predict the 3D trajectories of a channel confined squirmer. The idea is to utilize the results obtained in the previous sections, specifically the trajectories in two dimensions, and to use the principle of superposition to construct the behavior of channel confined swimmers in three dimensions.

Let us first consider the 2D trajectory of a squirmer confined to x-y plane of the channel. In general, the trajectory can be mathematically expressed as

$$y(x) = A \sin\left(\frac{2\pi}{\lambda}x + \Delta\right) \exp(-mx),\tag{17}$$

where A is the amplitude, λ is the wavelength, Δ is the phase of the oscillations, and m is a decay constant. The steady-state trajectories of pushers and neutral swimmers are oscillatory and periodic, which can be represented by this general expression with m = 0. The transient path of pullers which

follow an exponential decay, but settle to a sliding trajectory can be represented with $m \neq 0$. For pullers, the initial conditions determine the exact value of m. Similarly, for neutral swimmers A and λ are also determined by initial conditions.

An identical expression for the 2D trajectory of a squirmer in the x-z plane can be written as

$$z(x) = \tilde{A}\sin\left(\frac{2\pi}{\tilde{\lambda}}x\right)\exp(-\tilde{m}x),\tag{18}$$

where \tilde{A} , $\tilde{\lambda}$, and \tilde{m} have the same meaning as given above, and their values are determined by the squirmer parameters, the channel dimensions, and the initial conditions.

We propose that the 3D trajectory of a squirmer confined in a channel can be constructed by assuming a superposition of the 2D motions in the two orthogonal planes, namely, x-y and x-z planes. Therefore, the 2D position vector in the y-z plane, $\mathbf{T}_s(x) = y(x)\hat{y} + z(x)\hat{z}$, describing the trajectory of the squirmer as a function of x can be expressed as

$$\mathbf{T}_{s}(x) = A \sin\left(\frac{2\pi}{\lambda}x + \Delta\right) \exp(-mx)\hat{\mathbf{y}} + \tilde{A}\sin\left(\frac{2\pi}{\tilde{\lambda}}x\right) \exp(-\tilde{m}x)\hat{z}. \tag{19}$$

Thus, the assumption of superposition yields the above expression to determine the trajectory of the squirmer in three dimensions from its 2D counterparts. We now analyze the trajectory obtained from Eq. (19) with the actual trajectories.

Pushers: The amplitude and wavelength of 2D trajectories of pushers are independent of initial conditions. Therefore, in channels of square cross section $\tilde{A} = A$ and $\tilde{\lambda} = \lambda$. Eliminating x from Eq. (19), we can derive an expression for the motion of the squirmer in y-z plane as

$$\frac{z^2}{A^2} - \frac{2zy}{A^2}\cos\Delta + \frac{y^2}{A^2} = \sin^2\Delta,$$
 (20)

which is the general equation for an ellipse. Equation (20) implies that the 3D trajectory of a channel confined pusher will be a helix. The shape of the helix will be determined by the parameter Δ , the phase difference between the waves in the *x*-*y* and the *x*-*z* planes. For $\Delta = 0$, when there is no phase difference between the 2D trajectories considered in the two orthogonal planes, Eq. (20) reduces to the equation of a straight line y = z. This case corresponds to the edge to edge oscillations seen for a pusher. For $\Delta \neq 0$, the phase difference between the waves results in the 3D helical trajectory of the pushers [Fig. 8(a)].

In the above discussion, we approximated the 2D, oscillatory motion of pushers as a sinusoidal function to qualitatively explain the behavior in three dimensions. Instead of these expressions [Eqs. (17) and (18)], we can superpose the actual 2D trajectories obtained from the simulations to perform the superposition. Succinctly put, the trajectories of the squirmer when restricted (by symmetry) to move in the x-y and x-z planes in a square channel obtained from simulations can be superposed to construct the corresponding 3D trajectory, without actually solving for the 3D geometry. An example of such a result is plotted in Fig. 10(a). The trajectory from the full-scale 3D simulation (red line) is also shown in this figure. The agreement between the two approaches is quite good, indicating that the superposition principle works in generating 3D trajectories of pushers. More importantly, the principle of superposition explains the origin of the nature of the 3D trajectory.

Pullers: A similar comparison between the 3D trajectories from the full-scale numerical simulation and the superposition method for pullers also illustrates the efficacy of the latter. For example, consider the comparison made for pullers in a 3D channel with initial condition, $y_i^* = 0.0$, $z_i^* = 0.77$, and $\theta_i = 15^\circ$. We can split this problem into the superposition of two 2D cases: the first one in the x-y plane with $y_i^* = 0.0$, $z_i^* = 0.0$, $\theta_i = 15^\circ$, which isolates the effects of top and bottom wall, and the second one in the x-z plane with initial conditions $y_i^* = 0.0$, $z_i^* = 0.77$, and $\theta_i = 0^\circ$, which isolates the effects of the side walls. Using the results of these two simulations, a 3D trajectory is constructed in Fig. 10(b). This figure highlights the fact that the transient motion

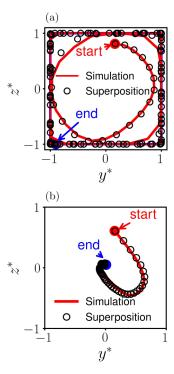


FIG. 10. Comparison of the trajectories (projected on to y^*-z^* plane) resulting from full-scale 3D simulation and from the superposition of 2D trajectories in the x-y and x-z planes (a) for pushers and (b) for pullers. In both (a) and (b) the initial conditions are $y_i^* = 0.0$, $z_i^* = 0.77$, and $\theta_i = 15^\circ$.

of pullers can be also understood as the superposition of two mutually perpendicular damping oscillations.

Neutral swimmers: We have already seen that the initial conditions affect the wavelength and amplitude of the oscillatory trajectory of neutral swimmers even in two dimensions. Therefore, the trajectories for a neutral swimmer obtained in the x-y and x-z plane can have different wavelengths and amplitude. Moreover they differ in the phase too. Thus, the trajectory of a neutral swimmer can be understood as the superposition of two mutually perpendicular waves with different wavelengths and amplitudes. Naturally, the resultant trajectory will be complex as illustrated in Fig. 8(e) (Sec. IV C 2). However, interestingly, there can be an exceptional initial condition for which the amplitude, wavelength, and phase of oscillatory motion in x-y and x-z planes exactly match, i.e., $A = \tilde{A}$, $\lambda = \tilde{\lambda}$, $\Delta = 0$, and hence, we get a edge-to-edge oscillation [Fig. 8(d)] as we reported before.

Further, this analysis also allows us to comprehend the mismatch in the predictions of numerical simulations and far-field calculations reported in Fig. 8(e) (Sec. IV C 2). As mentioned above, in two dimensions, a neutral swimmer performs oscillations whose amplitude and wavelength depend on initial conditions. The far-field calculations predict 2D oscillations with slightly different amplitudes and wavelengths. Such differences in amplitude and wavelength may create a noticeable change while constructing the 3D trajectory as can be seen in Fig. 8(e).

In summary, the superposition principle constructs the 3D trajectories of channel confined pushers, pullers, and neutral swimmers. The match obtained between the results of the numerical simulation and that from the superposition provides an insight into the dominant effects that drive the dynamics of a squirmer in a 3D channel. If the motion of the squirmer is analyzed from the perspective of method of reflections, the net effect due to the four walls of the channel can be considered as the sum of (1) hydrodynamic interaction with the channel walls parallel to the *x-y*

plane, (2) hydrodynamic interaction with the channel walls parallel to the x-z plane, and (3) the consequent series of reflections from the two pairs of walls in order to simultaneously satisfy the boundary conditions at all the walls. Clearly, the above results illustrate that the effect of the last contribution, i.e., the consequent reflections, is much weaker and the principle of superposition calculates the approximate trajectory of the squirmer in three dimensions. Although this method is not exact, it is sufficient to explain the origin of sliding motion of pullers and the helical motion of pushers in a square channel. This method also helps to reason the complex trajectory of neutral swimmers.

D. 3D motion of squirmers in rectangular channels

We conclude the study by extending our trajectory analysis to the case of squirmers in rectangular channels (AR > 0). We carried out simulations by keeping the height of the channel fixed, while increasing the width of the channel systematically. The trajectories of the squirmers appeared complex as we increased the aspect ratio AR. Note that AR = 0 for a channel with square cross section [Eq. (5)].

In this section, we report the trajectories of different types of squirmers in a rectangular channel of AR = 0.5, and explain the origin of the apparent complexity of the trajectory using the superposition principle.

We start by analyzing the case of pushers. The trajectories of pushers in this case are not easy to generalize since they do not follow the simple helical trajectories as observed in a square channel. Instead the trajectory appears complex, and these 3D trajectories strongly depend on the confinement ratio and aspect ratio of the channel. In Fig. 11(a), the trajectory of a pusher, initially located at $y_i^* = 0$, $z_i^* = 0.66$ and oriented at $\theta_i = -30^\circ$, is shown in the y^* - z^* plane. The left plot shows the simulation result, and the right plot is the far-field calculation. The trajectory is plotted with a color gradient for easy visualization of the trajectory. This seemingly complex trajectory can, again, be explained from the superposition method as follows. We have seen that the wavelength of the trajectory of a pusher increases with decreasing the confinement ratio, a/H [Fig. 4(c)]. In a rectangular channel, the width of the channel is more than the height. Therefore both the amplitudes and the wavelengths of the trajectories produced in the planar trajectories belonging to x-y and x-z planes will be different, i.e., $A \neq \tilde{A}$ and $\lambda \neq \tilde{\lambda}$. The superposition of these two different waves gives rise to the appearance of the complex 3D trajectory of the pusher shown in Fig. 11(a).

Next, we discuss the trajectories of pullers. They exhibit sliding motion in a rectangular channel as well. As discussed earlier, the sliding can be either along the channel center line or close to one of the walls. Moreover, the initial location and orientation dictate the path to reach the final trajectory. Consider the case of a puller initially located at $y_i^* = 0$, $z_i^* = 0.66$ and oriented at $\theta_i = -30^\circ$ in a rectangular channel. The y^* - z^* plane view, and the 3D trajectory followed by the puller is plotted in Figs. 11(c) and 11(d) respectively. This swimmer eventually slides along the channel center line for which $y^* = 0$ and $z^* = 0$. The transient path that leads to this final state can be analyzed as follows. During the motion, the puller first reaches the $y^* = 0$ plane, namely, the center of the x-y plane. Then it moves only along the z direction to the point $z^* = 0$, thus reaching the channel center line. Hence, the entire trajectory of the puller can be understood as a combination of two individual trajectories, consistent with the principle of superposition.

In rectangular channels, as we increase AR, neutral swimmers also generate quite complicated trajectories. An example of the trajectory of a neutral swimmer in the y^* - z^* plane is shown in Fig. 11(b), which has an initial position $y_i^* = 0$, $z_i^* = 0.66$ and orientation $\theta_i = -30^\circ$. Similar to the case of a pusher, the apparent complexity of the trajectory can again be attributed to the fact that it is a superposition of two planar waves of different wavelengths and amplitudes. But unlike the case of a pusher, here the characteristics of the planar waves are not determined by the channel dimensions alone, but are also dependent on the initial configuration of the swimmer [similar to Fig. 5(a)]. Consequently, slight changes in the initial position or orientation can entirely alter the trajectory of neutral swimmers, as we observed in the case of the square channel.

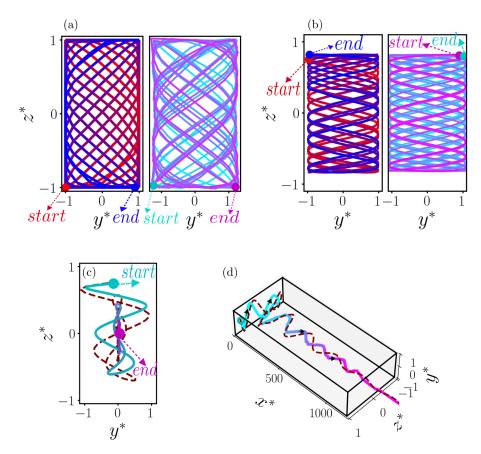


FIG. 11. Trajectories of a microswimmer in a 3D rectangular channel of AR = 0.5. Projection of the trajectory onto the y^* - z^* plane for (a) a pusher, (b) a neutral swimmer, and (c) a puller. Panel (d) shows the 3D trajectory of the puller reported in (c). The initial conditions for the swimmers are $z_i^* = 0.77$ and $y_i^* = 0.0$, $\theta_i = -30^\circ$. In (b) $z_i^* = 0.66$. The second plot in panels (a) and (b) and dashed lines in panels (c) and (d) are trajectories from the far-field calculations.

V. CONCLUSIONS

In this work, we present a detailed study of the 3D trajectories of microswimmers inside a microchannel with a square/rectangular cross section using two methods: (1) full numerical simulations based on the lattice-Boltzmann method and (2) an analytical method based on far-field hydrodynamic approximations combined with the method of images. We use a squirmer with tangential surface velocity as a model for the microswimmer. The hydrodynamic interactions between the channel walls and the squirmer plays a significant role in defining their 3D trajectories.

Analysis of the instantaneous velocity of the squirmer at different positions in the channel reveals that the channel confinement and near-wall swimming always decrease the swimming speed of a squirmer when it is oriented parallel to the channel axis, and induce a strong angular velocity. The far-field analysis captures the nature of these wall-induced dynamics fairly well until about 90% of the physically possible distance of the microswimmer away from the channel center and towards the walls. We also provide a detailed description of various two dimensional trajectories produced by the squirmer when it is confined between two parallel plates, which certainly depends

on the force/source dipole strengths and the extent of confinement. These observations helped us to underpin the behavior of swimmers in microchannels with square and rectangular cross sections.

In general, our analysis of the trajectories of different squirmers in a 3D channel with square and rectangular cross sections shows that the trajectories are closely dictated by three parameters: (1) strength of the confinement from different directions, (2) nature and strength of the swimmer, and (3) initial position and orientation of the swimmer. Pushers are seen to generally perform helical motion inside a square channel, and a qualitatively similar but more complex trajectory inside a rectangular channel. The far-field calculations predict that for a given force dipole strength of a pusher in a square channel, there is a critical value of the source dipole strength (and hence the size of the swimmer) beyond which the microswimmer only orbits the contours of the channel cross section without any displacement along the channel axis. On the other hand, pullers generally make a sliding motion inside the microchannel. Depending on the dipole strength, they choose different paths: weak pullers slide through the channel center line, while strong pullers slide through a straight line close to any one of the channel walls. Neutral squirmers show the most complex 3D trajectories due to their strong dependency on the initial conditions. The far-field hydrodynamic calculations, constructed from the superposition of the hydrodynamic interactions between a squirmer and two pairs of orthogonal walls, can predict the correct trajectories of microswimmers and all the essential features of swimming in two and three dimensions.

Our systematic analysis—determination of instantaneous velocity of the confined swimmers which in turn determines their trajectories, the planar trajectories when the motion of the swimmer is restricted to the symmetric planes of the microchannel, and the 3D trajectories in square channels and finally in the rectangular channels—help us to understand the factors that determine the nature of the 3D trajectory of a channel confined microswimmer. Based on this understanding, we propose a method of superposition to explain the apparent complexity of the 3D trajectories. According to our method, each 3D trajectory can be constructed from the superposition of two mutually perpendicular planar trajectories (trajectories produced when the squirmer is restricted to move in a plane in the channel). We demonstrate that the superposition method works well to explain the variety of trajectories of pushers, pullers, and neutral swimmers in 3D microchannels.

Hydrodynamics of microorganisms inside different 3D confinements are biologically relevant for understanding their locomotion, survival strategies, and collective behavior [38,56]. Clear understanding of the hydrodynamic interactions of microswimmers in strong confinements is also imperative for designing artificial microswimmers and formulating strategies for biomedical applications, like targeted drug delivery [57,58] and targeted interactions with biological cells [59]. However, as we have mentioned in the introduction, a detailed study of the hydrodynamics of microswimmers in 3D confinements remains scarce. In this context, the present research work provides a clearer picture of the possible dynamics of different kinds of microswimmers in 3D channels with a square/rectangular cross section. The unambiguous comparison between numerical simulations and far-field analytical calculations will greatly aid in designing active microfluidic systems for the aforementioned applications in an informed manner. Furthermore, the proposed superposition method for constructing 3D trajectories of microswimmers in strong confinements can be an efficient way of developing physical understanding without going into the complexities of full-scale numerical simulations of 3D systems and experiments.

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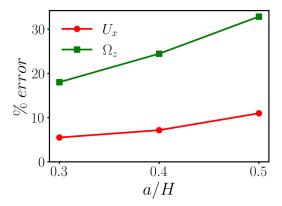


FIG. 12. The percentage error in the instantaneous translational and angular velocities estimated from the far-field predictions, relative to the estimates from the numerical simulations, plotted as a function of confinement, a/H. Initial condition and boundary conditions are the same as in Fig. 3, and $y^* = 0.6$. Note that the error estimates for U_v will be even smaller than that for U_x .

APPENDIX: ACCURACY OF FAR-FIELD CALCULATIONS AND SIGNIFICANCE OF HIGHER-ORDER TERMS

The accuracy of far-field calculations is estimated from the instantaneous translational and angular velocities shown in Fig. 3, relative to those obtained from numerical simulations. Figure 12 shows the percentage error in the instantaneous translational and angular velocities from the far-field calculation, relative to the lattice-Boltzmann simulation calculations. The error reduces as we decrease the confinement, a/H, illustrating the asymptotic convergence of the far-field calculations to the simulation results in the weak confinement limit.

The importance of the higher-order terms in the far-field calculations [Eqs. (11) and (12)] is further demonstrated in Figs. 13(a)–13(c). The trajectory of a pusher confined between two parallel plates is shown in Fig. 13(a). The figure consists of (1) the trajectory obtained from LB simulations (green solid line), (2) the predictions from the far-field calculations (blue dashed line), and (3) the predictions from the far-field calculations without accounting for the higher-order source dipole terms (red dashed and dotted line). The velocity from the far-field approximations without the source dipole terms is calculated by setting $\bar{q} = 0$ in Eqs. (11) and (12). Without these terms, the far-field calculations predict a sliding trajectory along the wall as shown in the figure. Hence, higher-order source dipole terms are necessary to produce an oscillatory trajectory of a pusher similar to results obtained from simulations. The force dipole strength for a neutral swimmer is zero ($\alpha = 0$), and far-field calculations without source dipole terms do not produce any confinement induced deviation in the trajectory because all the terms in Eqs. (11) and (12) vanish. In other words, the oscillatory motion of the confined neutral swimmer is purely the result of source dipole corrections [Fig. 13(b)]. In the case of pullers, far-field calculations without the source dipole terms predict that the puller will reach and get stuck at either one of the confining walls [Fig. 13(c)] depending on the initial conditions. One can obtain trajectories that are comparable to simulations only on inclusion of the higher-order source dipole terms in the far-field calculations. Figure 13 shows that the inclusion of the source dipole terms in the far-field calculations are necessary and sufficient (from the perspective of inclusion of higher-order terms) to capture the essential dynamics of spherical squirmers in confinements.

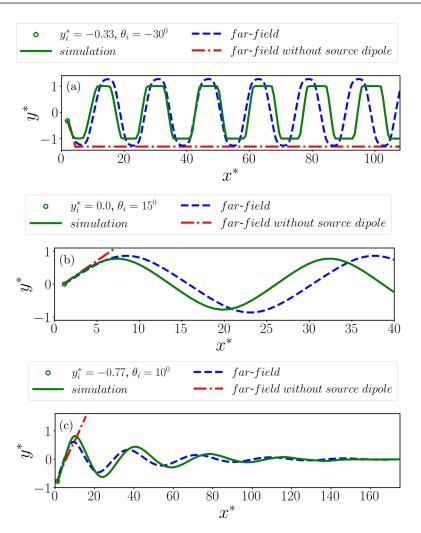


FIG. 13. Comparison between trajectories from simulation and far-field calculations with and without higher-order source dipole terms. The plots are for (a) pusher, (b) neutral swimmer, and (c) puller, and the initial conditions are specified in the legend of each panel.

^[1] D. L. Koch and G. Subramanian, Collective hydrodynamics of swimming microorganisms: Living fluids, Annu. Rev. Fluid Mech. 43, 637 (2011).

^[2] J. C. Conrad and R. Poling-Skutvik, Confined flow: Consequences and implications for bacteria and biofilms, Annu. Rev. Chem. Biomol. Eng. 9, 175 (2018).

^[3] H. Guidobaldi, Y. Jeyaram, C. Condat, M. Oviedo-Diego, I. Berdakin, V. Moshchalkov, L. Giojalas, A. Silhanek, and V. Marconi, Disrupting the wall accumulation of human sperm cells by artificial corrugation, Biomicrofluidics 9, 024122 (2015).

^[4] R. Zimmer and J. Riffell, Sperm chemotaxis, fluid shear, and the evolution of sexual reproduction, Proc. Natl. Acad. Sci. USA 108, 13200 (2011).

^[5] T. Wiles, B. Schlomann, E. Wall, R. Betancourt, R. Parthasarathy, and K. Guillemin, Swimming motility of a gut bacterial symbiont promotes resistance to intestinal expulsion and enhances inflammation, PLoS Biol. 18, e3000661 (2020).

- [6] C. Montecucco and R. Rappuoli, Living dangerously: How *Helicobacter pylori* survives in the human stomach, Nat. Rev. Mol. Cell Biol. **2**, 457 (2001).
- [7] F. Höfling and T. Franosch, Anomalous transport in the crowded world of biological cells, Rep. Prog. Phys. **76**, 046602 (2013).
- [8] C. Bechinger, R. Di Leonardo, H. Löwen, C. Reichhardt, G. Volpe, and G. Volpe, Active particles in complex and crowded environments, Rev. Mod. Phys. 88, 045006 (2016).
- [9] A. Purushothaman and S. P. Thampi, Hydrodynamic collision between a microswimmer and a passive particle in a micro-channel, Soft Matter 17, 3380 (2021).
- [10] E. Lushi, V. Kantsler, and R. E. Goldstein, Scattering of biflagellate microswimmers from surfaces, Phys. Rev. E 96, 023102 (2017).
- [11] W. Uspal, M. Popescu, S. Dietrich, and M. Tasinkevych, Active Janus colloids at chemically structured surfaces, J. Chem. Phys. 150, 204904 (2019).
- [12] A. T. Poortinga, R. Bos, W. Norde, and H. J. Busscher, Electric double layer interactions in bacterial adhesion to surfaces, Surf. Sci. Rep. 47, 1 (2002).
- [13] J. Humphries, L. Xiong, J. Liu, A. Prindle, F. Yuan, H. A. Arjes, L. Tsimring, and G. Süel, Species-independent attraction to biofilms through electrical signaling, Cell 168, 200 (2017).
- [14] K. Kroy, D. Chakraborty, and F. Cichos, Hot microswimmers, Eur. Phys. J. Spec. Top. 225, 2207 (2016).
- [15] Rothschild, Non-random distribution of bull spermatozoa in a drop of sperm suspension, Nature (London) 198, 1221 (1963).
- [16] E. Lauga and T. R. Powers, The hydrodynamics of swimming microorganisms, Rep. Prog. Phys. 72, 096601 (2009).
- [17] D. Woolley, Motility of spermatozoa at surfaces, Reproduction 126, 259 (2003).
- [18] J. Elgeti, U. Kaupp, and G. Gompper, Hydrodynamics of sperm cells near surfaces, Biophys. J. 99, 1018 (2010).
- [19] Y. Li, H. Zhai, S. Sanchez, D. B. Kearns, and Y. Wu, Noncontact cohesive swimming of bacteria in two-dimensional liquid films, Phys. Rev. Lett. 119, 018101 (2017).
- [20] A. P. Berke, L. Turner, H. C. Berg, and E. Lauga, Hydrodynamic attraction of swimming microorganisms by surfaces, Phys. Rev. Lett. 101, 038102 (2008).
- [21] E. Lauga, W. R. DiLuzio, G. M. Whitesides, and H. A. Stone, Swimming in circles: Motion of bacteria near solid boundaries, Biophys. J. **90**, 400 (2006).
- [22] R. Di Leonardo, D. Dell'Arciprete, L. Angelani, and V. Iebba, Swimming with an image, Phys. Rev. Lett. **106**, 038101 (2011).
- [23] S. E. Spagnolie, G. R. Moreno-Flores, D. Bartolo, and E. Lauga, Geometric capture and escape of a microswimmer colliding with an obstacle, Soft Matter 11, 3396 (2015).
- [24] J. S. Lintuvuori, A. T. Brown, K. Stratford, and D. Marenduzzo, Hydrodynamic oscillations and variable swimming speed in squirmers close to repulsive walls, Soft Matter 12, 7959 (2016).
- [25] M. Kuron, P. Stärk, C. Holm, and J. de Graaf, Hydrodynamic mobility reversal of squirmers near flat and curved surfaces, Soft Matter 15, 5908 (2019).
- [26] P. Ahana and S. P. Thampi, Confinement induced trajectory of a squirmer in a two dimensional channel, Fluid Dyn. Res. 51, 065504 (2019).
- [27] K. V. S. Chaithanya and S. P. Thampi, Wall-curvature driven dynamics of a microswimmer, Phys. Rev. Fluids 6, 083101 (2021).
- [28] C. Kurzthaler and H. A. Stone, Microswimmers near corrugated, periodic surfaces, Soft Matter 17, 3322 (2021).
- [29] A. Dhar, P. Burada, and G. Sekhar, Hydrodynamics of active particles confined in a periodically tapered channel, Phys. Fluids **32**, 102005 (2020).
- [30] A. Daddi-Moussa-Ider, C. Kurzthaler, C. Hoell, A. Zöttl, M. Mirzakhanloo, M.-R. Alam, A. M. Menzel, H. Löwen, and S. Gekle, Frequency-dependent higher-order stokes singularities near a planar elastic boundary: Implications for the hydrodynamics of an active microswimmer near an elastic interface, Phys. Rev. E 100, 032610 (2019).
- [31] R. Ledesma-Aguilar and J. M. Yeomans, Enhanced motility of a microswimmer in rigid and elastic confinement, Phys. Rev. Lett. 111, 138101 (2013).

- [32] A. J. Mathijssen, A. Doostmohammadi, J. M. Yeomans, and T. N. Shendruk, Hydrodynamics of microswimmers in films, J. Fluid Mech. 806, 35 (2016).
- [33] O. Liot, M. Socol, L. Garcia, J. Thiéry, A. Figarol, A.-F. Mingotaud, and P. Joseph, Transport of nanoobjects in narrow channels: Influence of Brownian diffusion, confinement and particle nature, J. Phys.: Condens. Matter 30, 234001 (2018).
- [34] L. Zhu, E. Lauga, and L. Brandt, Low-Reynolds-number swimming in a capillary tube, J. Fluid Mech. 726, 285 (2013).
- [35] J. Elgeti and G. Gompper, Run-and-tumble dynamics of self-propelled particles in confinement, Europhys. Lett. 109, 58003 (2015).
- [36] C. Liu, C. Zhou, W. Wang, and H. P. Zhang, Bimetallic microswimmers speed up in confining channels, Phys. Rev. Lett. 117, 198001 (2016).
- [37] J. de Graaf, A. J. T. M. Mathijssen, M. Fabritius, H. Menke, C. Holm, and T. N. Shendruk, Understanding the onset of oscillatory swimming in microchannels, Soft Matter 12, 4704 (2016).
- [38] R. Jeanneret, D. O. Pushkin, and M. Polin, Confinement enhances the diversity of microbial flow fields, Phys. Rev. Lett. 123, 248102 (2019).
- [39] R. D. Schulman, M. Backholm, W. S. Ryu, and K. Dalnoki-Veress, Undulatory microswimming near solid boundaries, Phys. Fluids 26, 101902 (2014).
- [40] C. Pozrikidis, *Boundary Integral and Singularity Methods for Linearized Viscous Flow*, Cambridge Texts in Applied Mathematics (Cambridge University Press, Cambridge, 1992).
- [41] A. T. Chwang and T. Y.-T. Wu, Hydromechanics of low-Reynolds-number flow. Part 2. Singularity method for stokes flows, J. Fluid Mech. 67, 787 (1975).
- [42] S. E. Spagnolie and E. Lauga, Hydrodynamics of self-propulsion near a boundary: Predictions and accuracy of far-field approximations, J. Fluid Mech. 700, 105 (2012).
- [43] O. S. Pak and E. Lauga, Theoretical models in low-Reynolds-number locomotion, in *Fluid–Structure Interactions in Low-Reynolds-Number Flows*, edited by C. Duprat and H. Stone (The Royal Society of Chemistry, Cambridge, 2015), pp. 100–167.
- [44] J. R. Blake, A note on the image system for a stokeslet in a no-slip boundary, Math. Proc. Camb. Philos. Soc. **70**, 303 (1971).
- [45] M. J. Lighthill, On the squirming motion of nearly spherical deformable bodies through liquids at very small Reynolds numbers, Commun. Pure Appl. Maths. 5, 109 (1952).
- [46] J. Blake, A spherical envelope approach to ciliary propulsion, J. Fluid Mech. 46, 199 (1971).
- [47] T. J. Pedley, Spherical squirmers: Models for swimming micro-organisms, IMA J. Appl. Maths. **81**, 488 (2016).
- [48] A. Zöttl and H. Stark, Emergent behavior in active colloids, J. Phys.: Condens. Matter 28, 253001 (2016).
- [49] A. J. T. M. Mathijssen, T. N. Shendruk, J. M. Yeomans, and A. Doostmohammadi, Upstream swimming in microbiological flows, Phys. Rev. Lett. 116, 028104 (2016).
- [50] C. de Blois, M. Reyssat, S. Michelin, and O. Dauchot, Flow field around a confined active droplet, Phys. Rev. Fluids 4, 054001 (2019).
- [51] R. Dey, C. M. Buness, B. V. Hokmabad, C. Jin, and C. C. Maass, Oscillatory rheotaxis of artificial swimmers in microchannels, Nat. Commun. 13, 2952 (2022).
- [52] S. Kim and S. Karrila, *Microhydrodynamics: Principles and Selected Applications*, Butterworth-Heinemann Series in Chemical Engineering (Dover Publications, New York, 2005).
- [53] T. Kruger, H. Kusumaatmaja, A. Kuzmin, O. Shardt, G. Silva, and E. M. Viggen, *The Lattice Boltzmann Method Principles and Practice* (Springer International Publishing, Cham, 2017).
- [54] N.-Q. Nguyen and A. J. C. Ladd, Lubrication corrections for lattice-Boltzmann simulations of particle suspensions, Phys. Rev. E 66, 046708 (2002).
- [55] C. K. Aidun and J. R. Clausen, Lattice-Boltzmann method for complex flows, Annu. Rev. Fluid Mech. 42, 439 (2010).
- [56] V. Tokárová Jr, A. Sudalaiyadum Perumal, M. Nayak, H. Shum, O. Kašpar, K. Rajendran, M. Mohammadi, C. Tremblay, E. A. Gaffney, S. Martel *et al.*, Patterns of bacterial motility in microfluidics-confining environments, Proc. Natl. Acad. Sci. USA 118, e2013925118 (2021).

- [57] F. Mou, C. Chen, Q. Zhong, Y. Yin, H. Ma, and J. Guan, Autonomous motion and temperature-controlled drug delivery of Mg/Pt-poly (*N*-isopropylacrylamide) Janus micromotors driven by simulated body fluid and blood plasma, ACS Appl. Mater. Interfaces **6**, 9897 (2014).
- [58] W. Gao, R. Dong, S. Thamphiwatana, J. Li, W. Gao, L. Zhang, and J. Wang, Artificial micromotors in the mouse's stomach: A step toward *in vivo* use of synthetic motors, ACS Nano 9, 117 (2015).
- [59] Y. Gao and Y. Yu, How half-coated Janus particles enter cells, J. Am. Chem. Soc. 135, 19091 (2013).