Experimental assessment of the contribution of electrodynamic interactions to long-distance recruitment of biomolecular partners: Theoretical basis

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Highly specific spatiotemporal interactions between cognate molecular partners essentially sustain all biochemical transactions in living matter. That such an exquisite level of accuracy may result from encountering forces solely driven by thermal diffusive processes is unlikely. Here we propose a yet unexplored strategy to experimentally tackle the long-standing question of a possibly active recruitment at a distance of cognate partners of biomolecular reactions via the action of resonant electrodynamic interactions. We considered two simplified models for a preliminary feasibility investigation of the devised methodology. By taking advantage of advanced experimental techniques nowadays available, we propose to measure the characteristic encounter time scales of dually interacting biopartners and to compare them with theoretical predictions worked out in both the presence and absence of putative long-range electromagnetic forces.

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I. INTRODUCTION

Living matter hosts a huge number of molecular players (e.g., proteins, nucleic acids) involved in simultaneous yet specific chemical reactions, despite an apparent lack of systematic spatial order. A phenomenological description of these biomolecular machineries at work often makes use of the concept of "recruitment," leaving usually unclear how biomolecules partners encounter or move toward their specific targets and sites of action. On this point significative progress has been made about DNA-protein interaction at a short distance. This followed the puzzling problem posed by the E. coli lac repressor-operator protein, which was found to locate its specific DNA-binding site several orders of magnitude faster than the upper limit estimated for a diffusioncontrolled process [1,2]. A widely accepted approach to tackle this problem is the so-called facilitated diffusion, on which a vast literature exists (see, for instance, Refs. [3-6] and citations therein). On the other hand, for DNA-protein interactions and, more generally, for any dually interacting biomolecules, the mutual approach from a long distance is called three-dimensional (3D) bulk diffusion and has not been further studied (by "long distance" is meant: much larger than the Debye screening length). Actually, at first inspection, the mutual approach of cognate partners might well be driven by Brownian motion only, as at living temperatures the ubiquitously distributed water molecules move chaotically in space, colliding with larger or heavier fluid components. For the latter, the total outcome of many simultaneous hits are forces of both random intensity and direction. Hence, by

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displacing themselves in a diffusive way through the inner cellular space, large molecules sooner or later will encounter their targets.

A complementary proposal, which remains hitherto largely unexplored, is the possibility for molecules to interact at a distance via an electromagnetic field [7,8]. In particular, electromagnetic attractive forces acting on a long range might, in specific conditions, facilitate the encounters of cognate partners, so that specific biomolecular reactions would occur more effectively than if dependent on stochastic motion only. Exploring this possibility, it should be stressed that the static dielectric constant ε_s of water is particularly high, $\varepsilon_s \simeq 80$, at physiological temperatures. In addition to this dielectric screening, freely moving ions in the cellular medium tend to make the environment electrically neutral; accordingly, the Debye length in a biological environment is found to be smaller than $\simeq 10$ Å, as was estimated on the basis of typical ionic strength of the cytosol [6,9]. Electrostatic interactions between electrically charged molecules at a distance larger than the Debye length are very unlikely. Conversely this is not necessarily the case for electrodynamic interactions [10,11] since the dielectric constant depends on the frequency of the electric waves under consideration. Among the latter, the interactions occurring between oscillating electric dipoles are of a particular interest since in many cases the long-range nature of the interaction potential is essentially "activated" by the proximity of the dipole frequencies (resonance). In other words, two molecules whose dipole moments oscillate at the same frequency may undergo a so-called resonant interaction [12,13], which is described by the potential $U(r) \propto -1/r^3$ with r the intermolecular distance (see Appendix). On the other hand, an off-resonance situation would produce a standard van der Waals-like potential, i.e., $U(r) \propto -1/r^6$, typically a shortrange interaction (see Appendix). Such a frequency-selective interaction, when applied to a biological context, might be of utmost relevance during the approach of a molecule toward its

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specific cognate partner(s). To the best of our knowledge this proposition dates back to Jordan, who advanced the idea that resonant interactions within a quantum framework could play a significant role in autocatalytic reactions or influence the process of biological synthesis in such a way that replicas of molecules present in the cell are formed [14]. His theory was questioned by Pauling [15], who estimated that such forces, supposed to occur only between identical molecules, could not be large enough to cause a specific attraction between proteins under the thermal conditions of excitation and perturbation prevailing in living organisms. Other attempts to explain biological selectivity have been made later on the basis of usual van der Waals forces [16,17]. In parallel, in 1968 H. Fröhlich proposed a dynamical model [18] to account for the capacity of biological systems to self-regulate, emphasizing that, under specific conditions of energy supply to these systems, part of this supply would not be totally thermalized but would be used to create order in response to environmental perturbations [19]. In particular, the normal polarization modes of a macromolecule (or of a part of it) may undergo a condensation phenomenon, characterized by the emerging of the mode of lowest frequency containing nearly all the energy supply [18]. Then, relying on this model, Fröhlich suggested [20-22] that, when occurring between two biomolecules, such dipole oscillations could be excited enough to overcome thermal noise leading to the above mentioned frequency-dependent forces. Fröhlich's seminal work has stimulated many theoretical investigations until the present day (see, for example, Refs. [8,23–25]). Moreover, a vast literature is available about the experimental observation of low-frequency modes in the Raman and far-infrared (THz) spectra of proteins [26] and DNA [27]. These spectral features are attributed to collective oscillation modes of the whole molecule (protein or DNA) or of a substantial fraction of its atoms. A priori these collective oscillations of the molecular electric dipole moment could activate the mentioned long-distance attractive and selective recruitment interactions. However, a clear-cut experimental confirmation of the existence of the latter ones within a biological context at the molecular level is still lacking.

In the present paper we consider a yet unexplored strategy to experimentally test, at least in simplified systems, whether these long-range recruitment forces are actually at work between typical actors of the broad variety of biomolecular reactions in living matter. On the basis of theoretical computations resorting to elementary and standard methods in the theory of stochastic processes on the one hand, and recent progress on experimental methods on the other hand, we make a first step toward the design of experiments to test whether such forces are actually at work in living matter.

In Sec. II we use two dynamical models to highlight qualitative and quantitative changes between Brownian and non-Brownian encounters of the macromolecular partners of a generic biochemical reaction. In Sec. III we apply our models to the case of attractive electrodynamic potential $U(r) \propto -1/r^3$ expected to have effects at long distance, and then we report the numerical results that have been obtained with realistic parameters. Finally, in Sec. IV we discuss how our findings can be used to design an experiment, and we conclude that fluorescence cross-correlation spectroscopy (FCCS) is an appropriate experimental tool to perform real-time measurements of the association kinetics of dually interacting biopartners. It thus seems experimentally feasible to address the issues formulated above by comparing the outcomes of the prospected experiments versus the theoretically predicted curves at different concentrations of the reactants.

II. FIRST-PASSAGE TIME MODELS

A. Generalities

Our idea is in principle a natural one: Different kinds of forces must have different dynamical effects. Thus we attempted to devise an experimental protocol in order to discriminate between the dynamics of purely random encounters between reaction partners versus encounters driven by both a stochastic force plus a deterministic long-range force. Then, by experiments resorting to available techniques, we wondered whether it could be possible to discriminate between these different dynamical regimes.

A natural way to proceed from the theoretical standpoint that may be closely related to experimental as well as physiological conditions is to consider an aqueous environment, initially containing N_A particles of a species A and N_B particles of a species B. Each molecule A is expected to interact with each molecule B in two ways:

(1) As soon as the distance between A and B diminishes below a threshold δ , a biochemical reaction instantaneously takes place, so that the two molecules are not functional anymore and are considered as out of the system.

(2) The particle A and the particle B interact at a distance via a two-body potential U(r) of an electrodynamic type, as long as the two molecules do not get closer than the distance δ .

From a general point of view, the equations describing the dynamics of the system include both random and deterministic forces, and therefore may be given in the form

$$m_{A}\frac{d^{2}\mathbf{r}_{A,i}}{dt^{2}} = -\gamma_{A}\frac{d\mathbf{r}_{A,i}}{dt} - \sum_{j=1}^{N_{B}}\nabla_{A}U(|\mathbf{r}_{A,i} - \mathbf{r}_{B,j}|) + \sqrt{2\gamma_{A}kT}\boldsymbol{\xi}_{A,i}(t),$$

$$m_{B}\frac{d^{2}\mathbf{r}_{B,j}}{dt^{2}} = -\gamma_{B}\frac{d\mathbf{r}_{B,j}}{dt} - \sum_{i=1}^{N_{A}}\nabla_{B}U(|\mathbf{r}_{A,i} - \mathbf{r}_{B,j}|) \qquad (1) + \sqrt{2\gamma_{B}kT}\boldsymbol{\xi}_{B,j}(t),$$

$$i = 1, \dots, N_{A}, \text{ and } j = 1, \dots, N_{B}.$$

Here m_A , m_B correspond to the masses, $\mathbf{r}_{A,i}$, $\mathbf{r}_{B,j}$ to the positions, and γ_A , γ_B to the friction coefficients of the constituents of each species. *T* stands for the temperature in the solution, and *k* is the Boltzmann constant. $\boldsymbol{\xi}(t)$ is the random process modeling the fluctuating force due to the collisions with water molecules, usually represented as a Gaussian white-noise process for which $\langle \xi_{A,i}^{\alpha}(t)\xi_{A,k}^{\beta}(t')\rangle = \delta_{\alpha\beta}\delta_{ik}\delta(t-t')$, where $\alpha,\beta = 1, 2, 3$ are related to each component of $\boldsymbol{\xi}_{A,i}$. The same relation is valid also for $\boldsymbol{\xi}_{B,j}$.

A priori, Eqs. (1) describe a very complex dynamics, even in the absence of randomness. For example, assuming that the potential is, for each pair of molecules, of the form $U(r) = c_1/r^m - c_2/r^n$, with c_1, c_2 constants, $m, n \in \mathbb{N}$, m > n and $n \le d$ (long-range condition if *d* is the spatial dimension), the Hamiltonian subset of this system is actually a nonlinear classical *N*-body system whose phase space is entirely filled with chaotic trajectories [28]. At this stage, the addition of random forces may imply that the representative point of the system nontrivially wanders in phase space, despite the presence of dissipative terms, which, in principle, would generate trivial attractors. Indeed, in the overdamped limit, when the acceleration terms can be neglected, one is dealing with a randomly perturbed first-order nonlinear dynamical system which, as integrability is exceptional, is expected to display a complex (chaotic) dynamics. Nevertheless, instead of undertaking the numerical integration of Eqs. (1), we decided to look, as a first step, for some analytic result that can be obtained at the cost of simplification of the system.

Because the reaction between two particles A and B occurs the first time they come sufficiently close together, we will have to focus on first passage times of a simplified version of system (1). Generally, first passage or first return time statistics are difficult to examine in dynamical systems, then leading one to model the system under study by keeping only its salient characteristics, in either a deterministic or stochastic manner [29,30]. Here we choose to still work with Eqs. (1), but to reduce drastically the dimensionality of the system, then leading us to keep in the model under study its salient characteristics only. This was achieved by noting that Eqs. (1) describe the mutual interaction between the two sets of particles, A and B, but neither the A nor the *B* particles interact among themselves. The trajectories of the A particles are indirectly coupled only through the dynamics of the B particles. Thus, as a first simplifying hypothesis, we assumed that the B particles are fixed, and as a consequence the dynamical behaviors of the different A particles are independent.

This leads to the decoupling of the individual equations in (1) and hence to the introduction of a one-dimensional model representative of the generic dynamics of a single *A* particle. This model is considered below according to two different versions and solved according to standard methods [31].

B. Model 1: Absorbing plus reflecting boundaries

Let us consider one fixed molecule *B* located at the position z = 0 and one molecule *A*, initially located at z = x (see Fig. 1). We first suppose that if *A* reaches the boundary z = L of the domain, it is reflected back to z < L; whereas when *A* reaches the position $z = \delta$ for the first time, it is absorbed. The random trajectory z(t) of the molecule *A* may be given,



FIG. 1. (Color online) A generic initial condition of Model 1 (t = 0). Here x is the initial distance between A and B; δ is the distance at which A and B react, and L is the position of the reflecting barrier for A and the position of B is fixed.

as previously, in the form

$$\frac{dz}{dt} = v,$$

$$m\frac{dv}{dt} = -\gamma v + F(z) + \sqrt{2\gamma kT} \xi(t).$$
(2)

For times much larger than the characteristic time m/γ , Eqs. (2) will then relax to a state in which $dv/dt \rightarrow 0$. This approximation is justified by the fact that the biomolecules involved in reactions of interest (protein-protein or DNA-protein) typically weigh thousands of Daltons, and thus the characteristic relaxation times in aqueous medium are very short. Therefore, Eqs. (2) for the *A* particle can be simplified as

$$\frac{dz}{dt} = \frac{F(z)}{\gamma} + \sqrt{\frac{2kT}{\gamma}}\,\xi(t).$$
(3)

As is well known, the one-dimensional Langevin initial value problem [31]

$$\frac{dz}{dt} = a(z,t) + b(z,t)\xi(t), \quad z(t_0) = x,$$

is equivalent to the Fokker-Planck equation (FPE) for a probability $p(z,t|x,t_0)$ of finding the particle at z at time t, given it was at x at $t_0 \le t$

$$\frac{\partial}{\partial t}p(z,t|x,t_0) = -\frac{\partial}{\partial z}[a(z,t)p(z,t|x,t_0)] + \frac{1}{2}\frac{\partial^2}{\partial z^2}[b(z,t)^2p(z,t|x,t_0)].$$
(4)

From (3), one thus obtains

$$\frac{\partial}{\partial t}p(z,t|x,t_0) = -\frac{1}{\gamma}\frac{\partial}{\partial z}\left[F(z)p(z,t|x,t_0)\right] + \frac{kT}{\gamma}\frac{\partial^2}{\partial z^2}p(z,t|x,t_0),$$
(5)

which is also known as the Smoluchovski equation.

We now look at the time \mathcal{T} at which the reaction between A and B occurs, that is, the first time when particle A reaches $z = \delta$. Since we are considering an absorbing barrier at $z = \delta$ and a reflecting barrier at z = L, the probability $\mathbb{P}(\mathcal{T} \ge t)$ and the one that the particle would still be in the interval $[\delta, L]$ at time t are the same:

$$\mathbb{P}(\mathcal{T} \ge t) = \int_{\delta}^{L} dz \ p(z,t|x,0) := G(x,t).$$

In addition, as F(z) and kT do not explicitly depend on t, p(z,t|x,0), and thus G(x,t), are homogeneous processes, such that

$$G(x,t) = \int_{\delta}^{L} dz \, p(z,0|x,-t),$$
(6)

this implies that G(x,t) satisfies the same partial differential equation of p(z,0|x, -t) for z fixed, that is, a backward

Fokker-Planck equation

$$\frac{\partial}{\partial (-t)} p(z,0|x,-t) = -\frac{1}{\gamma} F(x) \frac{\partial}{\partial x} p(z,0|x,-t) - \frac{kT}{\gamma} \frac{\partial^2}{\partial x^2} p(z,0|x,-t),$$

leading to

$$\frac{\partial}{\partial t}G(x,t) = \frac{1}{\gamma} \left\{ F(x)\frac{\partial}{\partial x}G(x,t) + kT\frac{\partial^2}{\partial x^2}G(x,t) \right\}.$$
 (7)

Here the initial condition $p(z,0|x,0) = \delta(x-z)$ (here δ is the Dirac functional) clearly gives

$$G(x,0) = 1$$
, if $\delta \leq x \leq L$; and $G(x,0) = 0$, if not,
(8)

whereas the absorbing condition at δ and the reflecting boundary condition at *L* allow us to write, respectively,

$$G(\delta,t) = 0$$
 and $\frac{\partial}{\partial x}G(x,t)\Big|_{x=L} = 0, \quad \forall t > 0.$ (9)

If one focuses on the mean first-passage time $\tau(x)$, which represents a characteristic time scale of the reaction, one has by definition

$$\tau(x) = \int_0^\infty t \frac{\partial}{\partial t} \mathbb{P}(\mathcal{T} < t) dt$$
$$= -\int_0^\infty t \frac{\partial}{\partial t} G(x, t) dt = \int_0^\infty G(x, t) dt, \quad (10)$$

after integration by parts. Then, by integrating Eq. (7) between t = 0 and $t = \infty$, and using the fact that G(x,0) = 1 and $G(x,\infty) = 0$, we find that $\tau(x)$ must satisfy the following ordinary differential equation:

$$-1 = \frac{1}{\gamma} \left\{ F(x) \frac{d\tau(x)}{dx} + kT \frac{d^2 \tau(x)}{dx^2} \right\}$$

with boundary conditions $\tau(\delta) = \partial \tau(x)/\partial x|_{x=L} = 0$, as follows from Eqs. (9) and (10). The solution is found to be [31]

$$\tau(x) = \int_{\delta}^{x} dy \, \frac{1}{\psi(y)} \int_{y}^{L} dz \, \frac{\gamma}{kT} \psi(z),$$

with

$$\psi(x) = \exp\left[\int_{\delta}^{x} \frac{F(s)}{kT} ds\right] = \exp\left\{-\frac{U(x) - U(\delta)}{kT}\right\}, \quad (11)$$

since $F(x) = -\partial U(x)/\partial x$. This gives for $\tau(x)$:

$$\tau(x) = \frac{\gamma}{kT} \int_{\delta}^{x} dy \exp\left[\frac{U(y)}{kT}\right] \int_{y}^{L} dz \exp\left[-\frac{U(z)}{kT}\right].$$
(12)

It can easily be checked that the mean first-passage time in the presence of an attracting deterministic potential, generically written as $U(x) \propto -x^{-n}$ with a given n > 0, is smaller than



FIG. 2. (Color online) A generic initial condition of Model 2 (t = 0). Two molecules *B* are fixed at the boundaries x = 0 and x = l; *x* and l - x are the initial distances between *A* and the *B*'s and δ is the distance at which *A* and *B* react.

the mean first-passage time with Brownian motion only, i.e., when U = 0. Since $\exp(x^{-n})$ is a decreasing function of x, we can find an upper limit for the second integral and thus get

$$\tau(x) < \frac{\gamma}{kT} \int_{\delta}^{x} dy \exp\left(-\frac{1}{kT y^{n}}\right) \int_{y}^{L} dz \exp\left(\frac{1}{kT y^{n}}\right)$$
$$= \frac{\gamma}{kT} \int_{\delta}^{x} dy \int_{y}^{L} dz := \tau(x)^{\text{Bwn}}.$$

More explicitly,

$$\pi(x)^{\text{Bwn}} = \frac{\gamma}{2kT} [(L-\delta)^2 - (L-x)^2]$$
$$= \frac{\gamma}{2kT} (x-\delta)(2L-\delta-x).$$
(13)

C. Model 2: Two absorbing boundaries

Let us now consider the alternative model where two particles *B* are fixed at positions z = 0 and z = l, so that the particle *A*, initially located at z = x (see Fig. 2), is absorbed as soon as it reaches $z = \delta$ or $z = l - \delta$. Such a model is mathematically similar to the previous one.

Simply the deterministic force and the boundary conditions have to be modified. Consequently Eq. (7) is to be replaced by

$$\frac{\partial}{\partial t}G(x,t) = \frac{1}{\gamma} \left\{ \left[F(x) - F(l-x) \right] \frac{\partial}{\partial x}G(x,t) + kT \frac{\partial^2}{\partial x^2}G(x,t) \right\};$$
(14)

now G(x,t) is defined as $G(x,t) = \int_{\delta}^{1-\delta} dz \ p(z,t|x,0)$. The initial condition $p(z,0|x,0) = \delta(x-z)$ gives

$$G(x,0) = 1$$
, if $\delta \leq x \leq l - \delta$ and $G(x,0) = 0$, if not,
(15)

and the absorbing boundary conditions give

$$G(\delta,t) = 0$$
 and $G(l - \delta,t) = 0$, $\forall t > 0$. (16)

The mean first-passage time $\tau(x)$ defined above then satisfies the ordinary differential equation

$$-1 = \frac{1}{\gamma} \left\{ \left[F(x) - F(l-x) \right] \frac{d\tau(x)}{dx} + kT \frac{d^2 \tau(x)}{dx^2} \right\},$$

with boundary conditions

$$\tau(\delta) = \tau(l - \delta) = 0$$

$$\tau(x) = \left\{ \int_{\delta}^{l-\delta} dy \, \frac{1}{\psi(y)} \right\}^{-1} \\ \times \left\{ \int_{\delta}^{x} dy \, \frac{1}{\psi(y)} \int_{x}^{l-\delta} dw \, \frac{1}{\psi(w)} \int_{\delta}^{w} dz \, \frac{\gamma}{kT} \psi(z) \\ - \int_{x}^{l-\delta} dy \, \frac{1}{\psi(y)} \int_{\delta}^{x} dw \, \frac{1}{\psi(w)} \int_{\delta}^{w} dz \, \frac{\gamma}{kT} \psi(z) \right\}$$

with

$$\psi(x) = \exp\left\{\int_{\delta}^{x} \frac{F(s) - F(l-s)}{kT} ds\right\}$$
$$= \exp\left\{-\frac{U(x) + U(l-x) - U(\delta) - U(l-\delta)}{kT}\right\} (17)$$

since $F(x) = -\partial U(x)/\partial x$. After simplification, one has

$$\tau(x) = \frac{\gamma}{kT} \left\{ \int_{\delta}^{l-\delta} dy \,\phi(y) \right\}^{-1} \\ \times \int_{\delta}^{x} dy \int_{x}^{l-\delta} dw \int_{y}^{w} dz \frac{\phi(y)\phi(w)}{\phi(z)}$$
(18)

where

$$\phi(s) = \exp\left\{\frac{U(s) + U(l-s)}{kT}\right\} \ .$$

Similarly to Model 1, the expression for the mean reaction time with Brownian motion only (U = 0), is particularly simple:

$$\tau(x)^{\text{Bwn}} = \frac{\gamma}{2kT} (x - \delta)(l - \delta - x).$$
(19)

To summarize, in this section we have obtained the general form of the mean first-passage time $\tau(x)$, that is the average time needed by molecule *A* to reach the molecule *B* (or one molecule *B* in the case of Model 2), as a function of the initial intermolecular distance *x* and temperature *T*, for both Model 1 [Eq. (12)] and Model 2 [Eq. (18)], respectively. The same function is given for randomly driven encounters between the reaction partners in Eq. (13) for Model 1 and in Eq. (19) for Model 2.

III. QUANTITATIVE THEORETICAL PREDICTIONS

In order to answer the question of whether it would be feasible to *experimentally detect* the possible existence of a deterministic attractive force through which the cognate partners of biochemical reactions interact at long distance, we first have to delimit the physical context, choose the domain of physical parameters, and provide the analytic form of the two-body interaction potential. In what follows, a long-range resonant potential potential $U(x) = -C/x^3$ is considered. As discussed in the Introduction, this kind of interaction can have sizable effects at long distances at variance with London–Van der Waals $1/x^6$ interactions (see also Appendix). Following Fröhlich [22], a lower bound for the coefficient of this potential is given by $C \simeq \hbar e^2 (Z_A Z_B)^{1/2} / 2M\omega_0 \varepsilon'(\omega_0)$, where Z_A and Z_B denote the number of charges of averaged mass M and charge e contributing to the dipole moment of the molecules A and B, respectively, whereas ω_0 stands for their oscillation frequency; $\varepsilon'(\omega_0)$ is the real part of the dielectric constant of the interposed medium. In particular, it is interesting to remark that in the expected range of oscillation frequencies for the setup of collective dipole oscillations in macromolecules (that Fröhlich estimated to be $\omega_0 \simeq 10^{11}-10^{12}$ Hz) the value of $\varepsilon'(\omega_0)$ drops down to a few units [32], thus allowing a much smaller screening of the interactions with respect to the static case. In this context, we use $Z_A = Z_B = 1000$ (see Ref. [8]) and the proton mass for *M*. A convenient unit system remains to be chosen. For the numerical tabulations of $\tau(x)$, instead of c.g.s. units we use μ m, kDa, and μ s, with the following definitions: 1 μ m =10⁻⁴ cm, 1 kDa $\approx 10^{-21}$ g, and 1 μ s = 10^{-6} s.

In this system of units we evaluate the lower bound of C, which is found to be $C \simeq 10^{-30}$ erg cm³ = 0.1 kDa μ m⁵ μ s⁻². Henceforth, we shall consider C varying from 0.1 to 10 kDa μ m⁵ μ s⁻². These values are given with a degree of arbitrariness that can be reduced by considering that C = 10corresponds to the physical situation where $U(x) \simeq kT$ at $x \sim 0.1 \ \mu m$. Hence the choice $\mathcal{C} \in [0.1, 10]$ is a very cautious estimate with respect to those existing in the literature about a possibly larger range of action [it has been surmised by Fröhlich and others that U(x) might become comparable with kT at $x \sim 1 \ \mu m$ or more [8,23,24]]. Among other constants appearing in Eqs. (12) and (18), the friction coefficient γ of the molecule A has been estimated according to Stokes' law $\gamma = 6\pi \eta(T)\mathcal{R}$, where $\eta(T)$ corresponds to the viscosity of water at temperature T and \mathcal{R} stands for the hydrodynamic radius of the molecule. The value of \mathcal{R} has been set equal to $5 \times 10^{-3} \,\mu\text{m}$, which is the typical diameter of a biomolecule with a mass in the interval 50-100 kDa (proteins and DNA fragments); the same value has been fixed for the reaction radius δ introduced in both models: $\mathcal{R} = \delta = 5 \times 10^{-3} \ \mu \text{m}.$

All the computations of $\tau(x)$ have been performed by means of MATLAB programs. Also, as MATLAB does not allow us to perform direct integrations over nonrectangular domains, integrals with variable limits in Eqs. (12) and (18) have been first "vectorized" [33] for each x to calculate $\tau(x)$ with a recursive adaptive Simpson quadrature (MATLAB quadl function). Further checks on the reliability of the method have been done through direct numerical integration of the Langevin equation (3) by means of a standard Euler-Heun algorithm and by averaging over 10⁴ different realizations of the random walk. Minor precision problems, especially when $x \sim \delta$, have been thus detected and corrected in what follows.

A. Model 1

We have computed $\tau(x)$ and $\tau(x)^{Bwn}$ by means of Eqs. (12) and (13), respectively, where we have set $R = \delta = 5 \times 10^{-3} \mu \text{m}$ and $U(x) = -C/x^3$, as detailed above. The position L of the reflecting barrier characteristic of Model 1 has been fixed so as $x \ll L$ for all x. In particular, $L = 10 \mu \text{m}$, and a maximal value for x equal to 1μ m have been used. Figure 3 displays the numerically found shapes of both functions $\tau(x)$ and $\tau(x)^{Bwn}$ computed at T = 300 K and for different values of the attractive potential coefficient C. A first check on the reliability of the plotted results is done by observing that $\tau(x) < \tau(x)^{Bwn}$ for all the x values while both curves merge at large x values, as expected when the resonant attraction



FIG. 3. (Color online) Model 1: Mean encounter time τ between two molecules *A* and *B*, initially placed at a distance *x* one from the other. Dotted lines are asymptotic behaviors. Dashed line refers to purely random encounters. Solid line refers to the combined effect of a random force plus a deterministic one derived from the potential $U(x) = -C/x^3$. Panel (a) refers to C = 0.1. Panel (b) refers to C = 10.

is wiped out by thermal noise. In particular, as we always considered $x \ll L$, the asymptotic behavior of $\tau(x)^{\text{Bwn}}$ is then proportional to x as required by Eq. (13). On the contrary, at smaller x, $\tau(x)^{Bwn}$ bends downward to slightly smaller values with respect to the extrapolated linear dependence (this happens when x is no longer much larger than δ). At variance, the pattern of $\tau(x)$ has two asymptotic limiting behaviors: At large x values it joins the Brownian curve $\tau(x)^{Bwn}$, and at small x values it is $\tau(x) \sim x^5$ (a power law characteristic of the $1/x^3$ form of the potential); the latter might be anticipated on the basis of simple dimensional arguments since the left-hand side of Eq. (3) has the dimensions $[dz/dt] = lt^{-1}$ while the right-hand side leads to $[\mathcal{C}/x^4] = [\mathcal{C}]l^{-4}$ in a purely deterministic regime. By combining the two, for a generic time scale $[\tau] = t$ associated with a displacement length x we get $\tau \sim [x]^5$ [34]. The two limiting behaviors are bridged by a steep transition pattern, which moves rightward or leftward according to the value of C, as shown in Fig. 3; the stronger the potential the larger the x values at which $\tau(x)$ displays the knee joining the x^5 functional dependence. The transition pattern of $\tau(x)$ is steep since the reflecting barrier is located far from the only molecule *B*.



FIG. 4. (Color) Model 1: Temperature dependence of $\tau(x)$ for C = 1.0. Red solid and dashed curves refer to T = 200 K, green dashed and solid curves refer to T = 300 K, blue dashed and solid curves refer to T = 400 K.

In any case, $\tau(x)^{Bwn}$ is found to exceed $\tau(x)$ by a factor of, e.g., 10 at definitely larger x values and, what is more relevant, at longer values of first encounter time: With C = 0.1, such a difference occurs at $x \simeq 300$ Å where $\tau^{Bwn} \simeq 6$ ms and $\tau \simeq 600 \,\mu$ s, while with C = 1.0, it occurs at $x \simeq 640$ Å, where $\tau^{Bwn} \simeq 10$ ms and $\tau \simeq 1$ ms, and with C = 10 at $x \simeq 1400$ Å, where $\tau^{Bwn} \simeq 20$ ms and $\tau \simeq 2$ ms. As we shall see in the next section, should we interpret x as the average distance between any two reacting molecules in three dimensions, this range of x values (between a few hundred angstroms and 1 μ m) is easily attained by varying the concentrations of the reactants between a few micromoles down to one nanomole. Notably, the encounter times belong to an interval of values easily accessible by means of optical detection methods.

A priori, further qualitative indications on the possible presence of attractive deterministic forces between cognate partners could be observed by modifying the temperature of the system. As shown in Fig. 4, $\tau(x)$ and $\tau(x)^{Bwn}$ plotted for three different values of T confirm that the x^5 functional dependence is purely deterministic as T has no influence within this domain. On the contrary, $\tau(x)^{Bwn}$ displays the same dependence on T for all values of x. Surprisingly, the steep transition pattern of $\tau(x)$ at intermediate values of x is characterized by a temperature dependence which is inverted compared to the Brownian case: In the presence of an intermolecular potential, the higher the temperature, the larger the first-passage time of A at $x = \delta$. Finally note that the temperature range considered in Fig. 4 is a broad one (T = 200, 300, and400 K). Nevertheless, as the physiological temperature range corresponds only to a few percent around 300 K, it is likely that variations of the first passage time at different temperatures are too weak to be experimentally detectable within such an interval. In particular, computations performed for temperature differences of 10K (typically 290, 300, 310K) with the Model 1 show variations of τ less than 5% of its value in the Brownian case as well as in the case of Brownian plus deterministic force.



FIG. 5. (Color online) Model 2: Mean encounter time τ between two molecules *A* and *B*, initially placed at a distance *x* from each other. Dotted lines are asymptotic behaviors. Dashed line refers to purely random encounters. Solid line refers to the combined effect of a random force plus a deterministic one derived from the potential $U(x) = -C/x^3$. Panel (a) refers to C = 0.1. Panel (b) refers to C = 10.

B. Model 2

We have also plotted $\tau(x)$ and $\tau(x)^{Bwn}$ computed according to Eqs. (18) and (19), respectively, where $R = \delta = 5 \times 10^{-3} \mu \text{m}$, l = 2x, and $U(x) = -C/x^3$ as in the case of Model 1. Figure 5 displays the numerically found shapes of both functions $\tau(x)$ and $\tau(x)^{Bwn}$ computed again at T = 300 K and for different values of the attractive potential coefficient C.

At first check, the characteristics of $\tau(x)$ prevailing for Model 1 [i.e., the x^5 functional dependence at small x, and the tendency of $\tau(x)$ to join $\tau(x)^{Bwn}$ at large x] also apply here. Nevertheless, asymptotic x dependence of $\tau(x)^{Bwn}$ is now proportional to x^2 as obtained from Eq. (19) by replacing l by 2x when $x \gg \delta$. On the other hand, the absence of a reflecting barrier makes the steep transition feature for $\tau(x)$ disappear and be replaced by a mild crossover at intermediate values of x. This steeper pattern for the first passage time as a function of x, in the case of purely Brownian diffusion, entails a less pronounced separation between $\tau(x)$ and $\tau(x)^{Bwn}$. In fact, a separation between these two curves by a factor of 10 (to make the same kind of comparison that we did for Model 1) occurs for C = 0.1 at $x \simeq 200$ Å for which $\tau^{Bwn} \simeq 1.1 \ \mu s$ and $\tau \simeq 0.1 \ \mu s$; while for C = 1.0 this happens at $x \simeq 600$ Å



FIG. 6. (Color) Model 2: Temperature dependence of $\tau(x)$ for C = 0.1. Red solid and dashed curves refer to T = 200 K, green dashed and solid curves refer to T = 300 K, blue dashed and solid curves refer to T = 400 K.

for which $\tau^{Bwn} \simeq 30 \,\mu s$ and $\tau \simeq 3 \,\mu s$; and, finally, for C = 10 at $x \simeq 1170$ Å for which $\tau^{Bwn} \simeq 120 \,\mu s$ and $\tau \simeq 12 \,\mu s$. Therefore, we can see that Model 2 is more constraining than Model 1, in the sense that, at equal values of C (that is, at equal strength of the long-range interaction), smaller intermolecular distances and a much faster tracking of the dynamics of the reactants are needed to discriminate with the same degree of confidence (arbitrarily set as a factor of 10) between random and nonrandom encounters of the reactant molecules.

The temperature dependence of both $\tau(x)^{\text{Bwn}}$ and $\tau(x)$ is reported in Fig. 6. The main features of $\tau(x)$ in Model 1 are likewise present in Model 2. In addition, we can see that the inversion in the temperature dependence with respect to the Brownian case (which was characteristic of the steep transition pattern of Model 1) is no longer there. Thus, the peculiar temperature dependence of this steep transition pattern could be mostly attributed to the presence of the reflecting barrier characteristic of Model 1. Likewise in Model 1, we used again T = 200, 300, and 400 K, even though computations carried out at physiological temperature again yield too weak variations of $\tau(x)$ to be experimentally detectable.

IV. DISCUSSION AND CONCLUDING REMARKS

The numerical results reported in the preceding section are in favor of a positive answer to the main question addressed by the present work. In fact, the numerical study of Models 1 and 2 revealed qualitative differences in the mean first-passage time τ between the case of a pure Brownian diffusion of the molecule A (see Sec. II) and the case in which an attractive (resonant) potential U is added to a random force. In particular, in the latter case, the functional dependence of τ on the initial distance x between the molecule A and its target [molecule(s) B] demonstrates the existence of different patterns in the two models depending on the range of the x values considered:

(1) A deterministic pattern at small x values (small initial separations), characterized by a power law representative of the potential under consideration (x^5 for the resonant potential used, x^{p+2} for a general potential of the form $U(x) \propto x^{-p}$)

(2) A Brownian pattern at large initial separations, proportional to x or to x^2 depending upon the symmetry of the system (x for an asymmetric situation as described by Model 1, and x^2 for a symmetric one as described by Model 2)

(3) A steep transition pattern joining the two asymptotic ones in the case of Model 1, and a smooth crossover joining the two asymptotic ones in the case of Model 2.

Although complementary computations revealed some interesting features in the temperature dependence of τ , the corresponding degree of variation in a laboratory accessible interval of temperature is too weak to be experimentally detectable.

In any case it is obvious that x must constitute an experimentally accessible control parameter so that the results mentioned above may be used to predict the possible role of long-range intermolecular forces in biological processes. Notably, such an approach is not very usual. Indeed, most of the attempts made hitherto in this direction have resulted in experimental measurements of association constants k_a (characteristic of a reaction medium), which are predictable from the Smoluchowski theory also when intermolecular forces are considered [35,36]. The focus of Smoluchovski theory is on the association constant k_a , which represents the probability for two molecules to react per time unit, irrespective of their position. In the case of Brownian encounters, this is given by [4] $k_a^B = 4\pi RD \equiv 4\pi \delta kT/\gamma$, where R is the reaction radius that can be approximated to δ from the current study, and D is the sum of the diffusion coefficients of the two cognate partners. In the presence of some interaction potential U, one has $k_a^* = 4\pi R^* D$, where R has been replaced by [4,35,36] $R^* = R(\int_R^{\infty} r^{-2} e^{-U(r)/kT} dr)^{-1}$. Now, if k_a is experimentally measured for some reaction and it turns out that $k_a^B < k_a$, then this would indicate that some deterministic force is in action, but one can hardly determine the law of the interaction potential because after integration over r there is no one-to-one correspondence between R^* (thus k_a^*) and the functional form of U(r) [24]. On the other hand, in measuring $k_a < k_a^B$, we cannot be sure that the reaction is simply diffusion driven because, in this case, chemical times could be long enough to make k_a smaller than the corresponding Brownian value. The advantage of our dynamical approach is that our models still apply by choosing δ as the distance at which A and B get in contact without reacting, and with the experimental technique discussed below (FCCS) we can make a distinction between the association time and the chemical times.

In the present situation x values might to some extent be considered in three dimensions as the *average distance* between two molecular partners, A and B, while this quantity can be easily controlled in laboratory experiments by varying the concentrations of the reactants. Given the concentrations $C_A = N_A/V_A$ and $C_B = N_B/V_B$ (with $V_{A,B}$ = the initial volumes and $N_{A,B}$ = the number of molecules of the two species, respectively; note that these numbers are controlled through the molarity, i.e., a definite fraction of the Avogadro number), we get the estimate $x = C_{av}^{-1/3}$ for the average intermolecular distance from the average concentration $C_{av} = (N_A + N_B)/V$, where the reaction volume $V = V_A + V_B$. In practice, as an example, with $C_{av} = 1$ nM we have $x \simeq 1 \,\mu$ m as the average distance between any two molecules, while with $C_{av} = 1 \,\mu$ M we have $x \simeq 1000$ Å. By working at equimolarity, that is, $C_A = C_B$, then $C_{av}^{-1/3}$ is a good estimate of the average distance between one A and one B molecule. Working with nanomoles of DNA and proteins (enzymes, transcription factors) is quite standard in molecular biology experiments. With such concentrations of reactants, both models (1 and 2) predict that the first passage time, which can be interpreted as the average encounter time between one A and one Bmolecule initially located at intermolecular distances of a few thousands of angstroms, varies in the interval between a few tens of microseconds to about 1μ s in the presence of an attractive deterministic force that would sum up to the random force. On the contrary, in the very same conditions, random only driven encounters would exceed the above mentioned encounter times by one or two orders of magnitude. Again, the distance at which sizable differences could be observed may vary significantly depending on the actual value of the resonant potential parameter C. On the other hand, estimates in literature [8,23,24] suggest that these long-range resonance interactions could be effective up to distances in the order of 1 μ m (the action range is estimated by computing the distance at which the resonance interaction energy equals the level of thermal noise kT). In this respect, in the preceding section we have limited ourselves to cautious estimates for the parameter \mathcal{C} , focusing on conservative assumptions for average encounter time varying in the interval $10^{-5}-10^{-3}$ s, which can be readily detected with the aid of fluorescence cross-correlation spectroscopy (FCCS) technology. This is a powerful technique which is being increasingly applied to the study of diffusion and chemical reaction rates in complex biological systems using fluorescently labeled macromolecules [37–39]. FCCS measures the spontaneous fluctuations of fluorescences $\delta F_1(t)$ and $\delta F_2(t)$ that arise from the diffusion of fluorescently labeled molecules of type 1 and 2, respectively, illuminated by two laser light beams of different colors, into or out of an open sampling volume. Even though the size of the detection volume is diffraction limited, the autocorrelation functions of $\delta F_{1,2}(t)$ and the cross-correlation function $\langle \delta F_1(t) \delta F_2(t) \rangle$ can be altered by processes occurring on smaller spatial scales. These correlation functions provide information on diffusion properties of fluorescent molecules.

Of course, we are well aware of the fact that the models studied here are simplified descriptions of the reality. Indeed, protein-protein and protein-nucleic acid interactions in vivo generally take place within complex structural scaffolds such as the membrane cytoskeleton or the chromatin envelope, which are themselves the subject of highly dynamical regulations (e.g., Refs. [40,41]) and may also possibly interfere with the spatiotemporal control of the given reactions (e.g., Refs. [42,43]). Should resonant electrodynamic interactions be involved within such an intricate context, it seems illusory at this stage to assess realistic C values simply based on the proposed experiments. In fact, regarding protein-DNA in vivo (physiological) interactions, for instance, it may well be that the putative values fluctuate depending on a host of variables, possibly including, in a nonmutually exclusive way, charges on proteins and DNA, the effect of surrounding electrolytes, the nucleic- or amino-acid compositions, the length of accessible DNA, etc. However, we stress here that our initial goal, as described in this article, is to merely probe whether or not biological partners also can take advantage, besides thermic diffusion, of long-distance (0.1–1 μ m) forces of electrodynamic origin to eventually interact. If established, this novel concept would then in turn open new avenues of research to investigate long-standing biological issues, e.g., on the precise definition of which variables exactly pertain on protein-DNA interactions, and how a diffusing protein particle may actually recognize the particular cognate DNA site among many other locations also available. Since we do not expect dramatic qualitative changes out of the numerical simulations of Eqs. (1) in three dimensions [44], an experimental setup providing a practical realisation of what has been investigated in the present work could be devised by resorting, as experimental probes, to three broad classes of interactions: protein-DNA, protein-RNA, and protein-protein (ligand-receptor). As DNA and RNA molecules do not have a preassigned length, it is implicitly understood that only short fragments are to be considered (some tens or a few hundreds of base pairs, that is, oligonucleotides or plasmides, respectively). The proteins interacting with DNA or RNA can be processing enzymes (helicases, polymerases, recombinases) or transcription factors normally bound at promoters, enhancers, insulators, or silencers. Thus, for example, one could choose two molecular species consisting, respectively, of a short double-stranded DNA molecule (for example, a synthetic oligonucteotide of ~ 100 base pairs or even less) and a protein with a site-specific affinity for the chosen DNA molecule (i.e., a transcription factor). By labeling the DNA molecules and proteins with standard fluorophores their dynamical behavior can be followed by means of FCCS microscopy at different concentrations $C = C_A = C_B$ of the reactants to get a characteristic time scale as a function of $x = C^{-1/3}$. In this way such an experimental setup should provide, after data fitting, an estimate of the constant C for the resonant potential considered above. Thus, C = 0 would mean that the reactants meet only under the action of Brownian diffusion, whereas $\mathcal{C} \neq 0$ would prove the existence at the same time of the long-range interactions evoked throughout this paper and give quantitative information about them.

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APPENDIX

For the sake of clarity and to help the reader to get hold of the physical origin of the $U(R) \propto -1/R^3$ potential referred to throughout the present work, this Appendix provides theoretical elements about the interaction of oscillating electric dipoles.

To begin, let us recall basic facts about this subject. Two atoms (or two small molecules) A and B in their ground states with no net charge excess and vanishing average dipole moment (i.e., both are nonpolar) interact through the London-Van der Waals dispersive force. The origin of this interaction is as follows. Though the expectation values of the dipole operators are zero for nonpolar atoms, quantum fluctuations are responsible for their instantaneous nonvanishing dipolar moments. This entails a nonzero dispersion of the dipole moment operator. The energy of the two isolated atoms is corrected at first order by the dipole-dipole interaction energy, which is proportional to the average dipole moments; thus it vanishes when both atoms are in their ground states. Instead, the second-order perturbative correction, due to the coupling between instantaneous dipole fluctuations, is found to be proportional to $1/R^6$. (In a QED framework the London–Van der Waals interaction stems from the exchange of virtual photons between the atoms.) This is a short-range potential, so called because the exponent of the power law of Ris strictly larger than 3, the dimension of physical space. London-Van der Waals interactions are of generically weak intensity, whereas they likely become of prime importance in a biological context when acting at short distances (below the Debye length) together with additional interactions of chemical type [45].

Remarkably the first-order perturbative correction may be nonvanishing under a degeneracy condition. Indeed, if one or both atoms are in an excited state, provided that the condition for exchange symmetry is fulfilled, that is, they have common eigenenergies in their spectra, it can be shown [46] that the interaction energy is now proportional to $1/R^3$, a long-range potential.

Interactions of similar kinds to those just mentioned between two atoms (or small molecules) could exist between macromolecules with an oscillating electric dipole moment. In this case the oscillating dipole moment would not be due to the electron motions but, rather, to conformational vibrations. As already mentioned in the Introduction, this interaction between the oscillating electric dipole moments of reacting macromolecules could play a relevant role in living matter. In fact, as already recalled in the Introduction, the high static dielectric constant of water together with the considerable amount of ions present in living cells tend to screen any electrostatic interaction beyond a distance of a few angstroms. However, this electrostatic opaqueness does not hold for an oscillating field: The higher the frequency of an oscillating field, the more transparent an aqueous salted medium. In fact, the value of the dielectric constant of water at room temperature is a decreasing function of the frequency [32] and, for example, already at 1 THz $\varepsilon(\omega = 10^{12}) \simeq 4$; likewise, the imaginary (dissipative) part of the dielectric constant (which is proportional to the conductivity of the medium due to the presence of free ions) is inversely proportional to the frequency of the oscillating electric field (according to the Drude equation [48]), so that at suitably high frequency it can be negligible.

Let us now study the basic mechanism of interaction between two oscillating electric dipoles before discussing its application to biomolecules [47]. As we show below, these oscillating dipoles can activate long-range forces that will be shown to be frequency selective. We consider a onedimensional simplified model in which the dipoles oscillate at frequencies ω_A and ω_B , respectively. Then a computation of the interaction energy between *A* and *B* can be given, which, despite the simplified treatment, allows us to grasp basic physical facts.

Let μ_A and μ_B be the masses of the two oscillators, let their dipole moments be parallel and given by qZ_Ar_A and qZ_Br_B , and assume that their mutual separation *R* is such that $R \gg r_A, r_B$; then we can write the interaction Hamiltonian as

$$H = \frac{p_A^2}{2\mu_A} + \frac{p_B^2}{2\mu_B} + \frac{1}{2}\mu_A\omega_A^2 r_A^2 + \frac{1}{2}\mu_B\omega_B^2 r_B^2 + \frac{\zeta q^2 Z_A Z_B}{4\pi\varepsilon_0 R^3} r_A r_B, \qquad (A1)$$

where Z_i , i = A, B, stands for an effective number of charges which account for the average value of the dipole moment of the oscillator *i*; ζ is a geometrical factor depending on the orientation of the dipoles with respect to the line joining them (on which the distance *R* is measured). Then, introducing a mean mass *M* defined so that $\mu_A = MZ_A$ and $\mu_B = MZ_B$, the Hamiltonian becomes

$$H = \frac{1}{2M} \left(p_A^2 + p_B^2 \right) + \frac{1}{2} M \omega_A^2 r_A^2 + \frac{1}{2} M \omega_B^2 r_B^2 + \frac{\beta}{R^3} r_A r_B,$$
(A2)

where the transformations $(Z_i)^{1/2}r_i \rightarrow r_i$ and $p_i/(Z_i)^{1/2} \rightarrow p_i$, i = A, B, have been introduced (the variables r_i and p_i are still canonically conjugated), and we put $\beta = \zeta q^2 (Z_A Z_B)^{1/2} / 4\pi \varepsilon_0$, where ε_0 is the dielectric constant of vacuum, in the absence of a material medium between the

oscillators, to be replaced by $\varepsilon(\omega)$ when a medium is present. In matrix form this also reads

$$H = \frac{1}{2M} \left(p_A^2 + p_B^2 \right) + \frac{1}{2} M \left(r_A \ r_B \right) \underbrace{\begin{pmatrix} \omega_A^2 \ \beta/MR^3 \\ \beta/MR^3 \ \omega_B^2 \end{pmatrix}}_C \begin{pmatrix} r_A \\ r_B \end{pmatrix}.$$
(A3)

Matrix *C* is real and symmetric, and thus diagonalizable by means of an orthogonal transformation. Let ω_+^2 and ω_-^2 the eigenvalues of *C* (homogeneous to squared frequencies). Under the action of this transformation the Hamiltonian can be cast in the form of the sum of two decoupled oscillators, that is,

$$H = \frac{1}{2M}(p_{+}^{2} + p_{-}^{2}) + \frac{1}{2}M\omega_{+}^{2}r_{+}^{2} + \frac{1}{2}M\omega_{-}^{2}r_{-}^{2}, \quad (A4)$$

and it can be easily shown that

$$\omega_{\pm} = \frac{1}{\sqrt{2}} \bigg[\left(\omega_A^2 + \omega_B^2 \right) \pm \left\{ \left(\omega_A^2 - \omega_B^2 \right)^2 + \frac{4\beta^2}{M^2 R^6} \right\}^{1/2} \bigg]^{1/2}.$$
(A5)

By considering r_A , r_B , p_A , and p_B as observables subject to standard commutation relations, the energy values of the system are obviously given by

$$E = \hbar \omega_{+} \left(n_{+} + \frac{1}{2} \right) + \hbar \omega_{-} \left(n_{-} + \frac{1}{2} \right),$$
 (A6)

where $n_+, n_- \in \mathbb{N}$, that is, are integers. Now, let us consider two opposite physical situations depending on the relative values of the frequencies ω_A and ω_B of the oscillators.

(1) Consider $\omega_A \gg \omega_B$ (or, equivalently, $\omega_A \ll \omega_B$), we have

$$\omega_{\pm} = \frac{1}{\sqrt{2}} \left[\left(\omega_A^2 + \omega_B^2 \right) \pm \left(\omega_A^2 - \omega_B^2 \right) \left\{ 1 + \frac{4\beta^2}{\left(\omega_A^2 - \omega_B^2 \right)^2 M^2 R^6} \right\}^{1/2} \right]^{1/2}, \tag{A7}$$

and the denominator of the last term is large enough to give at the lowest-order expansion

$$\omega_{\pm} = \frac{1}{\sqrt{2}} \left[\left(\omega_A^2 + \omega_B^2 \right) \pm \left(\omega_A^2 - \omega_B^2 \right) \pm \frac{2\beta^2}{\left(\omega_A^2 - \omega_B^2 \right) M^2 R^6} + \cdots \right]^{1/2} \\ = \frac{1}{\sqrt{2}} \left[2\omega_{A,B}^2 \pm \frac{2\beta^2}{\left(\omega_A^2 - \omega_B^2 \right) M^2 R^6} + \cdots \right]^{1/2} = \omega_{A,B} \pm \frac{\beta^2}{2\omega_{A,B} \left(\omega_A^2 - \omega_B^2 \right) M^2 R^6} + \cdots,$$

where $\omega_{A,B}$ stands for ω_A in the computation of ω_+ and ω_B in the computation of ω_- . By substituting this expression in Eq. (A6) we get

$$E = \hbar\omega_A \left(n_+ + \frac{1}{2} \right) + \hbar\omega_B \left(n_- + \frac{1}{2} \right) + \frac{\hbar\beta^2}{2(\omega_A^2 - \omega_B^2)M^2R^6} \left\{ \frac{1}{\omega_A} \left(n_+ + \frac{1}{2} \right) - \frac{1}{\omega_B} \left(n_- + \frac{1}{2} \right) \right\} + \cdots$$
(A8)

The first two terms correspond to the unperturbed energies of the oscillators A and B considered as isolated $(R \rightarrow \infty)$, while the last term provides the lowest-order correction to the unperturbed energy of the system, and due to the interaction, this interaction potential energy is proportional to R^{-6} . Note that this is functionally the same as the London–Van der Waals interaction but of a remarkably different physical origin (real oscillations instead of quantum fluctuations).

(2) On the other hand, at resonance, that is, $\omega_A \simeq \omega_B = \omega$, the eigenfrequencies (A5) are simply given by

$$\omega_{\pm} = \omega \sqrt{1 \pm \frac{\beta}{M\omega^2 R^3}}.$$
 (A9)

At long distances (imposed by the reality condition for ω_{\pm} in this equation) we can develop ω_{\pm} near ω and replace such a development into Eq. (A6) to obtain

$$E = \hbar\omega(n_{+} + n_{-} + 1) + \frac{\hbar\beta}{2M\omega R^{3}}(n_{+} - n_{-}) - \frac{\hbar\beta^{2}}{8M^{2}\omega^{3}R^{6}}(n_{+} - n_{-} + 1) + \cdots$$
(A10)

The first-order correction to the energy of the system corresponds to the interaction energy between the two oscillators at resonance and is proportional to R^{-3} . If both oscillators are in their ground states, i.e., $n_+ = n_- = 0$, the first contribution to the interaction energy in Eq. (A10) vanishes as well as the force given above. The first nonvanishing term is again proportional PHYSICAL REVIEW E 85, 041904 (2012)

to R^{-6} . But if the lowest of these modes (ω_{-}) gets more excited than the other (ω_{+}), then the consequence is the activation of an *attractive* long-range frequency-selective force. A repulsive force could also be activated in case $n_{+} > n_{-}$.

In the context of Fröhlich's theory [8,18,49,50] the above described mechanism of resonant interaction between oscillating dipoles was surmised to have a great potential relevance for fundamental biological processes at the molecular level. Fröhlich proposed a model describing the coupling between the elastic vibrations of macromolecules and the resulting time variations of their dipole moment; the model predicts that one or a few Fourier modes of the dipole field oscillation should be strongly (coherently) excited provided that the energy supply rate exerted on the macromolecule by its environment exceeds a threshold value. This energy supply is assumed to depend on the biological activity of the environment (metabolic energy). The strongly excited mode of oscillation of the molecular dipole moment should be due to a collective oscillation either of the entire molecule or of a subgroup of its atoms. The consequence of such collective oscillations would be to activate selective long-range recognition and attraction between cognate macromolecular partners via the above described mechanism of resonant interaction. Experimental evidence of the existence of collective excitations in macromolecules of biological relevance is available for polynucleotides (DNA and RNA) [27] and for proteins [26] in the Raman and far-infrared (THz) spectroscopic domains.

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