

Uncertainty quantification and estimation in differential dynamic microscopyMengyang Gu^{1,*}, Yimin Luo^{2,3}, Yue He¹, Matthew E. Helgeson^{2,†} and Megan T. Valentine^{3,‡}¹*Department of Statistics and Applied Probability, University of California, Santa Barbara, California 93106, USA*²*Department of Chemical Engineering, University of California, Santa Barbara, California 93106, USA*³*Department of Mechanical Engineering, University of California, Santa Barbara, California 93106, USA*

(Received 2 May 2021; revised 31 August 2021; accepted 7 September 2021; published 24 September 2021; corrected 1 October 2021)

Differential dynamic microscopy (DDM) is a form of video image analysis that combines the sensitivity of scattering and the direct visualization benefits of microscopy. DDM is broadly useful in determining dynamical properties including the intermediate scattering function for many spatiotemporally correlated systems. Despite its straightforward analysis, DDM has not been fully adopted as a routine characterization tool, largely due to computational cost and lack of algorithmic robustness. We present statistical analysis that quantifies the noise, reduces the computational order, and enhances the robustness of DDM analysis. We propagate the image noise through the Fourier analysis, which allows us to comprehensively study the bias in different estimators of model parameters, and we derive a different way to detect whether the bias is negligible. Furthermore, through use of Gaussian process regression (GPR), we find that predictive samples of the image structure function require only around 0.5%–5% of the Fourier transforms of the observed quantities. This vastly reduces computational cost, while preserving information of the quantities of interest, such as quantiles of the image scattering function, for subsequent analysis. The approach, which we call DDM with uncertainty quantification (DDM-UQ), is validated using both simulations and experiments with respect to accuracy and computational efficiency, as compared with conventional DDM and multiple particle tracking. Overall, we propose that DDM-UQ lays the foundation for important new applications of DDM, as well as to high-throughput characterization.

DOI: [10.1103/PhysRevE.104.034610](https://doi.org/10.1103/PhysRevE.104.034610)**I. INTRODUCTION**

Microscopy has become an essential tool for probing dynamical processes in complex materials and systems, but typically requires sophisticated video image analysis to obtain quantitative information. Although real-space analysis methods retain information regarding individualistic processes within an image, feature tracking algorithms such as multiple particle tracking (MPT) are often computationally expensive and require user interactivity to determine algorithmic parameters to isolate the dynamical process(es) of interest [1,2]. By contrast, Fourier transform-based analysis retains the statistical information encoded within the entire image, and is therefore more sensitive to low-signal processes as well as more robust to nonideal imaging conditions and optically dense systems [3,4]. In this way, Fourier microscopy combines the advantages of real-space imaging in feature identification and segmentation with ensemble-level statistical precision of Fourier-space analysis.

Of the various Fourier-space based approaches available, differential dynamic microscopy (DDM) [5] has emerged as a powerful and versatile analysis method to quantify spatiotemporally correlated dynamics from video microscopy data. This versatility stems from its compatibility with a broad range of

microscopy imaging modes, easy setup with instrumentation available in most research laboratories, and straightforward analysis routines. DDM has been applied to study an ever-broadening range of phenomena in soft and biological matter systems [3,4,6], including analysis of the dynamics of concentrated particle suspensions [7], motions of swimming bacteria [8], binary mixture of molecular fluids [9], and the coarsening dynamics of phase separating colloidal gels [10]. While it is common to assume the material to be isotropic, anisotropic properties such as the viscoelasticity of nematic liquid crystals can also be extracted [11].

For a more comprehensive overview of DDM and its various applications, the interested reader is referred to various reviews on the topic [3,4,12]. This work is concerned with the development of a comprehensive statistical framework that aims at quantifying errors, reducing computational cost, and enhancing the robustness of the analysis of differential dynamic microscopy (DDM) data. Similar developments have been made previously for MPT analysis [2,13], and have greatly improved the robustness and algorithmic development of MPT in various applications. We therefore anticipate similar benefits from a more thorough investigation of uncertainty for DDM. To better motivate these developments, we first summarize the analysis procedure of DDM, estimators employed to extract physical parameters, and highlight the features and limitations of DDM that inspired this study.

In DDM, a time sequence of image stacks represented by the intensity matrix $I(\mathbf{x}, t)$ is processed using a Fourier-based technique where $\mathbf{x} = (x_1, x_2)$ denotes two spatial coordinates

*Corresponding author: mengyang@pstat.ucsb.edu

†Corresponding author: helgeson@ucsb.edu

‡Corresponding author: valentine@engineering.ucsb.edu

and $t \in [t_{\min}, t_{\max}]$. One first calculates the difference in intensity at each pixel location between two frames separated by a lag time Δt :

$$\Delta I(\mathbf{x}, t, \Delta t) = I(\mathbf{x}, t + \Delta t) - I(\mathbf{x}, t). \quad (1)$$

The intensity differences are then Fourier transformed and the absolute values squared to obtain the normalized squared intensity function in Fourier space:

$$|\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 = |\mathcal{F}(\Delta I(\mathbf{x}, t, \Delta t))|^2, \quad (2)$$

where $\mathcal{F}(\cdot)$ denotes the operator of the two-dimensional (2D) discrete Fourier transformation (DFT), $\mathbf{q} = (q_1, q_2)$ is a coordinate wave vector in reciprocal space.

The ensemble average of Eq. (2) is computed to obtain the dynamic image structure function $D(\mathbf{q}, \Delta t)$:

$$D(\mathbf{q}, \Delta t) = \langle |\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 \rangle = \frac{1}{n_{\Delta t} n_q} \sum_{t \in \mathcal{S}_{\Delta t}} \sum_{(q_1, q_2) \in \mathcal{S}_q} |\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2, \quad (3)$$

where $\langle \cdot \rangle$ denotes averaging across all instances of $t \in \mathcal{S}_{\Delta t}$ and $(q_1, q_2) \in \mathcal{S}_q$ with sets $\mathcal{S}_{\Delta t} = \{t : t_{\min} \leq t \leq t_{\max} - \Delta t\}$. The sizes of the two sets are denoted by $n_{\Delta t} = \#\mathcal{S}_{\Delta t}$ and $n_q = \#\mathcal{S}_q$, respectively.

We only consider isotropic materials in this work, in which case $D(\mathbf{q}, \Delta t) = D(q, \Delta t)$ where $\mathcal{S}_q = \{(q_1, q_2) : q_1^2 + q_2^2 = q^2\}$. However, the approach can be generalized to retain a multidimensional q dependence as desired [14].

The following representation is routinely used to relate observables commonly associated with scattering analysis to the observations of $D(q, \Delta t)$ [3,5,6]:

$$D_o(q, \Delta t) = A(q)[1 - f(q, \Delta t)] + B(q, \Delta t), \quad (4)$$

where $A(q)$ is determined by the properties of the imaged material and imaging optics, $B(q, \Delta t)$ is determined by the noise of the detection chain, and the subscript “*o*” denotes the observed value. As will be discussed in Sec. II A, the mean of $B(q, \Delta t)$ is a constant value shared across all q and Δt values, whereas the variance of $B(q, \Delta t)$ depends on the values of q and Δt . The intermediate scattering function (ISF), $f(q, \Delta t)$, is in principle the same as that measured in conventional light scattering measurements such as dynamic light scattering (DLS). The ISF quantifies how the dynamic structure decorrelates over the observed length scale $1/q$ in Fourier space and timescale Δt in real space, which encodes the physical dynamics of the observed system. In general, for randomly fluctuating, ergodic systems, $f(q, \Delta t \rightarrow 0) = 1$ and $f(q, \Delta t \rightarrow \infty) = 0$, and thus, $D_o(q, \Delta t \rightarrow 0) = B(q, \Delta t)$ and $D_o(q, \Delta t \rightarrow \infty) = A(q) + B(q, \Delta t)$.

DDM’s ultimate integration into the characterization workflows of a diverse range of systems is not without challenges. First, we note that, because DDM operates on a series of finite-exposure images taken of time-fluctuating processes, the measured DDM signal will contain inherent error related to both static and dynamic effects, much in the same way MPT incurs static and dynamic errors associated with the imaging process [13]. Because of this, the observed value $D_o(q, \Delta t)$ will in general not be equal to the “true” image structure function $D(q, \Delta t)$ that would be obtained from an ideal imaging system. Separating the signal from the noise in estimating the

image structure function requires properly quantifying the uncertainty of the background noise that propagates through the analysis [5]. The observed intensity $D_o(q, \Delta t)$ typically overestimates $D(q, \Delta t)$, as the mean of the noise term $B(q, \Delta t)$ is positive, and is twice as large as the variance of the noise in the original images [3]. In this study, we show that it is critically important to obtain an accurate estimate of the mean of the noise term $B(q, \Delta t)$, which we denote as B_{est} , to extract dynamic information from systems, such as the mean squared displacement, using DDM.

Several distinct methods to obtain the noise estimator B_{est} in DDM have been proposed in prior studies. However, to our knowledge, there has yet to be a detailed study of how the choice of estimator affects the estimation of dynamic properties. For instance, B_{est} has been assumed to be 0 [15], estimated by the minimum value of $D(q, \Delta t)$ at the temporal resolution Δt_{\min} , denoted as $D_{\min}(\Delta t_{\min})$ [6], as the average of the high- q limit of the observed image structure function $\langle D_o(q_{\max}, \Delta t) \rangle_{\Delta t}$ [16,17], or as the ensemble average of the static power spectrum $2\langle |\hat{I}_o(q_{\max}, t)|^2 \rangle_t$ [18]. Although a particular estimator of the noise term may work well under certain experimental conditions, we will show that all estimators are biased in general (see Sec. II A), and ultimately introduce a way to discern whether the bias is negligible, or if additional measurements are needed to estimate the noise. Indeed, in both simulated scenarios and real experiments, we found that the bias of noise estimation can substantially impact the estimation accuracy of system dynamics. Finally, we note that B_{est} is sometimes treated as a fitting parameter and estimated along with other parameters in the model of $f(q, \Delta t)$ [3,19,20]. This approach is applied to analyzing the active actin dynamics from [20] in Sec. IV C. In general, we find that estimation of the noise parameter is the most challenging among all parameters, and could lead to a poor fit to the observed values. The first contribution of this work is a formal analysis of error propagation and comparison of different estimators in representative experimental contexts.

After obtaining the estimate of the mean of the noise B_{est} , the amplitude parameter $A(q)$ may be estimated by the plateau of intensity through $D_o(q, \Delta t \rightarrow \infty) = A(q) + B_{\text{est}}$ [6,16], or through the connection $A(q) + B_{\text{est}} = \langle |\hat{I}_o(q, t)|^2 \rangle_t$ [5,18,21]. In practice, it may be difficult to accurately obtain $D_o(q, \Delta t \rightarrow \infty)$ as there are very few observations available for intensities at large Δt . Thus, we find that estimation of $A(q)$ through $A(q) + B_{\text{est}} = \langle |\hat{I}_o(q, t)|^2 \rangle_t$ is often more reliable, as it does not require the intensity to reach plateau at large Δt . These approaches will be compared using simulated and experimental observations.

A second challenge of DDM analysis is the computational cost: the most computationally expensive step for DDM is performing a 2D fast Fourier transformation for each pixel of the difference for each of the image pairs $[T \times (T - 1)]/2$ pairs, where $T \sim 10^3 - 10^4$ is the total number of time points. For T images with size $N \times N$ pixels ($N \sim 10^2 - 10^3$), this requires $O[T^2 N^2 \log_2(N)]$ computational operations. Although there has been recent progress in accelerated computation that takes advantage of contemporary computational efficiency for Fourier-based image analysis [7,22], these approaches still require resolving the DDM signal over the full sampled space of q and Δt . To overcome the computational challenge, we use

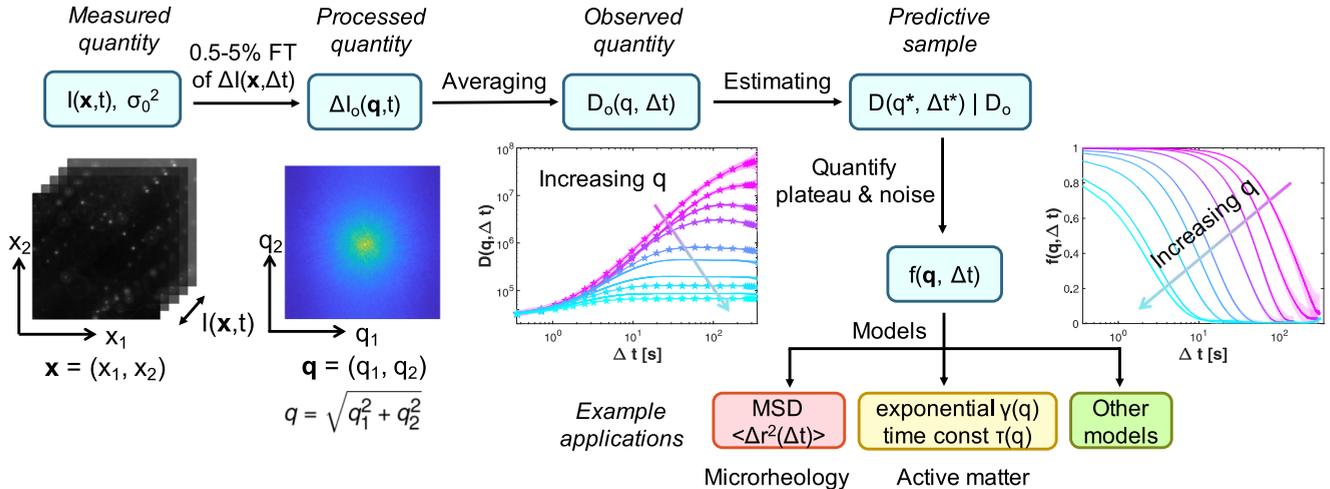


FIG. 1. Schematic representation of DDM-UQ-based data reduction, sampling, and fitting procedure used to determine material constants. From the stack of images acquired by microscopy, around 0.5%–5% of the Fourier transforms are performed to obtain the observed image structure function $D_o(q, \Delta t)$, from which a predictive sample is generated to estimate $D(q^*, \Delta t^*)$ at unobserved q^* and Δt^* , given $D_o(q, \Delta t)$. In the graphs of $D_o(q, \Delta t)$ versus Δt and $f(q, \Delta t)$ versus Δt , at select q 's, the asterisks denote data selected for fitting, the lines denote values at all Δt 's for a particular q . The shadow denotes 95% predictive interval, which is small compared to the range of the change in $D(q, \Delta t)$ over the range of Δt . The predictive samples preserve the quantiles of the distribution after transformation and are then used to find material quantities of interest.

a probabilistic approach to downsample the image stacks and reconstruct all image structure functions based on a fraction of observations, which dramatically reduces the computational cost of the required fast Fourier transform (FFT) algorithm.

The third, and often overlooked aspect, is the robustness of the algorithm(s) for decomposing $D_o(q, \Delta t)$ into its more physically meaningful components through Eq. (4). To ensure applicability to a wide range of materials, it is desirable to allow for an arbitrary form of $f(q, \Delta t)$, such that the method does not require prior knowledge of the system's dynamical properties. Moreover, the estimators used for $A(q)$ and $B(q)$ may contain bias, which can cause the algorithm to be less robust for both small and large Δt 's in estimating $f(q, \Delta t)$ and quantities derived from it [6]. Here we generate the predictive samples based on the observed image structure function $D_o(q, \Delta t)$ and use the predictive median to derive physical parameters within the systems; this approach is more robust than a simple ensemble based on $D_o(q, \Delta t)$ at selected wave vectors.

An exemplifying context for the potential advantages gained by overcoming these limitations is the recent application of DDM to passive probe microrheology as an alternative to conventional approaches such as MPT [6,23]. MPT-based passive probe microrheology involves imaging the Brownian fluctuations of embedded colloidal probes in order to resolve their mean square displacements (MSD) $\langle \Delta r^2(\Delta t) \rangle$ [1], which in the limit of homogeneous, uniform materials can be related to their linear viscoelastic moduli [24,25]. DDM offers an alternative approach to MPT in estimating the MSD through the estimation of $f(q, \Delta t)$ using Eq. (4). DDM presents a number of advantages in this regard, including applicability and better statistical precision to low-signal or optically dense probes and materials [6,17,26,27]. Importantly, since DDM requires no *a priori* user-input parameters associated with the probes or imaging system, it has the potential to provide

automated, user-free analysis that could enable high-throughput characterization [12].

In this work, we make three contributions towards overcoming these challenges of DDM: (1) We relate the mean and variance of the error in the observed image structure function to the variance of the error in the original image intensities. By propagating the error, we show that there exists potential bias in different estimators of the noise and amplitude parameter in DDM, and we propose a way to detect and reduce such bias. (2) We speed up the computation by using Gaussian process regression (GPR) [28] to overcome the computational bottleneck introduced by the Fourier transformations by subsampling the data at selected Δt (Fig. 1). (3) Finally, we use the median of the predictive samples from GPR to robustly estimate the ISF, MSD, and other quantities of interest. Furthermore, we illustrate through a broad range of examples, both simulated and experimental, how the choice of the estimators impacts the accuracy of the resultant MSD and other quantities of interests. We demonstrate that accurate estimation of the noise in image intensities is critical for obtaining an accurate estimation of dynamical information in DDM. We make available a user-friendly MATLAB software package [29] that implements fast computational techniques with uncertainty quantification developed in this work.

II. METHODOLOGY

We name our algorithm *differential dynamic microscopy with uncertainty quantification* (DDM-UQ). The analysis routine is described schematically in Fig. 1. First, image stacks $I(\mathbf{x}, t)$ are acquired with a microscope or are produced using a particle dynamics simulation algorithm. The variance of the background noise intensity σ_0^2 is either assessed independently or estimated from the image stack. Then, a small subsample of a few percent of the image differences are

squared and Fourier transformed to construct a set of observed quantities $D_o(\mathbf{q}, \Delta t)$. Thereafter, GPR is fit to $D_o(\mathbf{q}, \Delta t)$ to obtain a predictive distribution $D(q, \Delta t)$. Eventually, the predictive samples for each q are used to obtain predictive samples for quantities of interest [such as $f(q, \Delta t)$ and $\langle \Delta r^2(\Delta t) \rangle$]. Through analysis of a number of simulated and experimental data sets, we demonstrate that our approach not only reduces the computational time, but in many cases also improves the accuracy and robustness of estimation, as compared to previous DDM approaches and MPT.

A. Error quantification

To develop a statistical approach to error quantification and analysis, we write the observed intensity as

$$I_o(\mathbf{x}, t) = I(\mathbf{x}, t) + \epsilon(\mathbf{x}, t), \quad (5)$$

where $I(\mathbf{x}, t)$ is an (unknown) deterministic function of the observed sample and imaging system; $\epsilon(\mathbf{x}, t)$ is an independent random noise with mean zero and variance σ_0^2 ; the subscript “ o ” denotes the observed value. Several artifacts are known to impact the accuracy of DDM and are expected to contribute to $\epsilon(\mathbf{x}, t)$, including camera detection noise, edge effects arising from the finite field of view [30], and effects of finite exposure time [15]. Others are known to affect MPT, such as the depth of field [2] and finite pixel size [13], and are expected to impact DDM as well. Here we consider $\epsilon(\mathbf{x}, t)$ to be the difference between the measured signal and the “true” intensity $I(\mathbf{x}, t)$ at each pixel without regard to the actual physical origin of the error. We note that there are other known spatially or temporally correlated artifacts, such as illumination fluctuations, that do not satisfy the criteria assumed for $\epsilon(\mathbf{x}, t)$. These will not be considered in the present analysis.

We illustrate how the error in Eq. (5) propagates in the analysis of DDM. The derivation of Eqs. (6)–(10) is given in Appendix A. Assuming that Eq. (5) holds, we can express the observed squared intensity function in reciprocal space as

$$\begin{aligned} & |\Delta \hat{I}_o(\mathbf{q}, t, \Delta t)|^2 \\ &= |\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 + 2\Delta \hat{I}(\mathbf{q}, t, \Delta t)\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t) \\ & \quad + |\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)|^2, \end{aligned} \quad (6)$$

where the closed form expressions of $|\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2$, $\Delta \hat{I}(\mathbf{q}, t, \Delta t)\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)$, and $|\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)|^2$ are given in Eqs. (A1)–(A3) in Appendix A, respectively.

The expected value (i.e., mean) of $|I_o(\mathbf{q}, t, \Delta t)|^2$ is given by

$$\mathbb{E}[|\hat{I}_o(\mathbf{q}, t, \Delta t)|^2] = |\hat{I}(\mathbf{q}, t, \Delta t)|^2 + 2\sigma_0^2. \quad (7)$$

Note that the mean of the cross-product term $\langle \Delta \hat{I}(\mathbf{q}, t, \Delta t)\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t) \rangle$ is zero under the assumptions made for $\epsilon(\mathbf{x}, t)$.

By combining Eqs. (3) and (6), we can express the observations of the dynamic image structure function as follows:

$$\begin{aligned} D_o(q, \Delta t) &= \langle |\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 \rangle + 2\langle \Delta \hat{I}(\mathbf{q}, t, \Delta t)\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t) \rangle \\ & \quad + \langle |\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)|^2 \rangle, \end{aligned} \quad (8)$$

where $\langle \cdot \rangle$ denotes the ensemble with respect to $\mathbf{q} \in \mathcal{S}_q$ and $t \in \mathcal{S}_{\Delta t}$. The mean of $D_o(q, \Delta t)$ follows:

$$\mathbb{E}[D_o(q, \Delta t)] = D(q, \Delta t) + 2\sigma_0^2. \quad (9)$$

Further assuming $\epsilon(\mathbf{x}, t) \sim \mathcal{N}(0, \sigma_0^2)$ independently, we can calculate the variance

$$\begin{aligned} & \mathbb{V}[D_o(q, \Delta t)] \\ &= \frac{2\sigma_0^2}{n_q n_{\Delta t}} \left[2\sigma_0^2 + 2D(q, \Delta t) + \max(0, (T - 2\Delta t)) \right. \\ & \quad \left. \times \left(\frac{\sigma_0^2}{n_{\Delta t}} - \frac{2S_{q, \Delta t}}{(T - 2\Delta t)n_{\Delta t}n_q} \right) \right], \end{aligned} \quad (10)$$

where the expression of $S_{q, \Delta t}$ is given in Eq. (A5) in Appendix A.

The result in Eq. (10) is intuitive: the ratio $\frac{1}{n_q n_{\Delta t}}$ arises from the fact that the $D_o(q, \Delta t)$ is averaged from n_q and $n_{\Delta t}$ observations of $\langle \hat{I}(\mathbf{q}, t, \Delta t) \rangle^2$, as shown in Eq. (3), which decreases the variance. The other terms arise from the covariance between the sin and cos terms from the Fourier transform and that of the recursive sampling of the same image in different Δt .

Note that by Eq. (9), an unbiased estimator of $D(q, \Delta t)$ is $D_o(q, \Delta t) - 2\sigma_0^2$, while using the observations $D_o(q, \Delta t)$ alone typically overestimates the image structure function by $2\sigma_0^2$ on average. Potential practical procedures for estimating $2\sigma_0^2$ will be discussed later.

We have shown that $D_o(q, \Delta t)$ can be separated into a deterministic term of the signal $[D(q, \Delta t)]$ and a random term containing the noise and cross product of the noise and signal. This representation can be related to Eq. (4) by letting

$$B(q, \Delta t) = 2\langle \Delta \hat{I}(\mathbf{q}, t, \Delta t)\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t) \rangle + \langle |\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)|^2 \rangle, \quad (11)$$

where the mean and variance

$$\mathbb{E}[B(q, \Delta t)] = 2\sigma_0^2, \quad (12)$$

$$\mathbb{V}[B(q, \Delta t)] = \mathbb{V}[D_o(q, \Delta t)], \quad (13)$$

where $\mathbb{V}[D_o(q, \Delta t)]$ is given in Eq. (10). We observe that specifying B as 0 typically underestimates the mean of the noise term. On the other hand, specifying B as the average of the high- q limit of observed image structure function $\langle D_o(q_{\max}, \Delta t) \rangle_{\Delta t}$, or as the ensemble average of static power spectrum $2\langle |\hat{I}_o(q_{\max}, t)|^2 \rangle_t$ [18], tends to overestimate the mean of the noise by a small amount $\langle A(q_{\max})[1 - f(q_{\max}, \Delta t)] \rangle_{\Delta t}$. Note that $[1 - f(q_{\max}, \Delta t)]$ is close to 1 for large Δt . Thus, the bias is non-negligible when $A(q_{\max})$ is large. Also note that $A(q_{\max})$ typically increases when the number of objects in the image or their peak intensity increases, when the image pixel size increases, or when the object size decreases; the relationship of $A(q)$ to some of these quantities was considered in [31]. Indeed, we found that for a system with a large number of small objects, the bias can be large (Fig. 5, Appendix C). When the pixel size is large, such as in the case of the actively driven system considered in Sec. IV C, one may also tend to overestimate the noise. Similarly, using $D_{\min}(q, \Delta t_{\min})$, i.e., the minimum value of

TABLE I. Potential bias in estimation approaches of noise B_{est} in DDM.

B_{est}	Scenarios for non-negligible bias
0	Underestimation in all scenarios
$\langle D_o(q_{\text{max}}, \Delta t) \rangle_{\Delta t}$	Overestimation when particle number, peak intensity, or pixel size is large, or if particle size is small
$2\langle \hat{I}_o(q_{\text{max}}, t) ^2 \rangle_t$	Overestimation in similar scenarios as the above
$D_{\text{min}}(\Delta t_{\text{min}})$	Overestimation when $[1 - f(q, \Delta t_{\text{min}})]$ is not close to zero in similar scenarios as the above

intermediate scattering function at the smallest Δt , may also overestimate B in these scenarios. We found that the overestimation by $B_{\text{est}} = D_{\text{min}}(q, \Delta t_{\text{min}})$ may be smaller than the ones by $B_{\text{est}} = \langle D_o(q_{\text{max}}, \Delta t) \rangle_{\Delta t}$ or $B_{\text{est}} = 2\langle |\hat{I}_o(q_{\text{max}}, t)|^2 \rangle_t$, as the term $[1 - f(q, \Delta t_{\text{min}})]$ can be close to zero. Furthermore, both estimators may slightly underestimate the noise due to its stochastic nature. For instance, when the signal contained in $D_o(q, \Delta t_{\text{min}})$ is close to zero, the ensemble average across q 's serves as a good estimator, whereas the minimum tends to underestimate the noise. In these cases, $D_o(q, \Delta t)$ tends to underestimate the noise much less frequently than overestimating the noise.

We summarize the limitations for each of the available approaches in estimating the mean of the noise in DDM in Table I and offer a simple test to detect the bias in Appendix C. In such a scenario, one may change experimental conditions, such as by reducing the number of objects, or increasing the object size, reducing the pixel size, etc., to reduce the bias; alternatively, the noise can be measured independently, or a more accurate estimator of the noise may be used. One goal of this exercise is to illustrate the importance of noise quantification, and to provide a way to detect potential bias in a wide range of scenarios.

For applications of DDM to microrheology, and assuming dilute probes with diffusive particle dynamics involving Gaussian displacements, we can relate the MSD $\langle \Delta r^2(\Delta t) \rangle$ at each q to the image structure function and related quantities as follows [6,32]:

$$f(q, \Delta t) = \exp\left(-\frac{q^2 \langle \Delta r^2(\Delta t) \rangle}{4}\right) \times \left[1 + \frac{\alpha_2 q^4 \langle \Delta r^2(\Delta t) \rangle^2}{32} + \dots\right], \quad (14)$$

where the first order non-Gaussian parameter $\alpha_2 = \frac{d \langle r^4(\Delta t) \rangle}{(d+2) \langle \Delta r^2(\Delta t) \rangle^2} - 1$ (d = dimensionality) is a measure of the heterogeneity or nondiffusive dynamics of the sample [33]. As is common in microrheology, we assume the contribution of the non-Gaussian parameter is negligible [24], and thus $\Delta r^2(q, \Delta t)$ at each q may be estimated from $D(q, \Delta t)$ through the following approach:

$$\Delta r_{\text{est}}^2(q, \Delta t) = \frac{4}{q^2} \ln \left[\frac{A(q)}{A(q) - D_o(q, \Delta t) + B(q, \Delta t)} \right]. \quad (15)$$

It is common to assume sample ergodicity, and use the ensemble average of $\Delta r^2(q, \Delta t)$ to estimate the mean squared displacement $\langle \Delta r^2(\Delta t) \rangle$. However, we note that the variance

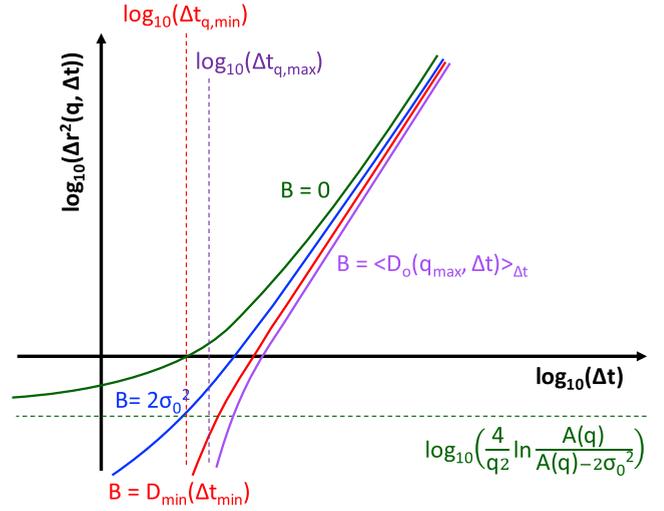


FIG. 2. Illustration of how four different approaches to estimating $B(q)$ influence the calculated MSD. For given q , the quantity $\Delta t_{q,\text{min}}$ satisfies $D_o(q, \Delta t_{q,\text{min}}) = D_{\text{min}}(\Delta t_{\text{min}})$.

of $D_o(q, \Delta t)$ given in Eq. (10) is not the same across different values of q and Δt . A simple ensemble average over wave vectors without addressing the weights due to different variances could introduce substantial bias in the estimation, when $D(q, \Delta t)$ at different wave vectors is not the same. We found that using the median, instead of the mean, can be more robust in such estimations. Numerical comparison between these estimation approaches will be discussed in the context of the simulated and experimental studies.

To illustrate the importance of proper error estimation in estimating MSD, we consider the outcomes using four different ways of estimating B , graphed in Fig. 2. If there exists $\Delta t_{q,\text{min}}$, such that $D_o(q, \Delta t_{q,\text{min}}) = D_{\text{min}}(\Delta t_{\text{min}})$ for a given q , and B is estimated by $D_{\text{min}}(\Delta t_{\text{min}})$ (red solid line), then from Eq. (15), the argument of the natural logarithm approaches unity, and $\Delta r^2(q, \Delta t_{q,\text{min}}) \rightarrow 0$, which in turn drives $\log_{10}(\Delta r^2(q, \Delta t_{q,\text{min}})) \rightarrow -\infty$, as shown. Choosing $B = \langle D_o(q_{\text{max}}, t) \rangle_t$ (purple solid line) will similarly overestimate the noise at small Δt , when $A(q)$ does not sufficiently approach 0 at q_{max} .

On the other hand, if B is estimated to be 0 (green solid line), then approximately $\log_{10}(\Delta r^2(q, \Delta t \rightarrow 0)) \rightarrow \log_{10}[\frac{4}{q^2} \ln(\frac{A(q)}{A(q)-2\sigma_0^2})]$ (an asymptotic value denoted by the green dotted line), as the expected value of $D_o(q, 0)$, $\mathbb{E}(D_o(q, 0)) = 2\sigma_0^2$. When we use the correct estimator $B = 2\sigma_0^2$ (blue solid line), the estimated $\Delta r^2(q, \Delta t) \rightarrow 0$ when $\Delta t \rightarrow 0$.

These asymptotic limits demonstrate the crucial importance of properly estimating the mean of B by σ_0^2 when extracting dynamical properties from DDM such as the ISF or MSD, particularly at small Δt , a result that is further validated in simulated and experimental examples (see, e.g., Figs. 4 and 5). A similar point about B was noted previously by other researchers: while uncertainty in $A(q)$ is considered to dominate the analysis because it pertains to the signal, overestimating $B(q)$ such as in [6] can lead to spurious results when computing $\langle \Delta r^2(\Delta t) \rangle$ [23], and hence other authors proposed an

iterative scheme to solve for $\langle \Delta r^2(\Delta t) \rangle$. Treating B as a fitting parameter was also commonly used in prior studies [5,20,34], although the optimized value of B can be unstable in some scenarios, particularly if the weights of $D_o(q, \Delta t)$ are not properly accounted for (see, e.g., Fig. 11). Through various simulation and real experiments in this study, we advocate that accurately estimating the noise parameter is crucial in DDM. In experiment, the noise term σ_0^2 may be measured, typically through independent experiments using immobilized particles under identical imaging conditions, if a large bias of the estimator is detected. The variance of the image difference is then computed to give σ_0^2 . In MPT, a “noise floor” is commonly quantified [13] and frequently subtracted from all data to give a more realistic estimate of the MSD [35–37]. We show that while many estimators have negligible bias in approximating $2\sigma_0^2$ in many cases, other times it may be necessary to evoke an independently measured $2\sigma_0^2$, and we propose that noise characterization should be routine for DDM as well.

B. Gaussian process regression

The second challenge of DDM is the computational bottleneck that arises when performing a massive number of Fourier transformations. We overcome this problem by representing the logarithm of image structure function by a Gaussian process regression (GPR) approach on a fraction (0.5%–5%) of data. GPR is a widely used machine learning tool for estimating nonlinear, smooth response surfaces and the predictions from GPR is the equivalence to the kernel ridge regression (KRR) estimator [38–46].

We apply GPR to the logarithm of $D_o(q, \Delta)$, denoted $\tilde{D}_o(\theta) = \ln[D_o(\theta)]$, with two input parameters: the natural logarithm of the wave vector (in reciprocal space) and time are denoted by $(\tilde{q}, \tilde{\Delta}) = (\ln(q), \ln(\Delta t)) = \theta$. After obtaining the predictive samples of the logarithm of the image structure function, we transform it back to obtain predictive samples for the image structure function. Because the logarithm of the ISF is smoother, the GPR approach works much better using logarithm of $D_o(q, \Delta)$ as observations. Assuming that n observations $\tilde{D}_o(\theta_i)$, $i = 1, 2, \dots, n$, are used, predictions by GPR can be represented through the following optimization, which simultaneously penalizes mean squared errors of the estimation with respect to the observations, as well as the complexity of the estimation:

$$\tilde{D}_* = \operatorname{argmin}_{\tilde{D} \in \mathcal{H}} \left\{ \frac{1}{n} [\tilde{D}_o(\theta_i) - \tilde{D}(\theta_i)]^2 + \lambda \|\tilde{D}\|_{\mathcal{H}}^2 \right\}, \quad (16)$$

where $\|\cdot\|_{\mathcal{H}}$ denotes the reproducing kernel Hilbert spaces regression (RKHS) norm (or native norm) [28] that penalizes the complexity of the estimation to avoid overfitting and λ is a regularization parameter. For any θ_* , the solution of (16), known as KRR, is a weighted average of observations

$$\tilde{D}_*(\theta_*) = \mathbf{w}^T \tilde{\mathbf{D}}_o = \sum_{i=1}^n w_i \tilde{D}_o(\theta_i), \quad (17)$$

where $\mathbf{w} = (w_1, \dots, w_n) = \mathbf{r}_{\theta_*}^T \tilde{\mathbf{R}}^{-1}$ is a row vector of weights, where $\tilde{\mathbf{R}} = \mathbf{R} + n\lambda \mathbf{I}_n$ with \mathbf{I}_n being an identity matrix of size n and \mathbf{R} is an $n \times n$ correlation with (i, j) th entry parametrized by a kernel function $K(\theta_i, \theta_j)$, and $\mathbf{r}_{\theta_*} =$

$(K(\theta_1, \theta_*), \dots, K(\theta_n, \theta_*))^T$ is the correlation between predictive output and observations.

Note that Eq. (17) is only a point estimator without giving assessment of uncertainty. One advantage of the GPR approach is the uncertainty of estimation can be quantified in a probabilistic framework. We model the latent function $\tilde{D}_o(\cdot)$ by a Gaussian process with noises, meaning that any marginal distribution $\tilde{\mathbf{D}}_o = (\tilde{D}_o(\theta_1), \dots, \tilde{D}_o(\theta_n))^T$ at n inputs $\{\theta_1, \dots, \theta_n\}$ follows a multivariate normal distribution:

$$((\tilde{D}(\theta_1), \dots, \tilde{D}(\theta_n))^T | \mathbf{m}, \tilde{\mathbf{R}}, \sigma^2) \sim \mathcal{MN}(\mathbf{m}, \sigma^2 \tilde{\mathbf{R}}), \quad (18)$$

where $\mathbf{m} = (m(\theta_1), \dots, m(\theta_n))^T$ is a vector of the mean [assumed to be a constant in this work, i.e., $\mathbf{m} = (m, \dots, m)^T$] and σ^2 is a variance parameter.

The power (stretched) exponential covariance function and Matérn covariance function are often used for GPR [28]. For any two inputs $\theta_a = (\tilde{q}_a, \tilde{\Delta}_a)$ and $\theta_b = (\tilde{q}_b, \tilde{\Delta}_b)$, we use a product covariance function $\sigma^2 K(\theta_a, \theta_b) = \sigma^2 K_1(\tilde{q}_a, \tilde{q}_b) K_2(\tilde{\Delta}_a, \tilde{\Delta}_b)$, with $K_l(\cdot, \cdot)$, $l = 1, 2$, following a Matérn correlation with roughness parameter $\frac{5}{2}$ such that

$$K_l(x_a, x_b) = \left(1 + \sqrt{5} \beta_l d + \frac{5 \beta_l^2 d^2}{3} \right) \exp(-\sqrt{5} \beta_l d), \quad (19)$$

where $d = |x_a - x_b|$ for any real valued input x_a and x_b with inverse range parameter $\beta_l \in \mathbb{R}^+$, $l = 1, 2$. The sample path of the Gaussian process with Matérn correlation in (19) is twice differentiable and is often used as a default correlation in GPR [47].

The parameters in GPR ($m, \sigma, \beta, \lambda$) can be estimated by the maximum likelihood approach discussed in Appendix B. Plugging in the estimated parameters ($m_{\text{est}}, \sigma_{\text{est}}^2, \beta_{\text{est}}, \lambda_{\text{est}}$), the predictive distribution of $\tilde{D}(\theta_*)$ at any θ_* follows a normal distribution [28]:

$$(\tilde{D}(\theta_*) | \tilde{\mathbf{D}}_o) \sim \mathcal{N}(\tilde{D}_{\text{est}}(\theta_*), \hat{\sigma}^2 K_*(\theta_*, \theta_*) + \tilde{\sigma}_*^2), \quad (20)$$

where $\tilde{\sigma}_*^2$ is the variance of the noise ϵ_{θ_*} with

$$\tilde{D}_{\text{est}}(\theta_*) = m_{\text{est}} + \mathbf{r}_{\theta_*}^T \tilde{\mathbf{R}}^{-1} (\tilde{\mathbf{D}}_o - m_{\text{est}} \mathbf{1}_n), \quad (21)$$

$$K_*(\theta_*, \theta_*) = \sigma_{\text{est}}^2 (K(\theta_*, \theta_*) - \mathbf{r}_{\theta_*}^T \tilde{\mathbf{R}}^{-1} \mathbf{r}_{\theta_*}). \quad (22)$$

We use the predictive median $\tilde{D}_{\text{est}}(\theta_*)$ for predicting the logarithm of the ISF to obtain the unsampled θ_* and the predictive median of $D_o(\theta_*)$ can be obtained by transforming the predictive median of $\tilde{D}_o(\theta_*)$ through the exponential function. The predictive median in Eq. (21) is equivalent to KRR in Eq. (17) when the mean parameter is zero $m_{\text{est}} = 0$. Here the uncertainty of predictions and predictive samples can be obtained by the predictive distribution in Eq. (20).

C. Predictive sampling by a downsampled data set

After obtaining the image intensities, DDM-UQ analysis starts by selecting a fraction of the Δt 's equally spaced logarithmically along the input space coordinates to compute $D_o(q, \Delta t)$ as shown in Fig. 3(a). Here, we choose not to downsample observations at wave vectors (q), but the approach can be extended to q as well, if one wishes to further reduce the computational cost. The Fourier transformation is only performed on this reduced set of image differences,

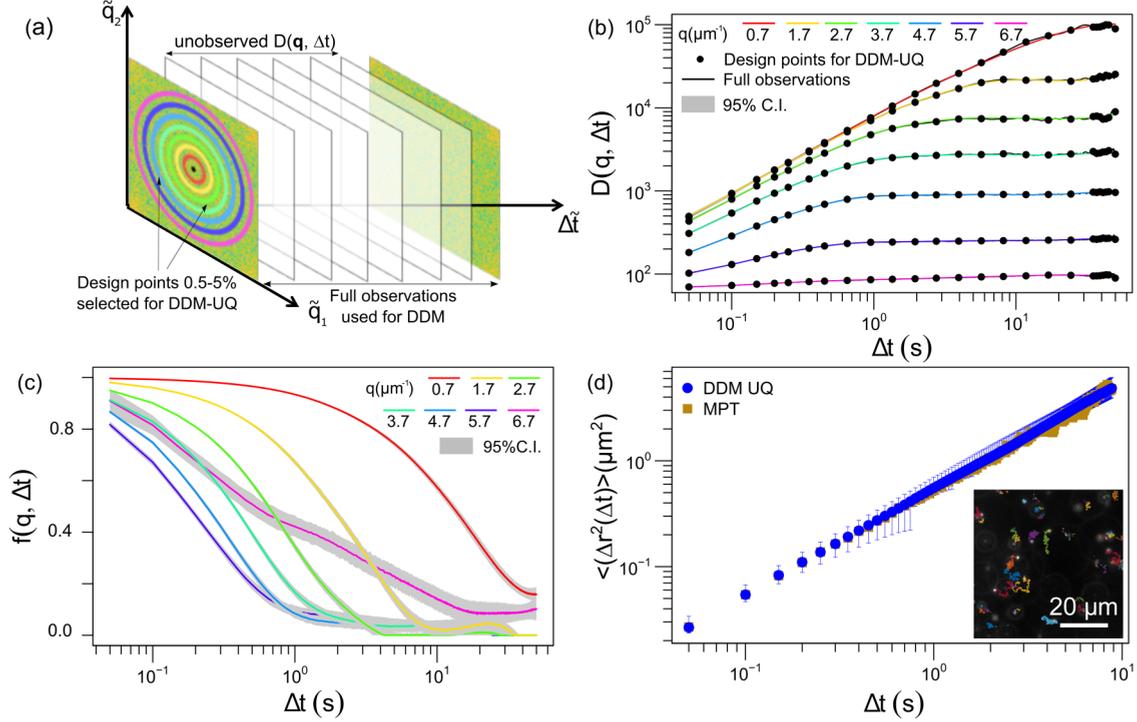


FIG. 3. (a) The design points (blue circles) are selected along the logarithmically scaled coordinates $(\mathbf{q}, \Delta t)$, requiring only a fraction of the Fourier transformations to compute $D_o(\mathbf{q}, \Delta t)$. (b) Predictive median and predictive samples by GPR for representative values of q 's. The variance of noise is proportional to the inverse of the number of pixels in ensemble. 300 predictive samples for each q were generated to obtain the 95% credible interval (gray shadow). The observations used for GPR are plotted as black dots. The full observations (black lines) overlap with the predictions (colored lines). (c) The predictive samples for the intermediate scattering function are plotted against the Δt . (d) Predictive mean squared displacement is plotted against the Δt using all three methods. The 95% predictive interval is shown with the error bar for DDM-UQ. The inset shows a snapshot of experiment with MPT trajectory overlay. Plots (b)–(d) are derived from a movie with 1- μm probe particles diffusing in a viscous fluid.

allowing for fast, high-throughput analysis. Since an estimate of the plateau of the image structure function at long times is often required by the analysis, we ensure that at least five design points lie on the interval $[0.7\Delta t_{\text{max}}, 0.9\Delta t_{\text{max}}]$ on the $\Delta \tilde{t}$ axis for each chosen \tilde{q} [Fig. 3(b)]. For all numerical results analyzed in this study, we only use $D_o(\mathbf{q}, \Delta t)$ at 25 Δt points in DDM-UQ, which represents around 0.5% to 5% of total observations. The predictive median of $D_o(\mathbf{q}, \Delta t)$ is smoother than the observed $D_o(\mathbf{q}, \Delta t)$ since spurious noise is effectively filtered out [see, e.g., the black curves and colored curves in Fig. 3(b)].

Note that our goal is to relate $D_o(\mathbf{q}, \Delta t)$ to other quantities of interest, such as the intermediate scattering function $f(\mathbf{q}, \Delta t)$ or mean squared displacement $\langle \Delta r^2(\Delta t) \rangle$, both of which are nonlinear transformations of $D_o(\mathbf{q}, \Delta t)$. To achieve this, we sample image structure functions from the predictive distribution in Eq. (20) and transform the samples to obtain other quantities of interest. The transformed predictive samples can be used to estimate the predictive interval of quantities of interest at a given set of input $(\mathbf{q}, \Delta t)$. In this work, we sample 1000 observations, denoted as $D_s(\mathbf{q}, \Delta t)$ for $s = 1, 2, \dots, 1000$ at any $(\mathbf{q}, \Delta t)$, and transform $D_s(\mathbf{q}, \Delta t)$ to obtain other quantities of interest, such as intermediate scattering function $f_s(\mathbf{q}, \Delta t)$ via Eq. (4) after estimating B_{est} and $A_{\text{est}}(\mathbf{q})$. The determination of these two estimators is discussed in Sec. III A. For each $(\mathbf{q}, \Delta t)$, the 95% predictive interval

of $f(\mathbf{q}, \Delta t)$ can be estimated by the lower 2.5% quantile and the upper 2.5% quantile of the transformed predictive samples $f_s(\mathbf{q}, \Delta t)$ for $s = 1, 2, \dots, 300$.

Since the predictive samples of $D(\mathbf{q}, \Delta t)$ preserve information such as quantiles of distribution for any transformation, transforming these samples can be used to estimate the ISF $f(\mathbf{q}, \Delta t)$ and MSD $\langle \Delta r^2(\Delta t) \rangle$ as shown in Figs. 3(c) and 3(d). The predictive median is used for estimating MSD at each Δt as it is typically more robust than the mean. Conditional on the observed values $D_o(\mathbf{q}, \Delta t)$, the predictive samples of $f(\mathbf{q}, \Delta t)$ and $\langle \Delta r^2(\Delta t) \rangle$ can be used to assess estimation uncertainty as well.

III. VALIDATION THROUGH SIMULATION

A. Image formation and analysis approach

To validate our methodology, we first employ simulations of a time series of images demonstrating particle motion. This approach has the significant advantage that the true particle motion is known *a priori* and the true MSD has a closed form expression, thus allowing quantitative comparison with results obtained using DDM, DDM-UQ, and MPT, the latter obtained using an open source tracking algorithm [1,48]. Moreover, it is possible to investigate systematically how different sources of noise influence algorithm performance, and whether different functional forms of $\langle \Delta r^2(\Delta t) \rangle$ perform differently.

We first examine simulated video images of particles in motion, each with a Gaussian intensity profile with peak intensity $I_c = 255$ and standard deviation of $\sigma_p = 2$ pixels. In principle, the intensity recorded at a single pixel $I_p(\mathbf{x})$ could arise from intensity contributions of multiple particles in the vicinity. The contribution of the j th particle located in $\mathbf{x}_j(t)$ is given by

$$I_p(\mathbf{x}, \mathbf{x}_j(t)) = I_c \exp\left(-\frac{[\mathbf{x} - \mathbf{x}_j(t)]^2}{2\sigma_p^2}\right). \quad (23)$$

To account for noise in the background intensity signal, a time-varying, random uniform noise $I_b(\mathbf{x}, t)$, centered around zero, in the range $[-10, 10]$ is added. Thus, we may compute σ_0^2 as the variance of the background noise $I_b(\mathbf{x}, t)$: $\sigma_0^2 = \frac{20^2}{12} \approx 33.3$. Thus, the signal at a time t is the sum of signals attributed to all particles as well as the background:

$$I(\mathbf{x}, t) = I_b(\mathbf{x}, t) + \sum_{j=1}^{n_p} I_p[\mathbf{x} - \mathbf{x}_j(t)]. \quad (24)$$

Note that the pixel intensities in simulation are not subjected to a cutoff ceiling value as are those obtained in imaging (e.g., 0–255 for an eight-bit image). Moreover, unlike MPT, where the relative brightness of particle and background significantly affects tracking precision [13], in DDM signal quality depends sensitively on the magnitude of the image difference. Keeping this context in mind, we simulate particles with brightness $I_p[\mathbf{x} - \mathbf{x}_j(t)]$ that does not vary with time, and we vary the step size by which the particles move in each time step instead. It is well known that DDM performance deteriorates, and can completely break down, in the limit of small probe displacements [6]. For diffusive particles taking a step with a variance σ_s^2 , we compare the performance of DDM, DDM-UQ, and MPT in calculating the MSD values.

The DDM-UQ analysis represents data obtained using our proposed approach based on the downsampled Fourier transformation of $\Delta I(\mathbf{x}, \Delta t)$'s with GPR and predictive samples. For all numerical results that extract the MSD using DDM-UQ, we estimate B_{est} to be the minimum of $D_{\min}(\Delta t_{\min})$ and $\langle D_o(q_{\max}, \Delta t) \rangle_{\Delta t}$, to reduce the overestimation bias, as summarized in Table I. The estimate of the amplitude parameter $A_{\text{est}}(q)$ is obtained by $A_{\text{est}}(q) + B_{\text{est}} = 2\langle I_o^2(q, t) \rangle_t$, as it applies to all wave vectors, regardless of whether or not a plateau is reached at the given Δt . Furthermore, extracting the MSD from the ISF at extremal wave vectors could lead to large errors. This is because the intensities of only a few pixels are used to calculate the ISF at small wave vectors, leading to large uncertainty in estimation. On the other hand, $A(q)$ can approach zero at large wave vectors, leading to unstable estimate of MSD in this regime. To avoid these extremes, DDM-UQ uses $D_o(q, \Delta t)$ based on 80 intermediate q values, typically ranging from the 4th up to 83rd largest q 's.

Additionally, we record the MSD estimations for Δt values that are no larger than 90% of the estimated plateau values according to Eq. (15). In this limit, the denominator is sufficiently different from zero to ensure a small variance of the estimation. Finally, we truncate the estimation of MSD at those Δt where fewer than 10 wave vectors are available, to avoid selection bias in estimation when sampling is limited. Other approaches to truncating the wave-vector range and lag

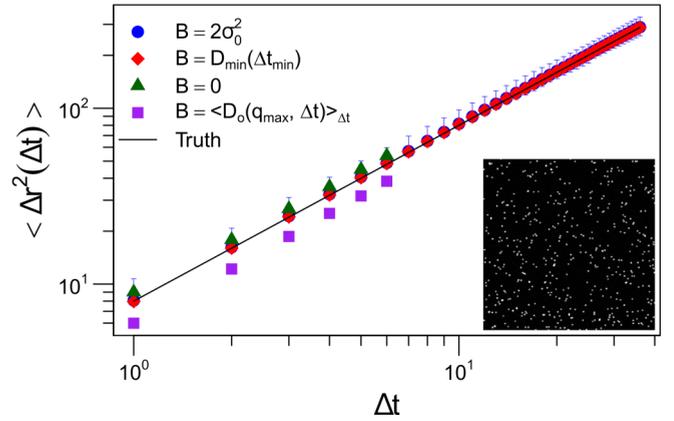


FIG. 4. The ensemble-averaged MSD calculated from the simulated 2D Brownian motion of 800 particles. Estimation of MSD by DDM-UQ with four different ways of estimating the mean of the noise. The error bars denote 95% predictive interval of DDM-UQ with $B = 2\sigma_0^2$. The truth of $\langle \Delta r^2(\Delta t) \rangle = 8\Delta t$ is denoted as the black line. The inset shows the initial position of particles, where particles are enlarged for better visualization. Note that it is not possible to perform MPT due to the large number of particles moving within the frame.

time based on standard deviation of the data were explored in prior DDM analyses [6]. Weighting $D(q, \Delta t)$ based on the variance of the data without truncation of wave vectors and time points could be an efficient way to reduce selection bias while allowing robust estimation of the extracted MSD at longer Δt . This is a potential future topic for research.

For DDM analysis in practice, we perform the Fourier transformation of all values of $\Delta I(\mathbf{x}, \Delta t)$, and estimate $B(q)$ using the average of $D(q_{\max}, \Delta t)$ over all Δt 's at q_{\max} , $\langle D(q_{\max}, \Delta t) \rangle$. Our first simulated case contains 800 small particles (Fig. 4), whereas we include 50 moderately large particles in the latter 6 simulations. As shown in Fig. 5, all estimators of the mean of the noise B for the six latter simulated examples are very similar, as expected. We then test how the DDM analysis is affected by estimating $A(q)$ two different ways. Following the procedures (I) from [6] we estimate $A(q)$ from the plateau or (II) using the static power spectrum [i.e., $A_{\text{est}}(q) + B_{\text{est}} = 2\langle I_o^2(q, t) \rangle_t$] per [16,17]. We largely follow the procedures defined by [6,31] to select the wave vectors and Δt values for estimation and data selection for both DDM analysis approaches.

Also included is the MPT approach, which was performed by locating particles in each frame, and searching in the vicinity to link trajectories of individual particles. The localization error is not characterized, given the high-particle intensity compared to the background intensity in all cases investigated. Note that for certain cases, such as optically dense cases of simple diffusive processes compared in Sec. III B, MPT cannot identify the large number of probes, so MPT results are not compared in these two studies.

In detail, in all cases, we generated videos of the motion of $n_p = 50$ particles, except for the case described in Sec. III B, where $n_p = 800$. The simulation box is a 2D square with sides $L = 480$. The movie spans time steps $t = 1, 2, \dots, 1000$. We further imposed displacements in each successive time step

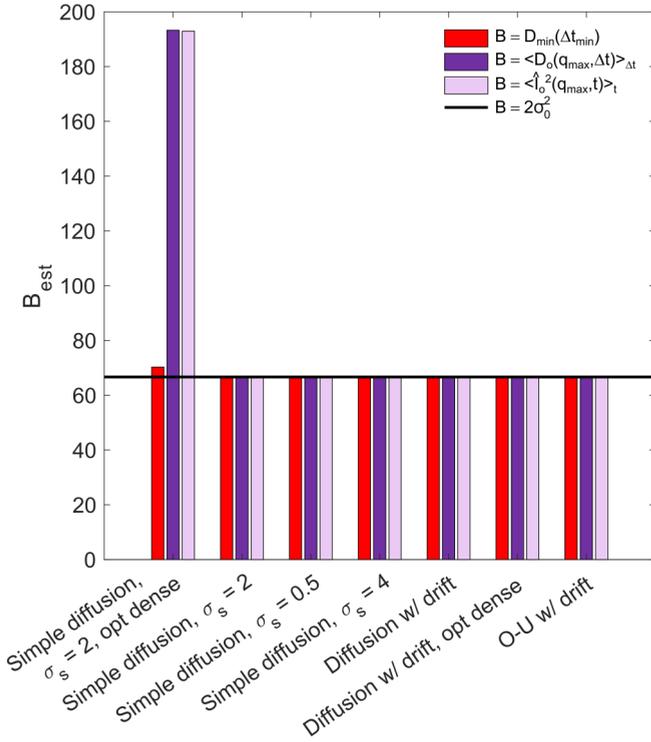


FIG. 5. Comparison of different estimators (denoted by different color bars) for all simulation scenarios. Note that the true noise $2\sigma_0^2$ is kept constant in all simulations and denoted by the thick black line.

$\Delta x_{i,j}(t) = x_{i,j}(t+1) - x_{i,j}(t)$ (where $i = 1, 2$ stands for the x_1 , or x_2 directions in the Cartesian coordinates in 2D).

We construct three scenarios that represent the general features observed in a broad range of experiments: simple diffusion, diffusion with drift, and constrained diffusion within a harmonic potential well (i.e., an Ornstein-Uhlenbeck process). These scenarios will result in distinct shapes of $\langle \Delta r^2(\Delta t) \rangle$ and highlight various challenges to each analytical approach. The derivations of the expected values of the MSDs for all simulated scenarios are given in Appendix D.

As a metric of the accuracy of a given analysis method, we compare the normalized root mean squared error (N-RMSE) for the estimated MSD relative to the known true MSD:

$$\text{N-RMSE} = \frac{\sqrt{\frac{1}{n_{\Delta t}} \sum_{\Delta t \in \Delta \mathcal{T}} [\langle \Delta \tilde{r}^2(\Delta t) \rangle - \langle \Delta \tilde{r}_{\text{est}}^2(\Delta t) \rangle]^2}}{\tilde{\sigma}_r}, \quad (25)$$

where $\langle \Delta \tilde{r}^2(\Delta t) \rangle$ is the logarithm of the true MSD with base 10, $\langle \Delta \tilde{r}_{\text{est}}^2(\Delta t) \rangle$ is the corresponding estimate using DDM

with two different $A(q)$ estimators, DDM-UQ or MPT, and $\tilde{\sigma}_r$ is the sample standard deviation of the logarithm of the true MSD with base 10. In practice, not all Δt values are available for every method, due to large fluctuations at large Δt values, and this provides a limit to the total Δt range captured in each case. To ensure that the four methods are evaluated on the same test set $\Delta \mathcal{T}$ and to ease quantitative comparisons, we determine the usable range of Δt 's by the smallest maximum Δt available among the three methods. The N-RMSE of different simulated cases is summarized in Table II.

B. Validating the use of σ_0^2 as an estimator for B

We have shown that the mean of B is $2\sigma_0^2$, and that the estimation of σ_0^2 is critically important to the analysis of DDM data. To illustrate this point, we first show the results of a simulation of $n_p = 800$ particles with $\sigma_p = 0.5$ moving in a purely viscous fluid. We generate movies demonstrating simple Brownian motion: at each time step, $\Delta x_{i,j}(t) \sim \mathcal{N}(0, \sigma_s^2)$ independently, where σ_s represents the step size with units of pixels. The expected MSD of the Brownian motion is $\mathbb{E}[\langle \Delta r^2(\Delta t) \rangle] = 2\sigma_s^2 \Delta t$. For 2D diffusive motion, we thus expect $\langle r^2(\Delta t) \rangle = 4D_m \Delta t$, where D_m is the diffusion coefficient. D_m can be associated with the step size $D_m = \frac{\sigma_s^2}{2}$. For $\sigma_s^2 = 4$, and $D_m = 2$, the truth is $\langle r^2(\Delta t) \rangle = 4D_m \Delta t = 8\Delta t$.

This simulation represents an optically dense case with a large number of small particles, which can induce a large bias in estimating the noise (Table I). We demonstrate this by comparing the estimation of noise by $B = D_{\min}(\Delta t_{\min})$, $B = \langle D_o(q_{\max}, \Delta t) \rangle_{\Delta t}$, and $B = 2\langle |\hat{I}_o(q_{\max}, t)|^2 \rangle_t$ to the truth $B = 2\sigma_0^2$ (horizontal line) as shown in Fig. 5. We found that all three estimators overestimate the mean of B in the analysis. The first approach, $B = D_{\min}(\Delta t_{\min})$, has the smallest bias among the three, as the term $1 - f(q, \Delta t_{\min})$ in Eq. (4) is relatively small at the smallest Δt .

Next, we calculated the MSDs by DDM-UQ using each of the following ways for estimating the noise, as shown in Fig. 4: $B = D_{\min}(\Delta t_{\min})$ (red diamonds), $B = \langle D_o(q_{\max}, \Delta t) \rangle_{\Delta t}$ (purple squares), $B = 2\sigma_0^2$ (blue circles), and $B = 0$ (green triangles). The truth is plotted as the thick black line. Our results reveal that estimating the noise accurately is necessary to obtain an accurate estimate of MSD, whereas overestimating or underestimating the noise leads to an inaccurate estimate, particularly for sufficiently small Δt , where the displacements are smaller and therefore more strongly impacted by a poorly estimated noise term.

A simple way to detect the estimation bias of B is to plot $D_o(q, \Delta t)$ as a function of q for a given Δt (here selected to be Δt_{\min}) across all q values, as shown Fig. 6. If the signal

TABLE II. N-RMSE of simulated cases. The analysis mode with the lowest N-RMSE is shown in bold in each scenario.

Scenario	DDM [$A(q)$ from plateau]	DDM [$A(q)$ from $\langle \hat{I}_o(q, t) ^2 \rangle_t$]	DDM-UQ	MPT
Simple diffusion, $\sigma_s = 2$	0.109	0.047	0.025	0.024
Simple diffusion, $\sigma_s = 0.5$	0.396	0.034	0.057	0.129
Simple diffusion, $\sigma_s = 4$	0.130	0.095	0.014	0.492
Diffusion with drift	0.061	0.052	0.074	0.026
Diffusion with drift, optically dense	0.041	0.049	0.029	0.424
O-U process with drift	0.243	0.063	0.183	0.213

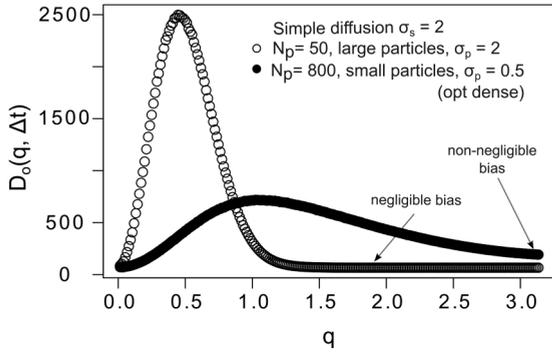


FIG. 6. A close look at the estimators of noise B_{est} in the first and second scenarios of simple diffusion with identical underlying dynamics $\sigma_s = 2$, above. The difference lies in that the second case includes $16\times$ as many particles while particle radius decreases by $4\times$ (filled circles). For the range of q probed, $D_o(q, \Delta t)$ does not decay to zero at q_{max} .

$A(q)[1 - f(q, \Delta t)]$ does not approach zero at the highest q values, then $D_o(q, \Delta t)$ continues to decrease as q increases. We found this to be the case for $D_o(q, \Delta t)$ for the optically dense case (with $N_p = 800$ probes, shown as filled circles, Fig. 6). Hence, estimating B using D_o at q_{max} introduces non-negligible bias in this example. In contrast, when there are only $N_p = 50$ particles (shown as open circles), the $D_o(q, \Delta t)$ approaches zero at high q and the bias for B is negligible across all methods of estimation (Fig. 5). To overcome the bias induced by the estimator, one may adjust the experimental conditions to avoid the scenarios summarized in Table I, or may attempt to measure the noise using a separate sample with probes immobilized in solid matrix under similar imaging conditions. Deriving a more accurate estimator of the noise at these experimentally challenging scenarios will be an interesting future direction.

C. Simple diffusion with different step size σ_s

To explore the effects of varying step size (which is a proxy for varying diffusivity) on our analysis, three additional scenarios with diffusive dynamics are explored, using simulations of particles taking different step sizes σ_s , but for which all other settings and conditions were held constant. When $\sigma_s = 2$, corresponding to an intermediate step size, all four methods (DDM with two different estimators, DDM-UQ and MPT) provide results that reasonably approximate the true values that are directly calculated from the inputted particle positions [Fig. 7(a)]. When the N-RMSE values are calculated and compared, we find that the results from MPT provide the best approximation of the true values for this case, whereas DDM-UQ provide nearly the same level of accuracy as MPT (Table II).

We next calculated the MSDs for simple diffusion with lower ($\sigma_s = 0.5$) and higher ($\sigma_s = 4$) step sizes [Figs. 7(b) and 7(c)]. The largest differences are observed at high step size [Fig. 7(c)] where particle displacements are large ($\sigma_s = 4$). In this limit, DDM-UQ outperforms MPT by a large margin (Table II). The reason is intuitive for MPT: as particle displacement becomes large, the likelihood of two or more particles exchanging positions within the search radius

increases significantly. This can lead to the algorithm misidentifying the particle, thereby resulting in erroneous linking of the trajectories.

D. Diffusion with drift

We next consider particles subjected to diffusive-convective motion (i.e., “drift”). At each time step, a particle moves by $\Delta x_{i,j} \sim \mathcal{N}(\mu_D, \sigma_s^2)$, resulting in a random walk with step size $\sigma_s = 0.56$ superimposed on a deterministic drift with mean velocity $\mu_D = 0.1$ in units of pixels and time step in the same direction for all particles. The expected value of the MSD in this case is $\mathbb{E}[\langle \Delta r^2(\Delta t) \rangle] = 2\sigma_s^2 \Delta t + 2\mu_D^2 \Delta t^2$. At short Δt , the motion is primarily diffusive [Fig. 7(d)], $\langle \Delta r^2(\Delta t) \rangle \sim \Delta t$ with a transition to convective motion $\langle \Delta r^2(\Delta t) \rangle \sim \Delta t^2$, at large Δt . This results in an increasing value of $\frac{d\langle \Delta r^2(\Delta t) \rangle}{d\Delta t}$ with Δt , and permits a measure of the relative strength of the two dynamic processes through measure of the local slope on a log-log scale.

We also compare the performance of the different analytical routines under differing initial conditions. In particular, we vary the initial positions where particles were released at $t = 1$, holding all other settings equal, to compare an optically dilute scenario [Fig. 7(d)], where particles were uniformly distributed throughout the simulation box, and an optically dense scenario [Fig. 7(e)], where particles were released from a small $\frac{L}{20} \times \frac{L}{20}$ square in the middle of the frame causing them to be near each other and even overlap for some frames. When particles are evenly distributed in the simulation box, all methods closely track the truth, with MPT having the lowest RMSE in the optically sparse scenario, whereas DDM-UQ performs the best in the optically dense scenarios. The optically dense scenario mimics situations where a high concentration of particles is present, which is known to lead to tracking issues in MPT since it is often difficult to distinguish particles in close proximity. By contrast, DDM and DDM-UQ perform strongly in this limit (Table II). The range of Δt that can be resolved by any method decreases as compared to the dilute case, due to the large variability of MSD at different wave vectors in this scenario.

E. Ornstein-Uhlenbeck process with drift

Finally, we simulated particles from an Ornstein-Uhlenbeck (O-U) process with drift. Such a process mimics thermally driven particle motion in an effective elastic medium with drifts distinct to each particle. The convective term is constant in magnitude μ_D , and fixed in direction θ for an individual particle, but randomized for all particles. A pure O-U process without a convective term can lead to a $\langle \Delta r^2(\Delta t) \rangle$ that plateaus at a certain Δt , and thus $f(q, \Delta t)$ does not decorrelate for some finite values of Δt . Adding a convective term leads to complete decorrelation and allows the dynamics at these Δt 's to be captured. Moving the sample to achieve ensemble averaging for sample that manifested constrained heterogeneity has previously been applied to light scattering on polymer gel samples [49].

Here, just like the original O-U process, successive steps have a weak correlation with previous steps:

$$X_{i,j}(t + 1) = \rho X_{i,j}(t) + \epsilon_{i,j,t}, \tag{26}$$

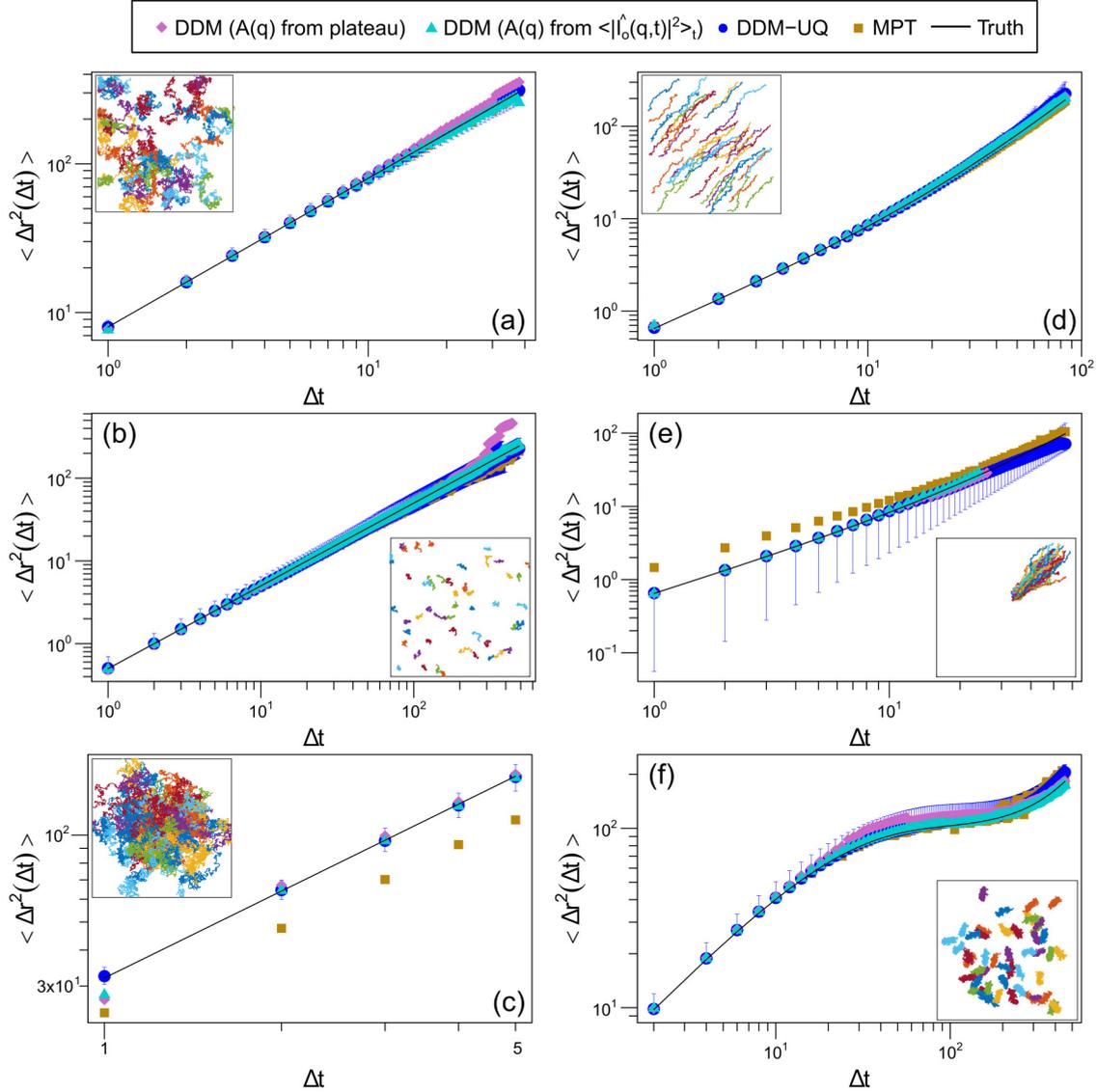


FIG. 7. Comparisons of MSDs calculated using DDM-UQ, DDM, and MPT for various simulated scenarios: (a)–(c) Simple diffusion with (a) $\sigma_s = 2$, (b) $\sigma_s = 0.5$, and (c) $\sigma_s = 4$, corresponding to intermediate, low, and high step sizes. (d), (e) Diffusion with drift in (d) optically dilute and (e) optically dense samples, and (f) diffusion of a particle within a harmonic potential well (Ornstein-Uhlenbeck process) subjected to drift. The pink diamonds and the cyan triangles indicate mean values obtained by estimating $A(q)$ from the plateau in $D(q, \Delta t)$, or from $\langle |\hat{I}_o(q, t)|^2 \rangle_t$, respectively, using DDM algorithm based on all values of $D(q, \Delta t)$. The blue circles and error bars depict the mean and 95% predictive interval estimated by DDM-UQ based on 1% of the $D(q, \Delta t)$, respectively. The golden squares and black solid line represent the output of MPT analysis and the true values as directly calculated from the known particle positions, respectively. The generated particle trajectories are shown in the insets, in a simulation box of 480×480 pixels.

where $\epsilon_{i,j,t} \sim \mathcal{N}(0, \sigma_s^2(1 - \rho^2))$, with $\rho = 0.95$, $\sigma_s = 5$, and $X_{i,j}(t) = x_{i,j}(t) - (t - 1)\mu_{ij} - x_{i,j}(1)$ for any t ; the indices represent the j th particle and i th direction ($i = 1, 2$) and $\mu_{1j} = \mu_D \cos\theta_j$, $\mu_{2j} = \mu_D \sin\theta_j$, respectively, with $\mu_D = 0.02$. Each particle's initial position was generated by a normal distribution centered around $x_{ij}(t_0)$ with variance σ_s^2 , i.e., $x_{ij}(t_1) \sim \mathcal{N}(x_{ij}(t_0), \sigma_s^2)$, where $x_{ij}(t_0)$ are randomly distributed within a square $\frac{3L}{4} \times \frac{3L}{4}$ in the middle of the simulation box to reduce the likelihood that a particle moves out of the frame during the simulation. The motion is subject to an attractive potential towards $x_{ij}(t_0) + (t - 1)\mu_{ij}$ with drift $(\mu_D \cos(\theta_j), \mu_D \sin(\theta_j))^T$ at the t th time point for the j th particle. The expected

value of MSD for O-U process with drift is $\mathbb{E}[\langle \Delta r^2(\Delta t) \rangle] = 4\sigma_s^2(1 - \rho^{\Delta t}) + \mu_D^2 \Delta t^2$. There is a diffusive contribution that dominates at sufficiently small Δt from the first term $1 - \rho^{\Delta t} \approx 1 - [1 + \ln(\rho)\Delta t] = \ln(\rho^{-1})\Delta t$, by Taylor expansion at small Δt . The second convective term captures the convective flow, and can represent the type of dynamics observed in some actively driven systems [20,50].

The process results in a trace of MSD versus Δt with multiple inflection points: $\langle \Delta r^2(\Delta t) \rangle$ first grows with decreasing slope until the plateau value is reached ($\frac{d\langle \Delta r^2(\Delta t) \rangle}{d\Delta t} \approx 0$); at much longer Δt 's, the MSD will grow with an increasing slope until eventually the convective motion dominates

($\frac{d\langle\Delta r^2(\Delta t)\rangle}{d\Delta t} = 2$). In contrast to all previous scenarios, we find that $D(q, \Delta t)$ exhibits nonmonotonic behavior at some wave vectors, and that the MSD shows apparent q dependence over Δt for many intermediate q values. Specifically, a local maximum in $D(q, \Delta t)$ appears before $D(q, \Delta t)$ reaches the plateau at long Δt . Such nonmonotonic behavior in the resulting $f(q, \Delta t)$ has previously been observed in a system consisting of self-catalytic Janus particles [15].

We find that all approaches are able to capture the general trends and magnitude of motion [Fig. 7(f)]. At short Δt , the response of all four methods is similar. We note that this scenario constitutes an “ideal” case for MPT: the particle locations are sparse, and each particle moves toward their respective attractive centers $x_{ij}(t_0)$, never to cross paths with one another. Yet, DDM-UQ still outperforms MPT in this case, even though it can be shown that some marginal benefit can be gained by using the full data set. Still, with limited sampling, DDM-UQ has performance on par with DDM and MPT, evaluated by the N-RMSE in Table II.

Overall, we find that DDM-UQ accurately determines $\langle\Delta r^2(\Delta t)\rangle$ for a range of experimentally relevant scenarios based on a fraction of the observations, which substantially reduces the computational cost. Note that DDM-based algorithms are more automated compared to MPT, as DDM does not require manually chosen inputs of model parameters such as particle sizes and search radius. On the other hand, MPT can provide more reliable dynamic information at larger Δt than the current model-free DDM-based algorithms. The DDM-UQ algorithm developed in this study enables a model-free automated DDM-based analysis with results that are comparable to MPT with optimized settings, with less computational cost and tuning parameters. Indeed, DDM-UQ even outperforms MPT for certain challenging experimental scenarios (e.g., high concentration, fast moving objects, etc). Furthermore, many other scenarios abound where the ISF $f(q, \Delta t)$ rather than the MSD can provide physical insight to the system, and the DDM-UQ algorithm provides an automated, model-free estimation of the ISF. These findings affirm the sensitivity of this ensemble-based method as well as the need for an unbiased estimator of noise and other model parameters, and validates our data reduction approach.

IV. ANALYSIS OF EXPERIMENTAL DATA

A. Newtonian fluid

We first measured the properties of a Newtonian fluid in which we expect simple diffusive particle dynamics. Experimentally, we suspended fluorescent polystyrene microspheres of diameter $2a = 100$ nm (yellow-green with excitation maxima of $\lambda_{ex} = 441$ nm and emission maxima at $\lambda_{em} = 485$ nm, Polysciences, Warrington, PA) in a 30 wt.% sucrose solution (Sigma-Aldrich, St. Louis, MO) at a particle volume fraction $\phi \approx 3 \times 10^{-5}$. This composition was previously studied by dark-field DDM [31], and the exact viscosity value of the sucrose is well documented [51]. The particle suspension is introduced into a homemade sample cell formed using a glass slide and glass cover slip separated by 100 μm spacers. The sample is imaged in epifluorescence using an Olympus IX73 inverted microscope, outfitted with a halogen lamp with

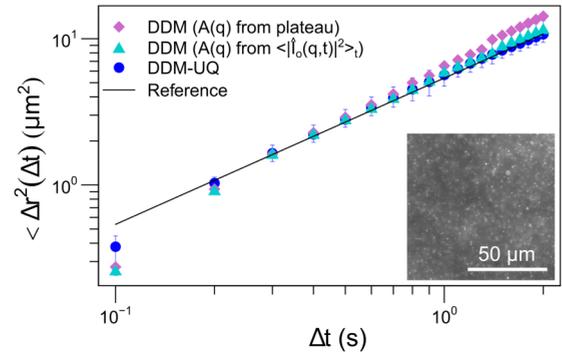


FIG. 8. Mean squared displacements estimated from the motions of 100 nm diameter probe particles in a 30 wt.% sucrose solution, which serves as a model Newtonian fluid. The pink diamonds and the cyan triangles indicate MSD obtained by using $A(q)$ determined from the plateau in $D_o(q, \Delta t)$, or from $\langle|\hat{l}_o(q, t)|^2\rangle_t$, respectively, based on all values of $D(q, \Delta t)$. Blue circles denote results obtained using the DDM-UQ analysis by a small fraction of the data; in this case $A(q)$ was only estimated using $\langle|\hat{l}_o(q, t)|^2\rangle_t$. The black solid line denotes reference values and is calculated using the known viscosity (at 20 °C) and the Stokes Einstein equation. The inset shows a single experimental image of the movie, where particles appear to be grainy and cannot be individually resolved.

green fluorescent protein (GFP) filter set ($\lambda_{ex} = 457\text{--}487$ nm, $\lambda_{em} = 502\text{--}538$ nm), using a 40 \times objective (NA = 0.6), which provides a spatial resolution of 97 nm/pixel. Images are collected using a 8-bit Point Grey Chameleon USB camera using a 100 ms exposure time, 10 Hz frame rate, and 960 pixel \times 960 pixel frame size.

In this experiment, the limited resolution of fluorescence microscopy relative to the particle size precludes identification of individual particles by MPT [see Fig. 8(a), inset], and thus prevents MPT analysis. Despite the lack of particle-level information, DDM is nevertheless capable of detecting the minute differences in image intensities due to particle motion, and recovers the correct diffusive dynamics [Fig. 8(a)]. Thus, DDM shows extraordinary sensitivity even when the particle size is below the diffraction limit of the microscope.

The reference value of the MSD determined by the Stokes-Einstein relation [31] is reported by the solid black line. We generally find quantitative agreement by both DDM and DDM-UQ over most of the measured Δt 's, with noticeable deviations from the linear trend and the expected values at the smallest Δt (Fig. 8). Here the effects of using the different estimators of noise are very small, as $D_o(q, \Delta t)$ levels off at high q values for any given Δt . When the approaches are compared, DDM-UQ (blue diamonds), which stabilizes large fluctuations through the use of the predictive median, leads to more accurate estimates of the MSD. Furthermore, we found that estimating $A(q)$ from $A_{\text{est}}(q) + B_{\text{est}} = \langle|\hat{l}_o(q, t)|^2\rangle_t$ (blue circles and cyan triangles) improves the estimate of the MSD at large Δt over cases where A is estimated from the plateau values only (pink diamonds). We attribute this improvement to averaging more $D_o(q, \Delta t)$'s from more q 's in determining the ISF, regardless of whether there is a plateau or not at the finite Δt that are accessible in experiment.

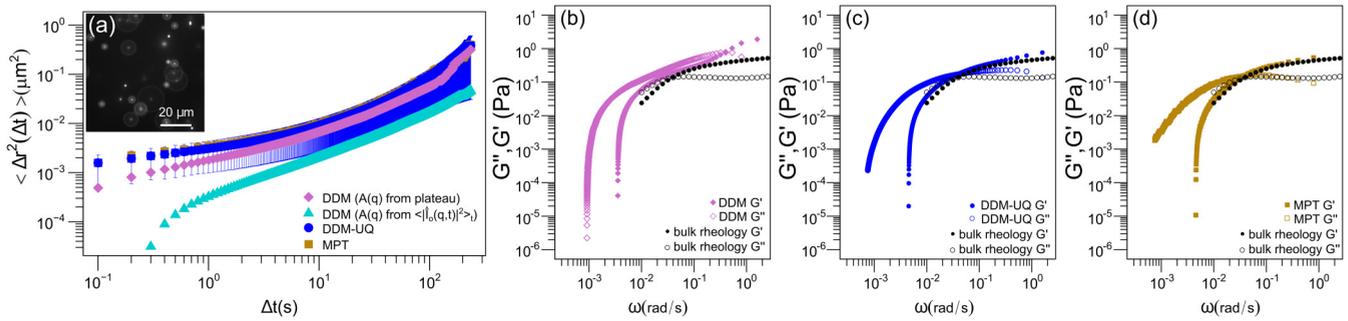


FIG. 9. Results of microrheology and bulk rheology measurements of solutions wormlike micelles, which form viscoelastic fluids. (a) Mean squared displacements obtained from full $D_o(q, \Delta t)$ with $A(q)$ estimated from the plateau in $D(q, \Delta t)$ (pink diamonds), from $\langle |\hat{L}_o(q, t)|^2 \rangle_t$ (cyan triangles), DDM-UQ (blue circles), and MPT (golden squares). The inset shows an experimental snapshot of the microrheology experiment. (b)–(d) Comparison of the frequency-dependent linear viscoelastic moduli obtained either from bulk rheology experiments (black symbols) or calculated from the MSDs obtained by either using (b) DDM with $A(q)$ estimated from $\langle |\hat{L}_o(q_{\max}, t)|^2 \rangle$ (pink diamonds), (c) DDM-UQ (blue circles), or (d) MPT (golden squares). Solid symbols denote the storage [elastic, $G'(\omega)$] modulus while open symbols denote the loss [viscous, $G''(\omega)$] modulus. MSDs estimated by MPT and by DDM-UQ nearly overlap.

B. Viscoelastic fluid

We next investigate the performance of DDM in probing the dynamics of a non-Newtonian fluid, namely, a viscoelastic wormlike micelle solution of 12.5 mM sodium salicylate (NaSal; Sigma-Aldrich, St. Louis, MO) and 15 mM cetylpyridinium chloride (CPyCl; Sigma-Aldrich, St. Louis, MO) that forms an entangled network. To this solution, fluorescent polystyrene microspheres of diameter $2a = 1500$ nm (carboxylated yellow-green ex/em = 505/515, Life Technologies, Carlsbad, CA) are added at a volume fraction $\phi \approx 2 \times 10^{-4}$. The sample is mixed and allowed to relax overnight prior to loading into a capillary tube, which is sealed on both sides with optical glue (Norland Products, Inc.) and cured under a UV lamp. We note that this sample is similar in composition, but not identical, to a solution characterized by DDM microrheology in previous work [6]. The sample is imaged using a Zeiss Axio Observer 7 microscope in fluorescence mode using a Colibri 7 light source, standard GFP filter sets and a $40\times$ water-immersion objective lens (NA = 1.2), which provides a magnification of 150 nm/pixel. Images were recorded with an Axiocam 702 monochromatic camera using 15 ms exposure time, 10 Hz frame rate, and 512×512 pixels frame size. In this case, the reference data set is obtained by a bulk rheology measurement of an identical sample without tracer particles using an AR-G2 stress-controlled rheometer (TA Instruments, New Castle, DE) to perform a frequency sweep in the linear viscoelastic limit using a 40-mm diameter cone-and-plate fixture, with a 2° cone angle and a $55 \mu\text{m}$ truncation, at 2% shear strain over a frequency range of 0.01–10 rad/s. The instrument is outfitted with a solvent trap to minimize evaporation during testing.

Wormlike micelles (WLMs) manifest complex frequency-dependent viscoelasticity that nonetheless follow simple scaling laws [6,52]. Such a system is challenging to characterize: small probe displacements at low Δt make it difficult to determine if the flattening of the MSD at low Δt is characteristic of system behavior or a result of “pixel biasing” due to particle localization error [13]. The slow dynamics also contributes to the sensitivity of q selection. In this case,

solidlike behavior at high frequency (low Δt) is confirmed by bulk rheometry measurements [Figs. 9(b)–9(d)] and thus a “flattening” of the MSD trace is expected at small Δt . With this information, we evaluate the four methods.

In this experiment, the noise estimators $B_{\text{est}} = \langle D_o(q_{\max}, \Delta t) \rangle_{\Delta t}$ and $B_{\text{est}} = D_{\min}(\Delta t_{\min})$ produce similar results by DDM, although neither method performs well because the estimation error for $A(q)$ at high values of wave vector is relatively large due to the small displacements, which approach the resolution limit at small lag times.

By contrast, the DDM-UQ algorithm using the median from the predictive sampling based on $D_o(q, \Delta t)$ on moderately large wave vectors is more robust than using a simple ensemble of $D_o(q, \Delta t)$ in DDM. The MSD trace for DDM-UQ is very close to the MPT result, and is more consistent with the Maxwell fluidlike behavior [Fig. 9(a)]. Note that we only compute the Fourier transformation of intensity difference at 25 selected Δt 's in DDM-UQ analysis, instead of 6000 Δt points in DDM, which reduces the computational

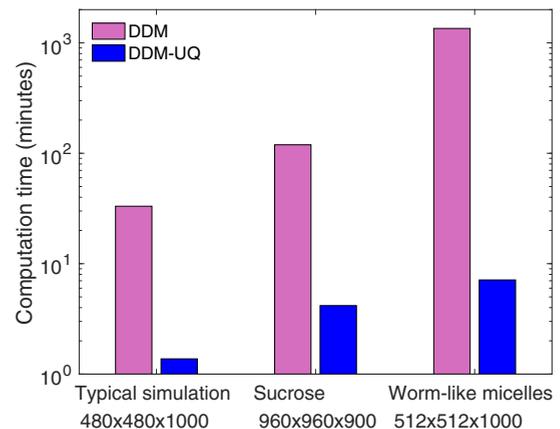


FIG. 10. Pair bar graphs demonstrating the computation efficiency of DDM-UQ scheme (blue) over current DDM approaches (pink). The size of each image stack is labeled underneath the data set, in terms of frame size \times time points ($L_x \times L_y \times T$).

cost by more than 100 times (shown in Fig. 10) while providing more accurate results.

Next we convert the measured MSD data into measures of the frequency-dependent viscoelastic moduli using the generalized Stokes Einstein relation (GSER) [53]:

$$|G^*(\omega)| \approx \frac{k_B T}{\pi a \langle \Delta r^2(1/\omega) \rangle \Gamma[1 + \alpha(\omega)]}, \quad (27)$$

where T is the temperature, k_B is the Boltzmann constant, and $\alpha(\omega)$ the power-law slope [on a log-log plot of $\langle \Delta r^2(\Delta t) \rangle$].

The procedure of determining the power law slope $\alpha(\omega)$ usually involves first taking the numerical derivative of $\langle \Delta r^2(\Delta t) \rangle$ with respect to Δt , and then fitting a polynomial of the data around a particular Δt , which is then Laplace transformed to frequency space [53]. As shown in Figs. 9(b)–9(d), the moduli we measured directly by bulk rheology generally agree with those computed from $\langle \Delta r^2(\Delta t) \rangle$ using microrheology approaches, even at high frequency, in both magnitude and in the frequency of the crossover. The result from DDM with $A(q)$ estimated from the plateau is shown in pink diamonds [Fig. 9(b)], the result using DDM-UQ is shown in blue circles [Fig. 9(c)], and MPT is shown in golden squares [Fig. 9(d)]. The MSD by DDM with $A(q)$ estimated from the relation $A_{\text{est}}(q) + B_{\text{est}} = \langle |\hat{I}_o(q, t)|^2 \rangle_t$ (cyan triangles) contains very large error, and thus it fails to estimate moduli, so the result is not shown here.

Both the frequency dependence and magnitude of the moduli in the high-frequency regime are extremely sensitive to the MSD at low Δt . Nevertheless, MPT and DDM-UQ are in an approximate agreement with the values obtained with macroscale rheology at higher frequencies. By contrast, in a slow moving system, or at large q 's (which correspond to small displacements), when the movement is less than a pixel on average over Δt , it is not possible to capture the system dynamics by calculating the image difference. As a result, estimates of the MSD from the $D_o(q, \Delta t)$ tend to provide underestimates as compared to the true values at small Δt .

We note that the numerical differentiation of $\langle \Delta r^2(\Delta t) \rangle$ introduces a high degree of uncertainty into the moduli that is not represented on Fig. 9. A more robust estimation method for the moduli is an area of future research.

C. Model fitting for actively driven systems

Beyond thermally driven dynamics and microrheology, DDM can be applied in the realm of non-Brownian, active systems where models of $f(q, \Delta t)$ can provide insight into the physics associated with the system dynamics. In such cases, the time variation of the image structure function does not arise from probe motion but from structural evolution of actively powered components within the system such as migrating bacteria [8] or advection due to internal stresses that arise from phase separation and aging [10], among other examples. Here we demonstrate that DDM-UQ analysis can be applied to such anomalous dynamics as well. Specifically, the statistical approach we have developed should be applicable to any system that sufficiently decorrelates over the experimental observation time, thus greatly reducing computational time and increasing the robustness of model fits.

As an example, we consider the dynamics of actively driven composite cytoskeletal networks of actin and microtubules, by (re)analyzing the experimental data recently reported by Lee *et al.* [20]. Actin and microtubules are ubiquitous and essential in eukaryotic cells, and *in vitro* networks of the purified filamentous proteins are widely studied for their potential to self-organize and form model nonequilibrium materials when acted upon by ATP-driven molecular motors such as myosin. In recent work, Lee *et al.* [20] investigated a composite network comprising actin and microtubules, which were each labeled with distinct fluorophores, and acted upon by myosin. DDM was then used as a means of disentangling the motions recorded through the individual fluorescence channels, allowing investigation of the mechanisms of cross correlation of actin and microtubule dynamics within the entangled network. By contrast, MPT can provide only information on the bulk network itself.

Here we follow [20] to model the dynamics of such an active system by a stretched exponential model [20]:

$$f(q, \Delta t) = \exp\left[-\left(\frac{\Delta t}{\tau(q)}\right)^{\gamma(q)}\right], \quad (28)$$

where $\gamma(q) > 1$ means the system shows contractile dynamics, while $\gamma(q) < 1$ when the system shows stretching dynamics. A relaxation time scaling where $\tau(q) = \frac{1}{vq}$ describes a system exhibiting ballistic motion with velocity v , whereas $\tau(q) = \frac{1}{D_m q^2}$ describes a system exhibiting diffusive motion where D_m represents the diffusion coefficient.

Given the intrinsically heterogeneous nature of such systems, multiple replicates of the same composition are often examined, increasing the already heavy computational costs that are typical of DDM analysis. To demonstrate this, we reanalyze six replicates of the active actin data set from [20] to show that despite computing the Fourier transformation of the intensity difference at only 25 selected Δt 's, which significantly reduces the computational cost, DDM-UQ extracts the information that is as accurate as the DDM approach that was originally employed. In detail, the full $D_o(q, \Delta t)$ is computed directly from the image stacks by DDM, from which a subset of values $D_o(q, \Delta t)$ are preselected to obtain the predictive distribution by DDM-UQ.

This example specifically compares the DDM and DDM-UQ analysis when a parametric model with four fitting parameters are specified. When a stretched exponential model [Eq. (28)] for $f(q, \Delta t)$ with coefficients $\tau(q)$ and $\gamma(q)$, along with $A(q)$ and $B(q)$, is fit to either the full matrix $D_o(q, \Delta t)$ (DDM) or the predictive distribution $D(q, \Delta t)$ (DDM-UQ) for each q , the estimated image structure function $D_e(q, \Delta t)$ is obtained. $D(q, \Delta t)$ contains the same number of entries as $D_o(q, \Delta t)$; it is reconstructed using the mean of values sampled from the predictive distribution $D(q, \Delta t)$ [300 instances at every $(q, \Delta t)$]. The purpose of this step is to extract coefficients $\tau(q)$ and $\gamma(q)$ relevant to the underlying physical process. Here for DDM, we use the conventional approach by minimizing the square error loss between the model and the $D(q, \Delta t)$, which is used as the loss function, whereas for DDM-UQ, we minimize the weighted square loss where the weights are calculated from the inverse variance of the predictive samples from GPR.

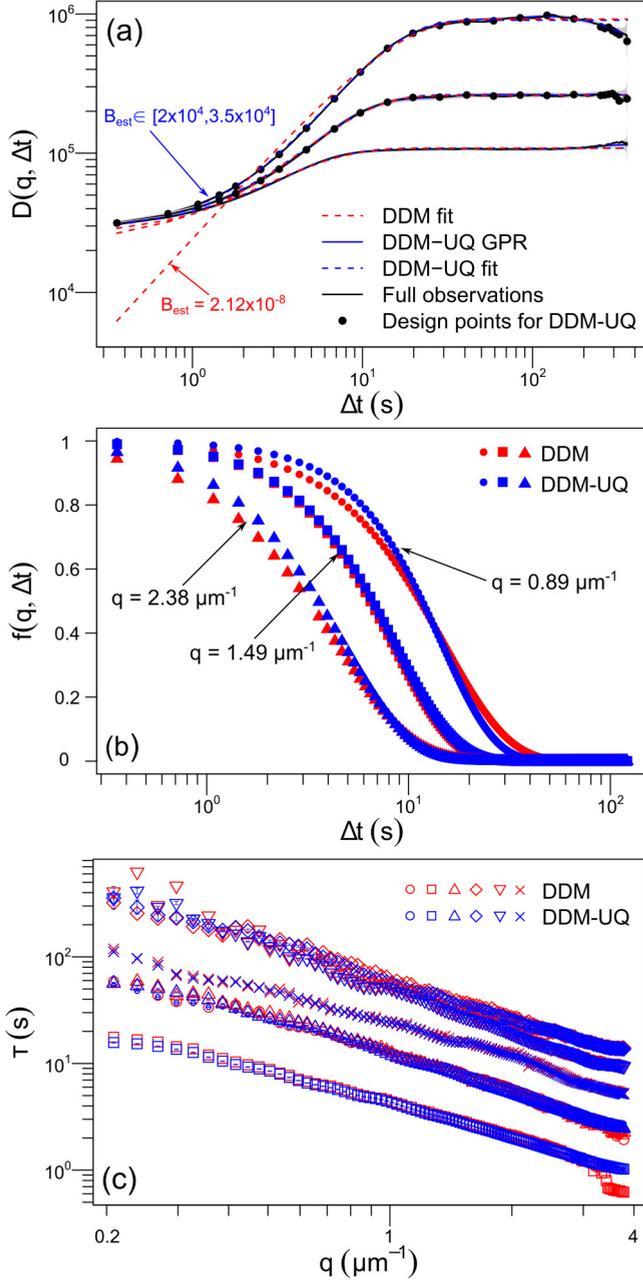


FIG. 11. Example of analysis of an actively driven system. Experimental data extracted from a fluorescently labeled actin-myosin-microtubule composite network system reported in [20], processed using DDM (red symbols) or DDM-UQ (blue symbols), and fit to a stretched exponential model to describe the system dynamics [Eq. (28)]. (a) Comparing DDM and DDM DQ fits to the original $D_o(q, \Delta t)$ matrix, where the uncertainty is denoted by the gray shaded area, which is very small as the uncertainty is low. The solid line denotes full $D_o(q, \Delta t)$ data, while the solid dots denotes $D_o(q, \Delta t)$ selected to obtain the predictive distribution. (b) Fits to $f(q, \Delta t)$ shown at several different q 's. (c) Parameter τ , which describes the system relaxation time, plotted as a function of q . Different symbols represent τ extracted from six different data sets. From this, the system velocity is obtained (Fig. 12).

The difference between the estimated quantity $D_e(q, \Delta t)$ from fitting stretched exponential model and observed quan-

TABLE III. N-RMSE of active actin networks.

Sample ID	<70% Plateau		Full data	
	DDM	DDM-UQ	DDM	DDM-UQ
1	0.0576	0.0417	0.0700	0.0507
2	0.0641	0.0689	0.0882	0.0948
3	0.0406	0.0265	0.0573	0.0373
4	0.0759	0.0699	0.0835	0.0769
5	0.0462	0.0285	0.0576	0.0355
6	0.0588	0.0317	0.0724	0.0390

tity $D_o(q, \Delta t)$ is evaluated by the N-RMSE:

$$\text{N-RMSE} = \frac{\sqrt{\frac{1}{n_{q\Delta t}} \sum_{\Delta t \in \Delta \mathcal{T}} \sum_{q \in \mathcal{Q}} [\langle \tilde{D}_o(q, \Delta t) \rangle - \langle \tilde{D}_e(q, \Delta t) \rangle]^2}}{\tilde{\sigma}_D}, \quad (29)$$

where $\tilde{D}_o(q, t)$ and $\tilde{D}_e(q, \Delta t)$ are the logarithms of the observed and estimated image structure function by different approaches, and $\tilde{\sigma}_D$ is the logarithm of sample standard deviation; $\Delta \mathcal{T}$ and \mathcal{Q} are the sets of Δt and q available for comparison, respectively.

Figure 11(a) shows reconstructed $D(q, \Delta t)$ (dashed lines) from fitting the full observation (solid line) using DDM as well as a reconstructed $D(q, \Delta t)$ obtained by resampling with only a fraction of design points (black dots) using DDM-UQ. There is general agreement between the two approaches at all q values, with some differences observed at short times because DDM-UQ weighs more heavily data at small Δt , as the predictive variance is small at these regions. Fitting via a weighted least squares minimization approach is more robust for estimating the noise term B . One $B(q)$ is estimated to be very close to 0 (2.13×10^{-8}) in DDM, whereas it is estimated to be around 3.3×10^4 in DDM-UQ. The significant underestimation of the noise by DDM explains the large deviation of the fit at this wave vector shown in Fig. 11(a). This example illustrates the importance of estimating the noise parameters accurately, and that fitting a parametric model of $f(q, \Delta t)$ without addressing the uncertainty by the least squared estimator can be unreliable in estimating the noise parameter.

In Fig. 11(b) we show the same fits of the stretched exponential model used to reconstruct $f(q, \Delta t)$ at different q 's using the full observed image structure function $D_o(q, \Delta t)$ obtained by the DDM (red lines) and DDM-UQ analysis (blue lines). Note that DDM-UQ only uses observations $D_o(q, \Delta t)$ at selected q and Δt , but it performs equally well even for unobserved q values for which there is no observation for any Δt [see Fig. 11(a), bottom curve].

The differences between the estimated and observed values of $D_o(q, \Delta t)$ for DDM and DDM-UQ are quantified by the N-RMSE in Eq. (29) as shown in Table III. Since $D(q, \Delta t)$ at large Δt contains fewer independent samples and shows large fluctuations, it is informative to compare the accuracy of the fit only up to a threshold value, chosen here to be 70% of the Δt values before the plateau is reached. In all cases, DDM-UQ outperforms DDM in more closely approximating the $D_o(q, \Delta t)$ when comparing the truncated data set, and

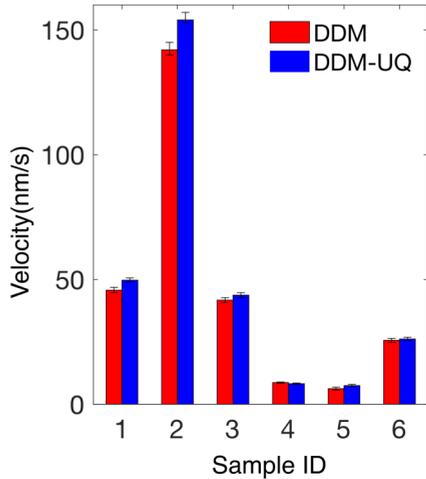


FIG. 12. Velocity estimates and confidence intervals extracted from fitting the stretched exponential model to either the full observations (DDM) or to reconstructed $D(q, \Delta t)$ from selected design points (DDM-UQ). The error bars denote 95% confidence intervals.

in almost all cases the fit of DDM-UQ, which has much lower computational cost, is comparable to that of DDM, as shown in Table III. Thus, DDM-UQ significantly accelerates the analysis without any observable sacrifice in accuracy with respect to postprocessing of the data such as model fitting.

As described in Eq. (28), $f(q, \Delta t)$ contains the fit parameters $\gamma(q)$ and $\tau(q)$. Following [20], we fit the data to a linear model $\tau(q) = \frac{1}{vq}$ and extract the characteristic velocities of the active actin mixture [Fig. 11(c)]. The maximum likelihood estimator (MLE) and the confidence interval (CI) for the velocities from different replicates are tabulated in Fig. 12. Importantly, DDM-UQ with its limited observations largely recovers similar characteristic velocities and confidence intervals as those obtained using the full matrix, paving the way for high-throughput analysis of the dynamic properties of complex biomaterial systems.

V. CONCLUDING REMARKS

DDM can be applied to a structurally evolving image stack to obtain the image structure function and intermediate scattering function. It provides an aggregated measure of dynamics, potentially offering higher accuracy in extracting physical quantities than using real-space data alone. While the theoretical framework of DDM is well established, to our knowledge, this work represents an exploration into propagating the uncertainty associated with measurement noise, and analyzing the effects of noise in parameter estimation through mathematical and numerical analysis.

Based on error propagation in estimating the image structure function, we derived the mean and variance of the noise term B , leading to more accurate estimation of the ISF and MSD at small Δt .

Moreover, we showed that only a small subset of $D_o(q, \Delta t)$ (around 0.5%–5%) at selected q and Δt need to be computed, and when they are used in a GPR model, it is possible to obtain the predictive median and samples in the image structure function at unobserved inputs and subsequent quantities of

interest could be robustly predicted. Both simulations and experiments were presented to demonstrate that our method has virtually no loss of information, while reducing the computational time by 25–120 times. The combined improvements offered by the error propagation and predictive median from GPR results in more robust estimation of the intermediate scattering function and mean squared displacements. Through the comparisons made here between DDM-UQ, other formulations of DDM, and MPT, we highlight the need for accurate noise estimation in the analysis and interpretation of DDM experiments.

We anticipate that these results will enable many new applications of DDM to complex biomaterial and soft material systems [54,55]. With the potential to carry out real-time analysis via downsampling, the proposed method can be extended to map out an entire phase space of material composition or physicochemical conditions in a high-throughput manner. This increased performance also places new demands on the general applicability of the algorithm, for instance, to provide meaningful analysis of stiffer materials that do not fully decorrelate as quickly as a more fluidlike samples, as well as heterogeneous samples, through analysis of subpopulations that demonstrate distinct features. Future extensions of DDM-UQ analysis should include reducing the selection bias by properly weighting the ISF by the inverse variance of the noise, which should provide a more reliable and fully automated estimation of physical quantities at larger lag times. Another potential direction is to derive a more robust estimator of the imaging noise σ_0^2 from the image stack that can contain less bias to some challenging experimental scenarios (summarized in Table I). These directions will be pursued in future work.

ACKNOWLEDGMENTS

This work was supported by the BioPACIFIC Materials Innovation Platform of the National Science Foundation under Award No. DMR-1933487 (NSF BioPACIFIC MIP), with partial support by the Materials Research Science and Engineering Center (MRSEC) Program of the National Science Foundation under Award No. DMR-1720256 (IRG-3). M.G. acknowledges partial support from the National Science Foundation under Award No. DMS-2053423. M.E.H. acknowledges partial support from the National Science Foundation under Award No. CBET-1729108. This work used computational facilities purchased with funds from the National Science Foundation (Grant No. CNS-1725797) and administered by the Center for Scientific Computing (CSC). The CSC is supported by the California NanoSystems Institute and MRSEC (NSF Grant No. DMR-1720256) at UC Santa Barbara. We thank G. Lee and R. Robertson-Anderson from University of San Diego for providing the active actin data set.

M.G. and Y.L. contributed equally to this work.

APPENDIX A

Proof of Eqs. (6) and (7). The observed difference of the intensity at two time points $(t + \Delta t)$ and t can be described as

$$\Delta I_o(\mathbf{x}, t, \Delta t) = \Delta I(\mathbf{x}, t, \Delta t) + \Delta \epsilon(\mathbf{x}, t, \Delta t),$$

where $\Delta I(\mathbf{x}, t, \Delta t) = I(\mathbf{x}, t + \Delta t) - I(\mathbf{x}, t)$ and $\Delta \epsilon(\mathbf{x}, t, \Delta t) = \epsilon(\mathbf{x}, t + \Delta t) - \epsilon(\mathbf{x}, t)$. We denote the minimum time interval by $\Delta t_{\min} = 1$ and $\Delta t = l \Delta t_{\min} = l$, where l is a positive integer smaller than T .

We apply 2D discrete Fourier transformations on $\Delta I_o(\mathbf{x}, t, \Delta t)$ and obtain

$$\begin{aligned} & |\Delta \hat{I}_o(\mathbf{q}, t, \Delta t)|^2 \\ &= \frac{1}{N^2} \left| \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta I_o(\mathbf{x}, t, \Delta t) \exp\left(-\frac{i2\pi \mathbf{x}^T \mathbf{q}}{N}\right) \right|^2 \\ &= \frac{1}{N^2} \left\{ \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta I_o(\mathbf{x}, t, \Delta t) \cos\left(\frac{2\pi \mathbf{x}^T \mathbf{q}}{N}\right) \right\}^2 \\ &\quad + \frac{1}{N^2} \left\{ \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta I_o(\mathbf{x}, t, \Delta t) \sin\left(\frac{2\pi \mathbf{x}^T \mathbf{q}}{N}\right) \right\}^2 \\ &= \Delta \hat{I}_{o,1}^2(\mathbf{q}, t, \Delta t) + \Delta \hat{I}_{o,2}^2(\mathbf{q}, t, \Delta t), \end{aligned}$$

where $\hat{I}_{o,1}(\mathbf{q}, t, \Delta t)$ and $\hat{I}_{o,2}(\mathbf{q}, t, \Delta t)$ are independent from each other by the orthogonality of the Fourier basis with

$$\mathbb{E}[\hat{I}_{o,1}(\mathbf{q}, t, \Delta t)] = \frac{1}{N} \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta I(\mathbf{x}, t, \Delta t) \cos\left(\frac{2\pi \mathbf{x}^T \mathbf{q}}{N}\right),$$

$$\mathbb{E}[\hat{I}_{o,2}(\mathbf{q}, t, \Delta t)] = \frac{1}{N} \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta I(\mathbf{x}, t, \Delta t) \sin\left(\frac{2\pi \mathbf{x}^T \mathbf{q}}{N}\right),$$

$$\mathbb{V}[\hat{I}_{o,1}(\mathbf{q}, t, \Delta t)] = \mathbb{V}[\hat{I}_{o,2}(\mathbf{q}, t, \Delta t)] = \sigma_0^2.$$

Furthermore

$$\begin{aligned} |\Delta \hat{I}_o(\mathbf{q}, t, \Delta t)|^2 &= |\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 + 2\Delta \hat{I}(\mathbf{q}, t, \Delta t) \\ &\quad \times \Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t) + |\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)|^2, \end{aligned}$$

where

$$|\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 = \Delta \hat{I}_1^2 + \Delta \hat{I}_2^2, \quad (\text{A1})$$

$$\Delta \hat{I}(\mathbf{q}, t, \Delta t) \Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t) = \Delta \hat{I}_1 \Delta \epsilon_1 + \Delta \hat{I}_2 \Delta \epsilon_2, \quad (\text{A2})$$

$$|\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)|^2 = \Delta \hat{\epsilon}_1^2 + \Delta \hat{\epsilon}_2^2 \quad (\text{A3})$$

with

$$\Delta \hat{I}_1 = \frac{1}{N} \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta I(\mathbf{x}, t, \Delta t) \cos\left(\frac{2\pi \mathbf{x}^T \mathbf{q}}{N}\right),$$

$$\Delta \hat{\epsilon}_1 = \frac{1}{N} \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta \epsilon(\mathbf{x}, t, \Delta t) \cos\left(\frac{2\pi \mathbf{x}^T \mathbf{q}}{N}\right),$$

$$\Delta \hat{I}_2 = \frac{1}{N} \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta I(\mathbf{x}, t, \Delta t) \sin\left(\frac{2\pi \mathbf{x}^T \mathbf{q}}{N}\right),$$

$$\Delta \hat{\epsilon}_2 = \frac{1}{N} \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \Delta \epsilon(\mathbf{x}, t, \Delta t) \sin\left(\frac{2\pi \mathbf{x}^T \mathbf{q}}{N}\right).$$

The expected value of $\mathbb{E}[|\hat{I}_o(\mathbf{q}, t, \Delta t)|^2]$ can be verified using properties of the Fourier basis. ■

Proof of Eqs. (8)–(10). The observations of image structure function can be obtained through computing ensemble average of the observed intensity

$$\begin{aligned} D_o(q, \Delta t) &= \langle |\Delta \hat{I}_o(\mathbf{q}, t, \Delta t)|^2 \rangle \\ &= \langle |\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 \rangle + \langle |\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)|^2 \rangle \\ &\quad + 2\langle \Delta \hat{I}(\mathbf{q}, t, \Delta t) \Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t) \rangle \end{aligned}$$

with expected value and variance:

$$\begin{aligned} \mathbb{E}[D_o(q, \Delta t)] &= \mathbb{E}[\langle |\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 \rangle] + \mathbb{E}[\langle |\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)|^2 \rangle] \\ &\quad + 2\mathbb{E}[\langle \Delta \hat{I}(\mathbf{q}, t, \Delta t) \Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t) \rangle] \\ &= \langle |\Delta \hat{I}(\mathbf{q}, t, \Delta t)|^2 \rangle + \frac{1}{n_{\Delta t} n_q} \sum_{t \in \mathcal{S}_{\Delta t}} \sum_{(q_1, q_2) \in \mathcal{S}_q} \mathbb{E}[\Delta \hat{\epsilon}_1^2 + \Delta \hat{\epsilon}_2^2] \\ &\quad + \frac{2}{n_{\Delta t} n_q} \sum_{t \in \mathcal{S}_{\Delta t}} \sum_{(q_1, q_2) \in \mathcal{S}_q} \Delta \hat{I}(\mathbf{q}, t, \Delta t) \mathbb{E}[\Delta \hat{\epsilon}(\mathbf{q}, t, \Delta t)] \\ &= D(q, \Delta t) + \mathbb{V}(\Delta \hat{\epsilon}_1) + \mathbb{V}(\Delta \hat{\epsilon}_2) \\ &= D(q, \Delta t) + 2\sigma_0^2, \end{aligned}$$

where $\langle \cdot \rangle$ denotes averaging over available time points for each Δt , and $(q_1, q_2) \in \mathcal{S}_q$ with $\mathcal{S}_q = \{(q_1, q_2) : q_1^2 + q_2^2 = q^2\}$, $n_q = \#\mathcal{S}_q$, and $n_{\Delta t} = T - \Delta t$:

$$\begin{aligned} \mathbb{V}[D_o(q, \Delta t)] &= \mathbb{V}[\langle |\Delta \hat{I}_o(\mathbf{q}, t, \Delta t)|^2 \rangle] \\ &= \mathbb{V}\left[\frac{1}{n_q n_{\Delta t}} \sum_{(q_1, q_2) \in \mathcal{S}_q} \sum_{t \in \mathcal{S}_{\Delta t}} |\Delta \hat{I}_o(\mathbf{q}, t, \Delta t)|^2\right]. \end{aligned}$$

The variance of the average of intensity over available time points for each Δt and $t_{\max} = T$ is

$$\begin{aligned} & \mathbb{V}\left[\frac{1}{n_{\Delta t}} \sum_{t \in \mathcal{S}_{\Delta t}} |\Delta \hat{I}_o(\mathbf{q}, t, \Delta t)|^2\right] \\ &= \mathbb{V}\left[\frac{1}{n_{\Delta t}} \sum_{t \in \mathcal{S}_{\Delta t}} [\Delta \hat{I}_{o,1}^2(\mathbf{q}, t, \Delta t) + \Delta \hat{I}_{o,2}^2(\mathbf{q}, t, \Delta t)]\right] \\ &= \frac{1}{n_{\Delta t}^2} \sum_{t \in \mathcal{S}_{\Delta t}} \mathbb{V}[\Delta \hat{I}_{o,1}^2(\mathbf{q}, t, \Delta t)] \\ &\quad + \frac{1}{n_{\Delta t}^2} \sum_{t \in \mathcal{S}_{\Delta t}} \mathbb{V}[\Delta \hat{I}_{o,2}^2(\mathbf{q}, t, \Delta t)] \\ &\quad + \frac{1}{n_{\Delta t}^2} \sum_{t_1 \neq t_2} \text{Cov}(\Delta \hat{I}_{o,1}^2(\mathbf{q}, t_1, \Delta t), \Delta \hat{I}_{o,1}^2(\mathbf{q}, t_2, \Delta t)) \\ &\quad + \frac{1}{n_{\Delta t}^2} \sum_{t_1 \neq t_2} \text{Cov}(\Delta \hat{I}_{o,2}^2(\mathbf{q}, t_1, \Delta t), \Delta \hat{I}_{o,2}^2(\mathbf{q}, t_2, \Delta t)), \end{aligned}$$

where the first two terms can be computed as

$$\begin{aligned} & \frac{1}{n_{\Delta t}^2} \sum_{t \in \mathcal{S}_{\Delta t}} (\mathbb{V}[\Delta \hat{I}_{o,1}^2(\mathbf{q}, t, \Delta t)] + \mathbb{V}[\Delta \hat{I}_{o,2}^2(\mathbf{q}, t, \Delta t)]) \\ &= \frac{1}{n_{\Delta t}^2} \sum_{t=1}^{n_{\Delta t}} (\mathbb{V}[(\Delta \hat{I}_1 + \Delta \hat{\epsilon}_1)^2] + \mathbb{V}[(\Delta \hat{I}_2 + \Delta \hat{\epsilon}_2)^2]) \end{aligned}$$

$$\begin{aligned}
 &= \frac{1}{n_{\Delta t}^2} \sum_{t=1}^{n_{\Delta t}} (\mathbb{V}[2\Delta\hat{I}_1\Delta\hat{\epsilon}_1 + \Delta\hat{\epsilon}_1^2] + \mathbb{V}[2\Delta\hat{I}_2\Delta\hat{\epsilon}_1 + \Delta\hat{\epsilon}_2^2]) \\
 &= \frac{1}{n_{\Delta t}^2} \sum_{t=1}^{n_{\Delta t}} (4\sigma_0^2(\Delta\hat{I}_1^2 + \Delta\hat{I}_2^2) + \mathbb{V}(\Delta\hat{\epsilon}_1^2) + \mathbb{V}(\Delta\hat{\epsilon}_2^2)) \\
 &= \frac{1}{n_{\Delta t}^2} \sum_{t=1}^{n_{\Delta t}} (4\sigma_0^2|\Delta\hat{I}(\mathbf{q}, t, \Delta t)|^2 + 4\sigma_0^4),
 \end{aligned}$$

and for $\Delta t = 1, 2, \dots, \lfloor (T-1)/2 \rfloor$, the last two terms follow:

$$\begin{aligned}
 &\sum_{t_1 \neq t_2} \text{Cov}(\Delta\hat{I}_{o,1}^2(\mathbf{q}, t_1, \Delta t), \Delta\hat{I}_{o,1}^2(\mathbf{q}, t_2, \Delta t)) \\
 &= 2 \sum_{t=1}^{T-2\Delta t} \text{Cov}(\Delta\hat{I}_{o,1}^2(\mathbf{q}, t, \Delta t), \Delta\hat{I}_{o,1}^2(\mathbf{q}, t + \Delta t, \Delta t)) \\
 &= 2 \sum_{t=1}^{T-2\Delta t} \text{Cov}([\Delta\hat{I}_1(\mathbf{q}, t, \Delta t) + \hat{\epsilon}_{1,t+\Delta t} - \hat{\epsilon}_{1,t}]^2 \\
 &\quad \times [\Delta\hat{I}_1(\mathbf{q}, t + \Delta t, \Delta t) + \hat{\epsilon}_{1,t+2\Delta t} - \hat{\epsilon}_{1,t+\Delta t}]^2) \\
 &= 2 \sum_{t=1}^{T-2\Delta t} \text{Cov}(\mathbb{E}[[\Delta\hat{I}_1(\mathbf{q}, t, \Delta t) + \hat{\epsilon}_{1,t+\Delta t} - \hat{\epsilon}_{1,t,\Delta t}]^2], \\
 &\quad \times \mathbb{E}[[\Delta\hat{I}_1(\mathbf{q}, t + \Delta t, \Delta t) + \hat{\epsilon}_{1,t+2\Delta t} - \hat{\epsilon}_{1,t+\Delta t}]^2]|\hat{\epsilon}_{1,t+\Delta t}) \\
 &\quad + 2 \sum_{t=1}^{T-2\Delta t} \mathbb{E}[\text{Cov}((\Delta\hat{I}_1(\mathbf{q}, t, \Delta t) + \hat{\epsilon}_{1,t+\Delta t} - \hat{\epsilon}_{1,t})^2, \\
 &\quad [\Delta\hat{I}_1(\mathbf{q}, t + \Delta t, \Delta t) + \hat{\epsilon}_{1,t+2\Delta t} - \hat{\epsilon}_{1,t+\Delta t}]^2|\hat{\epsilon}_{1,t+\Delta t})] \\
 &= 2 \sum_{t=1}^{T-2\Delta t} (\text{Cov}(\hat{\epsilon}_{1,t+\Delta t}^2, \hat{\epsilon}_{1,t+\Delta t}^2) - 4\Delta\hat{I}_1(\mathbf{q}, t, \Delta t) \\
 &\quad \times \Delta\hat{I}_1(\mathbf{q}, t + \Delta t, \Delta t)\text{Cov}(\hat{\epsilon}_{1,t+\Delta t}, \hat{\epsilon}_{1,t+\Delta t})) \\
 &= 2 \sum_{t=1}^{T-2\Delta t} \left(\frac{\sigma_0^4}{2} - 2\sigma_0^2\Delta\hat{I}_1(\mathbf{q}, t, \Delta t)\Delta\hat{I}_1(\mathbf{q}, t + \Delta t, \Delta t) \right),
 \end{aligned}$$

with

$$\begin{aligned}
 \hat{\epsilon}_{1,t} &= \frac{1}{N} \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \epsilon(\mathbf{x}, t) \cos\left(\frac{2\pi\mathbf{x}^T\mathbf{q}}{N}\right), \\
 \hat{\epsilon}_{2,t} &= \frac{1}{N} \sum_{x_1=0}^{N-1} \sum_{x_2=0}^{N-1} \epsilon(\mathbf{x}, t) \sin\left(\frac{2\pi\mathbf{x}^T\mathbf{q}}{N}\right).
 \end{aligned}$$

Similarly, for $\Delta t = 1, 2, \dots, \lfloor (T-1)/2 \rfloor$ and $t_{\max} = T$,

$$\begin{aligned}
 &\sum_{t_1 \neq t_2} \text{Cov}(\Delta\hat{I}_{o,2}^2(\mathbf{q}, t_1, \Delta t), \Delta\hat{I}_{o,2}^2(\mathbf{q}, t_2, \Delta t)) \\
 &= 2 \sum_{t=1}^{T-2\Delta t} \left(\frac{\sigma_0^4}{2} - 2\sigma_0^2\Delta\hat{I}_2(\mathbf{q}, t, \Delta t)\Delta\hat{I}_2(\mathbf{q}, t + \Delta t, \Delta t) \right).
 \end{aligned}$$

For general $\Delta t > \lfloor (T-1)/2 \rfloor$,

$$\begin{aligned}
 &\sum_{t_1 \neq t_2} \text{Cov}(\Delta\hat{I}_{o,1}^2(\mathbf{q}, t_1, \Delta t), \Delta\hat{I}_{o,1}^2(\mathbf{q}, t_2, \Delta t)) \\
 &= \sum_{t_1 \neq t_2} \text{Cov}(\Delta\hat{I}_{o,2}^2(\mathbf{q}, t_1, \Delta t), \Delta\hat{I}_{o,2}^2(\mathbf{q}, t_2, \Delta t)) \\
 &= 0.
 \end{aligned}$$

Combining the variance and covariance expressions developed above, the variance of the average of intensity is

$$\begin{aligned}
 &\mathbb{V}\left[\frac{1}{n_{\Delta t}} \sum_{t \in \mathcal{S}_{\Delta t}} |\Delta\hat{I}_o(\mathbf{q}, t, \Delta t)|^2\right] \\
 &= \frac{2\sigma_0^2}{n_{\Delta t}} \left[2\sigma_0^2 + \frac{2 \sum_{t=1}^{n_{\Delta t}} |\Delta\hat{I}(\mathbf{q}, t, \Delta t)|^2}{n_{\Delta t}} \right. \\
 &\quad \left. + \max(0, T - 2\Delta t) \left(\frac{\sigma_0^2}{n_{\Delta t}} - \frac{2S_{q_1, q_2, \Delta t}}{n_{\Delta t}(T - 2\Delta t)} \right) \right]
 \end{aligned}$$

with

$$\begin{aligned}
 S_{q_1, q_2, \Delta t} &= \sum_{t=1}^{T-2\Delta t} (\Delta\hat{I}_1(\mathbf{q}, t, \Delta t)\Delta\hat{I}_1(\mathbf{q}, t + \Delta t, \Delta t) \\
 &\quad + \Delta\hat{I}_2(\mathbf{q}, t, \Delta t)\Delta\hat{I}_2(\mathbf{q}, t + \Delta t, \Delta t)). \quad (\text{A4})
 \end{aligned}$$

Finally, we have

$$\begin{aligned}
 &\mathbb{V}[D_o(q, \Delta t)] \\
 &= \mathbb{V}\left[\frac{1}{n_q} \sum_{(q_1, q_2) \in \mathcal{S}_q} \frac{1}{n_{\Delta t}} \sum_{t \in \mathcal{S}_{\Delta t}} |\Delta\hat{I}_o(\mathbf{q}, t, \Delta t)|^2\right] \\
 &= \frac{1}{n_q^2} \sum_{(q_1, q_2) \in \mathcal{S}_q} \frac{2\sigma_0^2}{n_{\Delta t}} \left[2\sigma_0^2 + \frac{2 \sum_{t=1}^{n_{\Delta t}} |\Delta\hat{I}(\mathbf{q}, t, \Delta t)|^2}{n_{\Delta t}} \right. \\
 &\quad \left. + \max(0, T - 2\Delta t) \left(\frac{\sigma_0^2}{n_{\Delta t}} - \frac{2S_{q_1, q_2, \Delta t}}{n_{\Delta t}(T - 2\Delta t)} \right) \right] \\
 &= \frac{2\sigma_0^2}{n_q n_{\Delta t}} \left[2\sigma_0^2 + 2D(q, \Delta t) \right. \\
 &\quad \left. + \max(0, T - 2\Delta t) \left(\frac{\sigma_0^2}{n_{\Delta t}} - \frac{2S_{q, \Delta t}}{(T - 2\Delta t)n_{\Delta t}n_q} \right) \right]
 \end{aligned}$$

with

$$S_{q, \Delta t} = \sum_{(q_1, q_2): q_1^2 + q_2^2 = q^2} S_{q_1, q_2, \Delta t}. \quad (\text{A5})$$

APPENDIX B: PARAMETER ESTIMATION IN GAUSSIAN PROCESS REGRESSION

Let $\tilde{\mathbf{D}}_o = (\tilde{D}_o(\theta_1), \dots, \tilde{D}_o(\theta_n))^T$ denote the n observations. The parameters in the Gaussian process contain mean parameter m , variance parameter σ^2 , and inverse range parameters $\boldsymbol{\beta} = (\beta_1, \beta_2)$ in the kernel function. Conditional on $\boldsymbol{\beta}$ and the regularization parameter λ , the

maximum likelihood estimator of the mean parameter is $m_{\text{est}} = (\mathbf{1}_n^T \tilde{\mathbf{R}}^{-1} \mathbf{1}_n^T)^{-1} \mathbf{1}_n^T \tilde{\mathbf{R}}^{-1} \tilde{\mathbf{D}}_o$, with $\tilde{\mathbf{R}} = \mathbf{R} + n\lambda \mathbf{I}_n$ and $\sigma_{\text{est}}^2 = S^2/n$ with $S^2 = (\tilde{\mathbf{D}}_o - \mathbf{1}_n \hat{\mathbf{m}})^T \tilde{\mathbf{R}}^{-1} (\tilde{\mathbf{D}}_o - \mathbf{1}_n \hat{\mathbf{m}})$. Plugging $(m_{\text{est}}, \sigma_{\text{est}}^2)$, the profile likelihood of parameters $(\boldsymbol{\beta}, \lambda)$ in the covariance function follows $\mathcal{L}(\boldsymbol{\beta}, \lambda) \propto |\mathbf{K}|^{-\frac{1}{2}} (S^2)^{-\frac{n}{2}}$. Since no closed form expression of the maximum likelihood estimator for the range and regularization parameters is available, one often numerically maximizes the profile likelihood to obtain the estimates of $(\boldsymbol{\beta}, \lambda)$. When the sample size is small, the MLE can be unstable and marginal posterior mode estimation with robust parametrization is often used [56]. We implemented the parameter estimation and predictions of GPR by the ‘‘ROBUSTGASP’’ package available in R and MATLAB [47].

APPENDIX C: DETECTION OF ESTIMATOR BIAS

The overestimation by a series of previously used methods can be detected by plotting $D_o(q, \Delta t)$ over all q values at one Δt (see Fig. 6). If at high q , $D_o(q, \Delta t)$'s rate of change slows, then this implies that $A(q)[1 - f(q, \Delta t)]$ is close to zero, and the bias in estimating the noise term by the second to the fourth estimator in Table I is negligible. However, if $D_o(q, \Delta t)$ decreases (even slightly) as the value q increases, then the bias of the estimator is non-negligible.

APPENDIX D: DERIVATION OF MEAN SQUARED DISPLACEMENT

Here we derive the MSD for each of the different scenarios explored in simulation. First, the simulated particles in Secs. III B and III C all undergo Brownian motion. Without loss of generality, we may assume the variance of $\Delta x_{i,j}(t)$ is σ_s^2 . For any $\mathbf{x}(t)$, $\mathbf{x}(t + \Delta t) \in \mathbb{R}^2$, the MSD can be simply

computed by

$$\begin{aligned} & \mathbb{E}[[x_{ij}(t + \Delta t) - x_{ij}(t)]^2] \\ &= \mathbb{V}[x_{ij}(t + \Delta t) - x_{ij}(t)] + \{\mathbb{E}[x_{ij}(t + \Delta t) - x(t)]\}^2 \\ &= \mathbb{V}\left[\sum_{k=0}^{\Delta t-1} \Delta x_{ij}(t+k)\right] + 0 = \sigma_s^2 \Delta t, \end{aligned}$$

for any $j = 1, \dots, p$ and $i = 1, 2$. Since particles move isotropically in a 2D space, the MSD is $2\sigma_s^2 \Delta t$.

In simulated scenarios presented in Sec. III D, similarly we can split the MSD into two terms. Noting $\mathbb{E}[x_{ij}(t + \Delta t) - x_{ij}(t)] = \mu_D \Delta t$ and the process is isotropic, the MSD is $2\sigma_s^2 \Delta t + 2\mu_D^2 \Delta t^2$,

In simulated scenarios presented in Sec. III E, note that when $t = t_1$, $x_{ij}(t_1) \sim N(x_{ij}(t_0), \sigma_D^2)$. For any $t > t_1$, it is not hard to show

$$\begin{aligned} \mathbb{E}[x_{1j}(t)] &= (t - 1)\mu_D \cos(\theta_j) + x_{1j}(t_0), \\ \mathbb{E}[x_{2j}(t)] &= (t - 1)\mu_D \sin(\theta_j) + x_{2j}(t_0), \end{aligned}$$

and consequently

$$\mathbb{E}\left[\sum_{i=1}^2 (x_{ij}(t + \Delta t) - x_{ij}(t))^2\right] = \mu_D^2 \Delta t^2. \quad (\text{D1})$$

It is easy to verify $\mathbb{V}[x_{ij}(t)] = \sigma_s^2$. Thus, we have

$$\begin{aligned} & \mathbb{V}[x_{ij}(t + \Delta t) - x_{ij}(t)] \\ &= \mathbb{V}[x_{ij}(t + \Delta t)] + \mathbb{V}[x_{ij}(t)] - 2\text{Cov}(x_{ij}(t + \Delta t), x_{ij}(t)) \\ &= 2\sigma_s^2 - 2\text{Cov}(\rho^{\Delta t} x_{ij}(t), x_{ij}(t)) \\ &= 2\sigma_s^2 - 2\sigma_s^2 \rho^{\Delta t}. \end{aligned} \quad (\text{D2})$$

Since the process is on a two-dimensional space, combining (D1) and (D2), the MSD is $4\sigma_s^2 - 4\sigma_s^2 \rho^{\Delta t} + \mu_D^2 \Delta t^2$.

-
- [1] J. C. Crocker and D. G. Grier, Methods of digital video microscopy for colloidal studies, *J. Colloid Interface Sci.* **179**, 298 (1996).
- [2] T. Savin and P. S. Doyle, Statistical and sampling issues when using multiple particle tracking, *Phys. Rev. E* **76**, 021501 (2007).
- [3] F. Giavazzi, D. Brogioli, V. Trappe, T. Bellini, and R. Cerbino, Scattering information obtained by optical microscopy: Differential dynamic microscopy and beyond, *Phys. Rev. E* **80**, 031403 (2009).
- [4] F. Giavazzi and R. Cerbino, Digital Fourier microscopy for soft matter dynamics, *J. Opt.* **16**, 083001 (2014).
- [5] R. Cerbino and V. Trappe, Differential Dynamic Microscopy: Probing Wave Vector Dependent Dynamics with a Microscope, *Phys. Rev. Lett.* **100**, 188102 (2008).
- [6] A. V. Bayles, T. M. Squires, and M. E. Helgeson, Probe microrheology without particle tracking by differential dynamic microscopy, *Rheol. Acta* **56**, 863 (2017).
- [7] P. J. Lu, F. Giavazzi, T. E. Angelini, E. Zaccarelli, F. Jargstorff, A. B. Schofield, J. N. Wilking, M. B. Romanowsky, D. A. Weitz, and R. Cerbino, Characterizing Concentrated, Multiplex Scattering, and Actively Driven Fluorescent Systems with Confocal Differential Dynamic Microscopy, *Phys. Rev. Lett.* **108**, 218103 (2012).
- [8] V. A. Martinez, R. Besseling, O. A. Croze, J. Tailleur, M. Reufer, J. Schwarz-Linek, L. G. Wilson, M. A. Bees, and W. C. Poon, Differential dynamic microscopy: A high-throughput method for characterizing the motility of microorganisms, *Biophys. J.* **103**, 1637 (2012).
- [9] F. Giavazzi, A. Fornasieri, A. Vailati, and R. Cerbino, Equilibrium and non-equilibrium concentration fluctuations in a critical binary mixture, *Eur. Phys. J. E* **39**, 1 (2016).
- [10] Y. Gao, J. Kim, and M. E. Helgeson, Microdynamics and arrest of coarsening during spinodal decomposition in thermoreversible colloidal gels, *Soft Matter* **11**, 6360 (2015).
- [11] F. Giavazzi, S. Crotti, A. Speciale, F. Serra, G. Zanchetta, V. Trappe, M. Buscaglia, T. Bellini, and R. Cerbino, Viscoelasticity of nematic liquid crystals at a glance, *Soft Matter* **10**, 3938 (2014).
- [12] R. Cerbino and P. Cicuta, Perspective: Differential dynamic microscopy extracts multi-scale activity in complex fluids and biological systems, *J. Chem. Phys.* **147**, 110901 (2017).

- [13] T. Savin and P. S. Doyle, Static and dynamic errors in particle tracking microrheology, *Biophys. J.* **88**, 623 (2005).
- [14] M. Reufer, V. A. Martinez, P. Schurtenberger, and W. C. Poon, Differential dynamic microscopy for anisotropic colloidal dynamics, *Langmuir* **28**, 4618 (2012).
- [15] C. Kurzthaler, C. Devailly, J. Arlt, T. Franosch, W. C. K. Poon, V. A. Martinez, and A. T. Brown, Probing the Spatiotemporal Dynamics of Catalytic Janus Particles with Single-Particle Tracking and Differential Dynamic Microscopy, *Phys. Rev. Lett.* **121**, 078001 (2018).
- [16] M. Escobedo-Sánchez, J. Segovia-Gutiérrez, A. Zuccolotto-Bernez, J. Hansen, C. Marciniak, K. Sachowsky, F. Platten, and S. Egelhaaf, Microliter viscometry using a bright-field microscope: η -ddm, *Soft Matter* **14**, 7016 (2018).
- [17] R. Cerbino, D. Piotti, M. Buscaglia, and F. Giavazzi, Dark field differential dynamic microscopy enables accurate characterization of the roto-translational dynamics of bacteria and colloidal clusters, *J. Phys.: Condens. Matter* **30**, 025901 (2017).
- [18] F. Giavazzi, C. Malinverno, G. Scita, and R. Cerbino, Tracking-free determination of single-cell displacements and division rates in confluent monolayers, *Front. Phys.* **6**, 120 (2018).
- [19] H. Moon, A. M. Dean, and T. J. Santner, Two-stage sensitivity-based group screening in computer experiments, *Technometrics* **54**, 376 (2012).
- [20] G. Lee, G. Leech, M. J. Rust, M. Das, R. J. McGorty, J. L. Ross, and R. M. Robertson-Anderson, Myosin-driven actin-microtubule networks exhibit self-organized contractile dynamics, *Sci. Adv.* **7**, eabe4334 (2021).
- [21] F. Giavazzi, V. Trappe, and R. Cerbino, Multiple dynamic regimes in a coarsening foam, *J. Phys.: Condens. Matter* **33**, 024002 (2020).
- [22] M. Norouziadeh, G. Cerchiari, and F. Croccolo, Increased performance in DDM analysis by calculating structure functions through Fourier transform in time, *arXiv:2012.05695*.
- [23] P. Edera, D. Bergamini, V. Trappe, F. Giavazzi, and R. Cerbino, Differential dynamic microscopy microrheology of soft materials: A tracking-free determination of the frequency-dependent loss and storage moduli, *Phys. Rev. Mater.* **1**, 073804 (2017).
- [24] T. G. Mason, K. Ganesan, J. H. van Zanten, D. Wirtz, and S. C. Kuo, Particle Tracking Microrheology of Complex Fluids, *Phys. Rev. Lett.* **79**, 3282 (1997).
- [25] T. G. Mason and D. A. Weitz, Optical Measurements of Frequency-Dependent Linear Viscoelastic Moduli of Complex Fluids, *Phys. Rev. Lett.* **74**, 1250 (1995).
- [26] M. S. Safari, M. A. Vorontsova, R. Poling-Skutvik, P. G. Vekilov, and J. C. Conrad, Differential dynamic microscopy of weakly scattering and polydisperse protein-rich clusters, *Phys. Rev. E* **92**, 042712 (2015).
- [27] T. Sentjabskaja, E. Zaccarelli, C. De Michele, F. Sciortino, P. Tartaglia, T. Voigtmann, S. U. Egelhaaf, and M. Laurati, Anomalous dynamics of intruders in a crowded environment of mobile obstacles, *Nat. Commun.* **7**, 1 (2016).
- [28] C. E. Rasmussen, *Gaussian Processes for Machine Learning* (MIT Press, Cambridge, MA, 2006).
- [29] The MATLAB package is available at: <https://github.com/UncertaintyQuantification/DDM-UQ>.
- [30] F. Giavazzi, P. Edera, P. J. Lu, and R. Cerbino, Image windowing mitigates edge effects in differential dynamic microscopy, *Eur. Phys. J. E* **40**, 97 (2017).
- [31] A. V. Bayles, T. M. Squires, and M. E. Helgeson, Dark-field differential dynamic microscopy, *Soft Matter* **12**, 2440 (2016).
- [32] B. Nijboer and A. Rahman, Time expansion of correlation functions and the theory of slow neutron scattering, *Physica (Amsterdam)* **32**, 415 (1966).
- [33] E. R. Weeks, J. C. Crocker, A. C. Levitt, A. Schofield, and D. A. Weitz, Three-dimensional direct imaging of structural relaxation near the colloidal glass transition, *Science* **287**, 627 (2000).
- [34] L. G. Wilson, V. A. Martinez, J. Schwarz-Linek, J. Tailleur, G. Bryant P. N. Pusey, and W. C. K. Poon, Differential Dynamic Microscopy of Bacterial Motility, *Phys. Rev. Lett.* **106**, 018101 (2011).
- [35] L. Jawerth, E. Fischer-Friedrich, S. Saha, J. Wang, T. Franzmann, X. Zhang, J. Sachweh, M. Ruer, M. Ijavi, S. Saha *et al.*, Protein condensates as aging Maxwell fluids, *Science* **370**, 1317 (2020).
- [36] E. M. Furst and T. M. Squires, *Microrheology* (Oxford University Press, Oxford, 2017).
- [37] J. A. McGlynn, N. Wu, and K. M. Schultz, Multiple particle tracking microrheological characterization: Fundamentals, emerging techniques and applications, *J. Appl. Phys.* **127**, 201101 (2020).
- [38] M. Rupp, A. Tkatchenko, K.-R. Müller, and O. A. von Lilienfeld, Fast and Accurate Modeling of Molecular Atomization Energies with Machine Learning, *Phys. Rev. Lett.* **108**, 058301 (2012).
- [39] A. P. Bartók, R. Kondor, and G. Csányi, On representing chemical environments, *Phys. Rev. B* **87**, 184115 (2013).
- [40] S. Chmiela, A. Tkatchenko, H. E. Sauceda, I. Poltavsky, K. T. Schütt, and K.-R. Müller, Machine learning of accurate energy-conserving molecular force fields, *Sci. Adv.* **3**, e1603015 (2017).
- [41] F. Brockherde, L. Vogt, L. Li, M. E. Tuckerman, K. Burke, and K.-R. Müller, Bypassing the kohn-sham equations with machine learning, *Nat. Commun.* **8**, 1 (2017).
- [42] S. Chmiela, H. E. Sauceda, K.-R. Müller, and A. Tkatchenko, Towards exact molecular dynamics simulations with machine-learned force fields, *Nat. Commun.* **9**, 1 (2018).
- [43] A. P. Bartók, J. Kermode, N. Bernstein, and G. Csányi, Machine Learning a General-Purpose Interatomic Potential for Silicon, *Phys. Rev. X* **8**, 041048 (2018).
- [44] D. M. Wilkins, A. Grisafi, Y. Yang, K. U. Lao, R. A. DiStasio, and M. Ceriotti, Accurate molecular polarizabilities with coupled cluster theory and machine learning, *Proc. Natl. Acad. Sci. U.S.A.* **116**, 3401 (2019).
- [45] K. R. Anderson, I. A. Johanson, M. R. Patrick, M. Gu, P. Segall, M. P. Poland, E. K. Montgomery-Brown, and A. Miklius, Magma reservoir failure and the onset of caldera collapse at Kilauea volcano in 2018, *Science* **366**, eaaz1822 (2019).
- [46] J. Wu and M. Gu, Emulating the first principles of matter: A probabilistic roadmap, *arXiv:2010.05942*.
- [47] M. Gu, J. Palomo, and J. O. Berger, RobustGaSP: Robust Gaussian stochastic process emulation in R, *R Journal* **11**, 112 (2019).

- [48] Y. Gao and M. L. Kilfoil, Accurate detection and complete tracking of large populations of features in three dimensions, *Opt. Express* **17**, 4685 (2009).
- [49] J.-Z. Xue, D. J. Pine, S. T. Milner, X.-L. Wu, and P. M. Chaikin, Nonergodicity and light scattering from polymer gels, *Phys. Rev. A* **46**, 6550 (1992).
- [50] N. Monnier, S.-M. Guo, M. Mori, J. He, P. Lénárt, and M. Bathe, Bayesian approach to MSD-based analysis of particle motion in live cells, *Biophys. J.* **103**, 616 (2012).
- [51] J. F. Swindells, *Viscosities of Sucrose Solutions at Various Temperatures: Tables of Recalculated Values*, Vol. 440 (US Government Printing Office, Washington DC, 1958).
- [52] H. Rehage and H. Hoffmann, Viscoelastic surfactant solutions: Model systems for rheological research, *Mol. Phys.* **74**, 933 (1991).
- [53] T. G. Mason, Estimating the viscoelastic moduli of complex fluids using the generalized Stokes-Einstein equation, *Rheol. Acta* **39**, 371 (2000).
- [54] J. Fricks, L. Yao, T. C. Elston, and M. G. Forest, Time-domain methods for diffusive transport in soft matter, *SIAM J. Appl. Math.* **69**, 1277 (2009).
- [55] M. Lysy, N. S. Pillai, D. B. Hill, M. G. Forest, J. W. Mellnik, P. A. Vasquez, and S. A. McKinley, Model comparison and assessment for single particle tracking in biological fluids, *J. Am. Stat. Assoc.* **111**, 1413 (2016).
- [56] M. Gu, X. Wang, and J. O. Berger, Robust Gaussian stochastic process emulation, *Ann. Stat.* **46**, 3038 (2018).
- Correction:* Minor errors in some displayed and inline equations in Appendix A have been fixed.