Nutation-based longitudinal-sensing protocols for high-field NMR with nitrogen-vacancy centers in diamond

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Nitrogen-vacancy (N-V) centers in diamond enable nuclear magnetic resonance (NMR) spectroscopy of samples at the nano- and microscale. However, at typical tesla-scale NMR magnetic field strengths, N-V–NMR protocols become difficult to implement due to the challenge of driving fast N-V pulse sequences sensitive to nuclear Larmor frequencies above a few megahertz. We perform simulations and theoretical analysis of the experimental viability of N-V NMR at tesla-scale magnetic fields using a measurement protocol called DRACAERIS (Double Rewound ACquisition Amplitude Encoded Radio Induced Signal). DRACAERIS detects the longitudinal magnetization of the NMR sample at a much lower driven Rabi frequency, more suitable technically for N-V detection. We discuss how pulse errors, finite pulse lengths, and nuclear spin-spin couplings affect the resulting NMR spectra. We find that DRACAERIS is less susceptible to pulse imperfections and off-resonance effects than previous protocols for longitudinal magnetization detection. We also identify reasonable parameters for experimental implementation.

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I. INTRODUCTION

Nitrogen-vacancy (N-V) centers in diamond are widely used for micro- and nanoscale magnetic sensing applications due to their long electronic spin coherence times, optical spin-state preparation and readout at room temperature, and straightforward technical implementation [1-3]. With dynamical decoupling sequences, N-V centers can be made sensitive to oscillating (ac) magnetic fields from nearby spins, e.g., nuclear magnetic resonance (NMR) signals of small numbers of nuclei near the diamond surface [4-8]. Using coherent sensing protocols, N-VNMR can achieve sensitivity better than 30 pT/ $\sqrt{\text{Hz}}$ (for N-V ensembles) and a spectral resolution of approximately 1 Hz [9–11], enabling high-resolution NMR spectroscopy of microscale sample volumes [9,12,13]. Camera-based detection of ensemble N-V fluorescence can also enable wide-field NMR spectral imaging with high spatial

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resolution [6], providing a tool for chemical analysis in the biological and physical sciences. A simplified schematic of an ensemble N-V–NMR apparatus is shown in Fig. 1(a).

Distinguishing chemical species via conventional NMR spectroscopy requires chemical-shift resolution that is only achievable at high applied (bias) magnetic fields (typically approximately 1 T or larger). To detect a sample NMR signal (produced by the transverse nuclear magnetization of the sample oscillating at its Larmor frequency), a dynamical decoupling sequence is applied to the N-Vcenters with a pulse spacing on the time scale of half the nuclear Larmor precession period. At tesla-scale biasfield strengths, the NMR precession period is on the order of tens of nanoseconds, requiring that strong microwave (MW) pulses be applied to the N-V electronic spins at high carrier frequencies [10]. For example, for a 1-T bias field, N-V-NMR detection of a proton NMR signal requires that MW pulses be applied to the N-V centers at a carrier frequency of approximately 25 GHz, with a repetition rate of approximately 85 MHz (given by the proton Larmor frequency) and a Rabi frequency of approximately 500 MHz (to approximate instantaneous pulses in the N-V sensing protocol). Applying such large-amplitude high-frequency MW pulses while also maintaining sufficient field homogeneity over the diamond surface is technically daunting and has yet to be realized for an ensemble N-V-NMR system.

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FIG. 1. (a) The schematic of an ensemble N-V-NMR apparatus. A green laser optically excites an N-V layer near the diamond surface, is totally reflected, and a camera or photodetector collects the emitted N-V red fluorescence. A fluid sample is in near contact with the N-V layer, e.g., within a microfluidic apparatus. An integrated coplanar waveguide applies microwaves to the N-V ensemble. A nearby radio-frequency (rf) coil (not shown) is used to manipulate nuclear spins in the sample. A bias magnetic field (not shown) provides Zeeman shifts to both the N-V electronic spins and the sample nuclear spins. (b) The AERIS protocol from Ref. [14]. Nuclear spins are initialized with a $\pi/2$ rf pulse and their precession is then measured via a series of N acquisition steps. During each acquisition, the N-V centers are first initialized with an optical pulse. A $2\pi N$ rf nutation pulse with duration μ is applied to the nuclei. During this rf nutation pulse, a dynamical decoupling sequence such as XY8 is applied to the N-V electronic spins via microwave (MW) pulses, followed by readout of the N-V fluorescence. The time outside the rf pulse adds up to a delay τ , during which the nuclear spins freely precess. (c) The DRACAERIS protocol. Each acquisition now consists of two steps, a $2\pi N$ rf pulse on the nuclei driving forward nutation, followed by a second $2\pi N$ rf pulse with the opposite phase to rewind (i.e., reverse) the nutation. N-V dynamical decoupling measurements are performed during both nutation pulses. Subtracting the second N-V fluorescence signal from the first helps minimize common-mode laser noise. The duration μ is defined as the total time for both $2\pi N$ rf nutation pulses. The box to the lower left gives the color-coded definitions of the different types of MW pulses applied to the N-V centers for both the AERIS and DRACAERIS protocols. Note that for both AERIS and DRACAERIS, typical operational conditions will have $\tau > \mu$, as discussed in the text. (d) (e) Bloch-sphere diagrams highlighting errors caused by off-resonant driving of the nuclear spin magnetization. In (d), the green trajectory shows the nuclear magnetization vectors before, during, and after the AERIS nutation pulse. When the pulse is completed, the longitudinal magnetization vector M_z does not return to the x-y plane of the Bloch sphere. In (e), the red trajectory shows that the DRACAERIS protocol corrects to leading order for these off-resonance driving effects and resets M_z to zero after the two rf nutation pulses.

To address this challenge, an alternative measurement protocol called AERIS (Amplitude-Encoded Radio Intensity Signal) [14] has been proposed. Rather than detecting transverse NMR magnetization oscillating at the Larmor frequency, AERIS detects the longitudinal nuclear magnetization of the sample oscillating at a much lower drive frequency, i.e., the nuclear Rabi frequency, which can be tuned to a technically optimal frequency for N-V detection protocols (e.g., dynamic decoupling), typically hundreds of kilohertz to a few megahertz. The AERIS protocol, shown in Fig. 1(b), begins with an initial $\pi/2$ pulse, followed by a series of $2\pi N$ nutation pulses separated by regular intervals τ , all applied to the sample nuclear spins. During these nutation pulses, the nuclear spins are driven at the chosen Rabi frequency while dynamical decoupling magnetometry is simultaneously performed via the N-V centers, yielding a series of NMR amplitude measurements over time. For each time point, the amplitude of the longitudinal magnetization signal depends on the relative phase between the nuclear spins and the phase of the nutation pulse, a phase shift that builds up during the interpulse intervals due to the frequency difference between the nuclear spin dynamics and the reference oscillator. The resulting longitudinal magnetization signal is expected to contain information about the nuclear spin sample analogous to that of a conventional transverse magnetization NMR signal detected inductively with a heterodyne circuit. Likewise, the Fourier transform of the resulting time course using the AERIS protocol is expected to be comparable to a conventional NMR spectrum, albeit with the ability provided by N-V centers to detect very small samples.

As outlined above, AERIS provides an elegant solution to some challenges of high-field N-VNMR. However, nonideal experimental conditions and sample properties may degrade its performance. For example, the finite bandwidth of pulses and the possibility of imperfect pulse rotations can lead to undesired dynamics of the nuclear spins. Finite lengths of the induced rotations also affect spin-spin dynamics within real sample molecules where J couplings are present, thereby modifying the NMR spectra. In this work, we present an adapted protocol (DRA-CAERIS) designed to alleviate these effects and discuss how to interpret the resulting spectral data in relation to conventional NMR spectroscopy.

II. METHODS

To investigate how nuclear spins evolve under the AERIS and DRACAERIS protocols, a numerical simulation has been created to track the density operator of the sample spins under the effects of different Hamiltonians during both the free evolution τ and the nutation period μ . In the rotating frame, these Hamiltonians can be defined as

$$\hat{H}_0 = \sum_i \omega_i \hat{I}_{z,i} + \sum_{j>k} 2\pi J_{jk} \hat{\mathbf{I}}_j \cdot \hat{\mathbf{I}}_k$$
(1)

during free evolution and

$$\hat{H}_{\text{drive}} = \hat{H}_0 + \sum_i \Omega \hat{I}_{x,i}$$
(2)

during nutation pulses, where ω_i represents the *i*th chemical-shift precession frequency in the rotating frame, I and its components are the nuclear spin operators, J_{ik} represents the spin-spin coupling constant between the *j* th and kth spins of the sample molecule, and Ω is the Rabi frequency of the nutation pulse. Since neither Hamiltonian is time dependent, one can use the simple time-evolution operator $U(\hat{H}, t) = e^{i\hat{H}t}$ to evolve the density matrix of the sample spins under both free evolution τ and driving μ periods. Due to chemical shifts, the resonance frequencies of different nuclei are not equal. This results in small deviations of the effective Rabi frequencies for each class of sample spin. This is reflected in this formulation of the Hamiltonian, since the driving field ΩI_x is static in the rotating frame and therefore is not resonant for any of the $\omega_i I_{z,i}$ terms unless $\omega_i = 0$.

At the beginning of each simulation, all nuclear spins begin along the \hat{y} axis, representing the state following a $\pi/2$ excitation pulse along -x. This state is represented by a density matrix ρ and evolution under an arbitrary

Hamiltonian \hat{H} is calculated using the time-evolved state $\rho(t) = U(\hat{H}, t)^{-1} \rho U(\hat{H}, t)$. Excitation is followed by a series of repeated N-V-NMR signal-acquisition blocks. In each block, the free-evolution Hamiltonian \hat{H}_0 is applied to the nuclei for duration $\tau/2$, the state is subjected to the nuclear driving Hamiltonian \hat{H}_{drive} for nutation duration μ , and finally the free-evolution Hamiltonian \hat{H}_0 is applied to the nuclei for duration $\tau/2$ again. In an actual experiment, one or more ac magnetometry sequences are performed on the N-V centers during nuclear driving and the resulting fluorescence produced by the N-V centers is directly related to the amplitude and phase of the longitudinal nuclear magnetization (M_z^{N-V}) . For each acquisition block, we can therefore determine what the N-V response would be by calculating $\langle M_z^{N-V} \rangle = tr(\rho(t_{max})\hat{\mathbf{I}}_z)$ at the first maximum of the nutation oscillation t_{max} (i.e., a quarter of a period in a 2π rf pulse used for nuclear spin nutation). The signal-acquisition blocks are repeated on the order of hundreds to thousands of times, matching the duration of the nuclear T_2 . The Fourier transform of these acquisitions produces the longitudinally detected N-V-NMR spectrum.

The effects of nuclear spin decoherence are implemented by scaling the magnetization magnitude by a factor

$$\exp\left(-\frac{\tau+\mu}{T_2}\right) \tag{3}$$

after each acquisition step. In the case of conventional NMR, $\mu = 0$. Here, we make the assumption that the nuclear spin decoherence times during the nutation pulse and free precession are similar, which is typically valid for liquid samples in a homogeneous field [15,16].

To perform the pulse-error analysis shown in Fig. 5, we track the geometric position of the N-V spin magnetization vector \overline{M} as it accumulates phase in the presence of bias field B_0 and longitudinal nuclear spin signals produced by Rewind2 and DRACAERIS nutation pulses. Similar to our simulation of the nuclear spins, the magnetization vector of the N-V ensemble is initialized in the transverse plane along \hat{x} to reflect an initial MW $\pi/2$ pulse. The ensemble N-V magnetization orientation is represented by the 2×2 density matrix of a two-level system. MW π pulses on the N-V centers are applied as simple geometric rotation operators around the \hat{x} and \hat{y} axes. The degree of rotation induced by a π pulse is varied from $\theta \in [90, 270]$ degrees, corresponding to a set of pulse errors between -90 and +90 degrees. At the end of the sequence, a final MW $\pi/2$ pulse projects the ensemble N-V magnetization along \hat{z} for measurement as the \hat{z} projection $\langle M_z^{N-V} \rangle$, which is then plotted for each value of θ .

To account for N-V dephasing effects dictated by the characteristic time T_2^* , we have repeated the above protocol for 100 different values of bias-field intensity B_0 selected from a Gaussian distribution. The width of the Gaussian distribution $\sigma = (1/(\sqrt{2\pi}T_2^*))$ increases with shorter

 T_2^* [17] and $\langle M_z^{N-V} \rangle$ measurements for each value of B_0 are averaged together.

III. DRACAERIS PROTOCOL

The AERIS protocol [14] involves down-sampling nuclear magnetization signals via longitudinal detection of the sample spins during repeated $2\pi N$ rf nutation pulses. However, this proposal includes simplifications that are generally not applicable to real NMR samples, most importantly neglecting chemical-shift differences and J couplings during the nuclear drive pulses. Also, in practical applications, pulses are not perfect throughout the sample because of imperfect calibration and field inhomogeneities. Since the AERIS protocol relies on repeated application of these pulses, even small pulse errors can have a large cumulative effect. For example, these errors break the underlying assumption that during the free-precession period τ , the nuclear spins are in the *x*-*y* plane, which can lead to incomprehensible NMR spectra.

In order to address pulse errors, the DRACERIS protocol includes a key modification: every $2\pi N$ "forward" rf nutation pulse is immediately followed by an equivalent "rewinding" (or reverse) pulse of opposite phase, illustrated in Fig. 1(c). The rewinding pulse compensates to leading order for the effects of pulse errors and returns the spins to the x-y plane. We call this protocol DRACAERIS (Double Rewound ACquisition Amplitude Encoded Radio Induced Signal). To illustrate the effect of pulse errors schematically, in Fig. 1(d) we show the magnetization of an off-resonant nuclear spin during one cycle of the AERIS sequence. Following the nutation pulse, magnetization is shifted out of the transverse plane. In contrast, using DRACAERIS [Fig. 1(e)], magnetization is shifted out of the transverse plane following the first (forward) nutation pulse but is returned there by the second (rewinding) pulse.

Pulse errors can cause significant spectral shifts that make NMR spectral identification difficult. In Fig. 2(a), we show simulated measurements of three uncoupled spins with differing chemical shifts using conventional NMR detection, as well as N-V NMR with AERIS and DRA-CAERIS in a 1-T field and with a 2% error in the nutation rate used for AERIS and DRACAERIS. The extra phase build-up due to pulse errors causes AERIS measurements to shift toward higher frequencies, with the largest effects occurring for the lowest-frequency NMR spectral lines. For example, the spin at a 25-Hz chemical shift ($\delta = 0.59$ ppm) would be measured at 103 Hz ($\delta = 2.42$ ppm) using AERIS, corresponding to a spectral line error $\Delta = 1.83$ ppm. In comparison, DRACAERIS eliminates this effect and produces the correct NMR spectrum.

For weak nutation pulses, AERIS also produces small spectral shifts in off-resonance nuclear spins, an effect caused by limited nutation-pulse bandwidth. In Fig. 2(b), we show simulated measurements of the same spins as in

Fig. 2(a) but for perfect pulses and a weak nutation pulse with a Rabi frequency of just 5 kHz. In this example of an extreme case of weak pulses and thus small bandwidth,



FIG. 2. Longitudinal magnetization detection by an N-V sensor produces spectral line shifts, in comparison to conventional NMR detection of transverse magnetization, in the presence of pulse errors and off-resonant spins. Here, the conventional, AERIS, and DRACAERIS detection of an NMR sample is simulated. The sample contains a system with three types of uncoupled spins in a 1-T bias field with chemical shifts of 25, 250, and 500 Hz [$\delta = 0.59, 5.9, 11.7$ ppm (parts per million)], and a coherence time $T_2 = 1$ s for each spin type. (a) With a 2% pulse error, AERIS produces a significant shift Δ in some spectral lines, while DRACAERIS does not. Here, $\tau = 800 \ \mu s$, $\mu = 20 \ \mu s$ for AERIS and $\mu = 40 \ \mu s$ for DRACAERIS, with Rabi frequency $\Omega = 200$ kHz. (b) Even with perfect pulses, small errors can arise due to off-resonance effects if the Rabi frequency Ω is too low. Here, $\tau = 800 \ \mu s$, $\mu = 800 \ \mu s$ for AERIS and $\mu = 1600 \ \mu s$ for DRACAERIS, with $\Omega = 5$ kHz. The different line widths reflect the significantly longer total measurement time when μ is large (see also Fig. 4).

the AERIS nutation time μ equals the free-precession time τ and the nutation Rabi frequency is only about 10 times larger than the chemical shift of the farthest NMR spectral line. As a result, off-resonant spectral lines are shifted slightly and the effect is larger for spins that are further off resonance. The maximum shift using these parameters is about 0.5%.

DRACAERIS does not correct for these effects and, in fact, produces about twice as large a shift due to the extra nutation pulse. Fortunately, off-resonance effects are much smaller for typical experimental parameters. To achieve high sensitivity with either protocol, the nuclear spin driving must be strong enough to produce multiple Rabi oscillations within the N-V dephasing time T_2 ; also, a shorter MW-pulse spacing during the dynamical decoupling sequence improves N-V T_2 . Given a typical N-V ensemble T_2 of 20 µs for an XY8 sequence [18], a minimum nuclear spin rf nutation-pulse Rabi frequency $\Omega = 200 \text{ kHz}$ is necessary to create four periods of nuclear oscillations. Adding in rewinding for DRACAERIS gives a total nutation-pulse time μ of at least 40 μ s. An additional constraint arises from the required sampling bandwidth. For example, to cover roughly 15 ppm of a proton NMR spectrum at 1 T, one would need a sampling frequency of at least $f_{sr} = 1.25$ kHz to achieve the Nyquist limit. Thus $\tau = 1/f_{sr}$ must be shorter than approximately 800 μ s. A typical ratio τ/μ , i.e., the amount of free precession versus acquisition time, is therefore <20. In this scenario, the nutation Rabi frequency is about 300 times stronger than the largest chemical shifts and the offresonance effects for AERIS or DRACAERIS are much smaller than the NMR line width for a typical sample.

Because data acquisition is only possible during μ , sensitivity is expected to scale by a factor of $\sqrt{\mu/(\mu + \tau)}$ compared with N-V–NMR methods using near-continuous acquisition, such as CASR [9]. However, utilizing DRA-CAERIS together with a quantum logic enhanced readout scheme, as described in Ref. [18], would allow N-V readout to continue during the nuclear spin free-evolution period τ , offering sensitivity closer to continuous acquisition without changing the nutation duration μ .

IV. MAGNETIC FIELD RESCALING

A third cause of measurement error for N-V–NMR longitudinal-sensing protocols is nuclear spin-spin coupling in the target sample. During conventional inductive NMR detection, the signal resulting from transverse magnetization $\langle M_y \rangle$ is sampled at a repeated time interval τ . For conventional inductive detection, τ tracks the free-precession time of the nuclear spins. However, for AERIS and DRACAERIS, the signal is measured at intervals of $\tau + \mu$. If evolution of the sample nuclear spin system were suspended during the acquisition time μ , then the NMR spectra created by the three techniques would be identical



FIG. 3. Longitudinal magnetization detection by an N-V sensor using DRACAERIS produces spectral distortions for coupled spins, which can be corrected using magnetic field rescaling. Here, conventional and DRACAERIS detection of an NMR sample is simulated. The sample contains three types of nuclear spins at a 1-T bias field. Spin 1 is uncoupled with chemical shift $\nu = 25 \text{ Hz} \ (\delta = 0.59 \text{ ppm})$. Spins 2 and 3 have $\nu = 50, 150 \text{ Hz}$ $(\delta = 1.17, 3.52 \text{ ppm})$ and have J = 10 Hz scalar coupling. (a) When the free-precession time τ is used as the evolution time for the frequency axis, DRACAERIS yields spectral lines with the correct chemical shift but incorrect splitting due to J coupling. (b) When the total evolution time $\tau + \mu$ is used, DRACAERIS gives the correct splitting due to J coupling but incorrect chemical shifts. (c) A match is achieved for a conventional NMR spectrum at a magnetic field B_0 rescaled by $\tau/(\tau + \mu)$. Due to the longer experimental time for longitudinal sensing, the DRACAERIS spectral features have lower amplitudes and broader line widths than conventional NMR.

when using τ to measure free-precession time. This condition applies if the sample nuclei are described only by the chemical-shift Hamiltonian, since during strong driving (i.e., nutation-pulse Rabi frequency Ω much larger than chemical-shift differences $\delta \omega$) chemical-shift evolution is suppressed. In this case, the spin system at the end of the nutation pulse is nearly identical to its state at the beginning of the pulse.



FIG. 4. Simulated ethanol NMR spectra using conventional inductive and longitudinal DRACAERIS detection disagree due to the effects of scalar coupling between the sample spins: (a) field 1 T, $\tau/\mu = 10.0$; (b) field 1 T, $\tau/\mu = 1.0$; (c) field 10 T, $\tau/\mu = 10.0$; (d) field 10 T, $\tau/\mu = 1.0$. These effects become more pronounced for smaller ratios of τ/μ . The insets illustrate that the magnitude of disagreement is similar for 1-T and 10-T bias magnetic fields, although at 1 T extra splittings also appear due to higher-order effects. Uncoupled spins are unaffected. The 0-ppm line is a tetramethylsilane reference. Here, $\tau = 1$ ms, $\Omega = 20$ kHz, and nuclear spin $T_2 = 1$ s.

However, for coupled spin systems, the *J*-coupling terms of the Hamiltonian are not suppressed by strong nutation driving; i.e., the *J*-coupling Hamiltonian is effective for the entire time $\tau + \mu$. In this case, if one uses τ

as the measure of evolution time, our simulations indicate that the chemical-shift frequencies match the conventional spectrum but the splittings caused by J coupling are too large [Fig. 3(a)]. If the sampling time step is instead



FIG. 5. Simulations indicate that DRACAERIS measurements of longitudinal signals are much less sensitive to MW-pulse errors than the rewound acquisition version of AERIS [14]. (a) Here, we compare the performance of the multicycle XY8-N sequences used in DRACAERIS to the modified AERIS sequence over multiple cycles (Rewind2-2N). These two sequences have the same phaseaccumulation time. (b) Simulations of the normalized N-V ensemble longitudinal magnetization (proportional to the measured N-V fluorescence) show that XY8 sequences perform significantly better than Rewind2 in the presence of pulse errors, particularly for large numbers of repetitions. In the absence of pulse error, the sequences perform about equally (within 5%; the results shown here are normalized to the maximum measured signal to make bandwidth comparisons easier). The simulations are for a 50-pT-amplitude signal driven at 1 MHz and measured using an N-V ensemble with Gaussian-averaged $T_2^* = 100$ ns, a typical value [17].



FIG. 6. A simulated ethanol DRACAERIS NMR spectrum at 1-T bias field with $\tau/\mu = 1$ is reproduced from Fig. 4 and compared against an inductive NMR spectrum simulated for 0.5 T. There is good agreement among the locations of all spectral features. Nuclear spin decoherence causes the DRACAERIS spectral features to have lower amplitudes and somewhat broader line widths due to the additional experiment time required for the longitudinal-sensing protocol.

considered to be $\tau + \mu$, then our simulations show that the *J*-coupling splittings have the correct value but the chemical shifts are too small [Fig. 3(b)].

These systematic errors can be corrected with a straightforward modification to the analysis of the longitudinalsensing data. From simulations and average Hamiltonian theory, we find that for all nonzero μ , the AERIS or DRACAERIS spectrum matches the conventional NMR spectrum acquired at a bias magnetic field rescaled by $\tau/(\tau + \mu)$ (where τ is again the measure of freeprecession time for the time-domain signal). For example, if $\tau/\mu = 1$, a DRACAERIS spectrum acquired at 1 T would match the conventional spectrum acquired at 0.5 T [Fig. 3(c)].

This effect can be significant for the N-V–NMR spectra of real molecules. In Fig. 4, we show an example simulation for ethanol. Both low- and high-field spectra show better equivalence with conventional NMR spectra as τ/μ increases, particularly when $\tau/\mu \ge 10$. At 1 T and $\tau/\mu = 1$, the spectra begin to show extra splittings typical of those acquired at much lower field strengths, which result from higher-order effects of *J* coupling. In Fig. 6, we show that scaling the field of the reference spectrum by $\tau/(\tau + \mu)$ produces a match with DRACAERIS for all spectral features.

V. DISCUSSION

Longitudinal detection is an elegant way of shifting an NMR signal to a more convenient detection frequency and has been implemented in magnetic resonance in the past [19,20], including for some of the first N-V-NMR measurements [4]. However, the required driving pulses are subject to errors in amplitude and frequency in any realistic NMR apparatus, due to both intrinsic factors such as

off-resonant spins and extrinsic factors such as B_1 inhomogeneity. A common way to correct for pulse errors is to perform a second operation that reverses the leading order effect of an error; this can occur by either cycling the phases of pulses over the course of a measurement or by directly rewinding the error during the measurement step. The original description of AERIS [14] proposed an error-correction protocol, which, like DRACAERIS, performs a rewinding immediately before any free precession can occur under imperfect conditions. However, the protocol in Ref. [14] performs the forward and reverse pulses within a single modified N-V magnetometry sequence, while DRACAERIS instead splits this correction between two separate N-V magnetometry sequences. The DRACAERIS approach has two key benefits. First, it refrains from altering the N-V sensing protocol (such as XY8), which could corrupt its error-correction properties for N-V pulses. Second, it provides a differential measurement that reduces the effects of laser intensity noise [21].

To quantify the advantage of DRACAERIS in the presence of MW-pulse errors, we simulate and compare the performance of the modified AERIS protocol [14] and the forward-nutation portion of a single DRACAERIS measurement. We use ideal NMR signals but vary the lengths of MW pulses applied to the N-V ensemble to model common rotational pulse error. We take several steps to ensure that the simulation reflects meaningful measurements. To improve contrast, it is beneficial to use many cycles of XY8, designated XY8-N, where N is the number of cycles. To create a fair comparison, we also extend the modified AERIS sequence over multiple cycles, which we designate as the Rewind2-2N protocol. A single Rewind2 and a single XY4 cycle have the same sensing time, similar sensitivity, and similar levels of degradation due to pulse errors. However, as shown in Fig. 5, the XY8 protocol used in DRACAERIS maintains its sensitivity much better than Rewind2 as the number of cycles and sensing time increases. This superior performance is because extended Rewind2 sequences have vanishing error-correction benefits of alternating X and Y phases, making them analogous to Carr-Purcell-Meiboom-Gill (CPMG) sequences in the long-sequence-duration limit.

Regardless of the measurement scheme utilized, a rewind nutation is critical to the performance of any longitudinal protocol. Without rewind correction, a consistent pulse error applied to a resonant nuclear spin causes evolution along the *x*-*z* plane, which leads to an oscillating *X*-axis magnetization that appears as a frequency shift in the NMR spectrum (see the Appendix Fig. 7). For offresonant spins, this effect is somewhat suppressed once a spin precesses toward 180° around the Bloch sphere. By keeping the nuclear spins sufficiently far from resonance, it might be possible to minimize the effects of pulse errors without resorting to rewinding. However, this approach



FIG. 7. Time-domain simulations of a single nuclear spin during the AERIS sequence help elucidate the dynamics caused by pulse errors. (a) For an on-resonance spin with $\nu = 0$ Hz, perfect pulses lead to a simple exponentially decaying signal, the same type of signal that would be acquired with conventional NMR. (b) With 2% pulse errors and $\nu = 0$ Hz, AERIS produces a constant rotation in the *x*-*z* plane that causes an oscillating signal for *X*-axis magnetization. Spectroscopically, this makes the spin appear to be precessing in the transverse plane at a higher frequency. Since the *Y*-axis magnetization is not measured with this scheme, it cannot discriminate this effect from actual higher-frequency precession in the transverse plane. (c),(e) When applying perfect pulses to spins detuned by frequency ν , some magnetization oscillates between the longitudinal and transverse planes but the amount is small, as are the spectral effects: (c) $\nu = 100$ Hz; (e) $\nu = 500$ Hz. (d),(f) The effect of pulse errors is smaller for off-resonant spins because the signal is partially refocused as it precesses: (d) $\nu = 100$ Hz; (f) $\nu = 500$ Hz. For example, in (f) the spins spend less time along the *Z* axis than in (d). For these simulations, $\tau = \mu = 800$ ms, $\Omega = 5000$ Hz, and $T_2 = 1$ s.

wastes bandwidth and increases the effects of shifts caused by off-resonant excitation.

Even after correcting for pulse errors using DRA-CAERIS, it is important to account for spectral effects caused by the nutation pulses. NMR spectra typically have features produced by both chemical shifts (δ), which are field-strength dependent, as well as field-independent scalar couplings (J) between spins. The interplay between these effects causes the spectra to change significantly as a function of the magnetic field. At sufficiently high bias-field strengths, such that chemical-shift differences are significantly larger than scalar couplings ($\Delta \delta \gg J$), chemical shifts dominate the nuclear spin Hamiltonian and only the secular coupling term $2\pi J_{jk}I_{z,j}I_{z,k}$ is significant. In this case, NMR spectra typically consist of multiplets with splitting equal to J, separated by larger chemicalshift differences. A typical example is the ethanol NMR spectrum at a 1-T bias field, as shown in Fig. 4. At lower bias fields where $\Delta \delta \leq J$, higher-order effects caused by the nonsecular coupling terms $2\pi J_{ik}(I_{x,i}I_{x,k} + I_{v,i}I_{v,k})$ also become significant.

The effective chemical shift exhibited by the NMR sample also can be controlled by the measurement protocol. For example, the chemical shift can be suppressed during sufficiently strong pulses or by spin locking applied to nuclear spins, as well as by time reversal through one or more spin echoes. Morris et al. have demonstrated these effects in inductively detected NMR with a chemical-shift scaling protocol in which an XY dynamical decoupling sequence was performed between signal acquisitions [22]. The result is an NMR spectrum acquired at bias field B_0 matching a conventional NMR spectrum acquired at $\tau_2/(\tau_1 + \tau_2)B_0$, where τ_1 is the dynamical decoupling time and τ_2 is the spin free-precession time. This result matches our field scaling for DRACAERIS, consistent with the effects of dynamical decoupling and strong spin driving being analogous. Additionally, as in Ref. [22], altering the nutation duration may provide access to different spectral behavior in a high-field setting.

VI. CONCLUSIONS

We perform simulations of longitudinal detection techniques for N-V NMR and find that these techniques can create measurement errors due to pulse imperfections and off-resonance effects. We introduce a longitudinal

detection protocol called DRACAERIS (Double Rewound ACquisition Amplitude Encoded Radio Induced Signal) that simulations show can greatly suppress the errors from imperfect pulses. Additionally, we provide a model based on quantum simulations and confirmed by average Hamiltonian theory to help interpret longitudinally detected NMR spectra. Our results indicate that magnetic field rescaling can easily correct for predictable spectral distortions, due to the effects of *J* couplings between sample nuclear spins present in such spectra. This study provides a basis for experimental realization of the DRACAERIS protocol, which we intend to pursue in future work.

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D.D. initiated the investigation and then developed the DRACAERIS protocol together with E.H. and J.W.B. Authors D.D. and S.J.D., in consultation with J.W.B. and J.C., developed the theory and performed the simulations. R.L.W. supervised the work. All authors discussed the research and contributed to the final manuscript.

APPENDIX: AVERAGE HAMILTONIAN THEORY

Using average Hamiltonian theory (AHT), we confirm the magnetic field scaling behavior observed in the simulated spectra. Consider a system in the rotating frame with two spin-1/2 nuclei interacting via spin-spin coupling J_{12} . The resonance frequencies are ω_1 and ω_2 . The Hamiltonian is then

$$\hat{H}_0 = \omega_1 I_{z,1} + \omega_2 I_{z,2} + J_{12} \mathbf{I}_1 \cdot \mathbf{I}_2.$$
(A1)

The driving Hamiltonian for this system is written in the following way:

$$\hat{H}_1 = \Omega(I_{x,1} + I_{x,2}).$$
 (A2)

The Hamiltonians during free precession and nutation pulses are then

$$\hat{H}_{\text{int}} = \begin{cases} \hat{H}_{0}, & \text{if } t \in [0, \tau], \\ e^{-\Omega i (\mathbf{I}_{X})(t-\tau)} \hat{H}_{0} e^{\Omega i (\mathbf{I}_{X})(t-\tau)}, & \text{if } t \in [\tau, \tau + \mu]. \end{cases}$$
(A3)

The average Hamiltonian for a single acquisition cycle is given by the integral

$$\hat{H}_{\text{avg}} = \frac{1}{\tau + \mu} \left(\int_0^\tau H_{\text{int}} dt + \int_\tau^{\tau + \mu} H_{\text{int}} dt \right).$$
(A4)

This integral evaluates to

$$\begin{split} \hat{H}_{\text{avg}} &= \frac{\tau}{\tau + \mu} \left(J_{12} \mathbf{I}_1 \cdot \mathbf{I}_2 + \omega_1 I_{1,z} + \omega_2 I_{2,z} \right) \\ &+ \frac{\mu}{2(\tau + \mu)} \left[J_{12} (I_1^- \cdot I_2^+ + I_1^+ \cdot I_2^- + 2(I_{1,z} I_{2,z})) \right. \\ &- \frac{2i(\omega_1 (I_1^- - I_1^+) + \omega_2 (I_2^- - I_2^+)) \sin^2(\mu \Omega/2)}{\mu \Omega} \\ &+ \frac{2(\omega_1 I_{1,z} + \omega_2 I_{2,z}) \sin(\mu \Omega)}{\mu \Omega} \right]. \end{split}$$

The terms divided by $\mu\Omega$ become zero if the nutation pulses are perfect so that $\mu\Omega = 2\pi N$, i.e., at least one full nutation is performed. Even if the pulses are imperfect, these terms are typically small, as they are suppressed by the factor $\Omega(\tau + \mu)$. The remaining terms are

$$\hat{H}_{\text{avg}} = \frac{\tau}{\tau + \mu} \left(J_{12} \mathbf{I}_1 \cdot \mathbf{I}_2 + \omega_1 I_{1,z} + \omega_2 I_{2,z} \right)$$
(A5)

$$+\frac{J_{12}\mu}{2(\tau+\mu)}(I_1^- \cdot I_2^+ + I_1^- + \cdot I_2^- + 2(I_{1,z}I_{2,z})).$$
(A6)

Using $I_i^{\pm} = I_{i,x} \pm I_{i,y}$,

$$\hat{H}_{avg} = \frac{\tau}{\tau + \mu} \left(J_{12} \mathbf{I}_1 \cdot \mathbf{I}_2 + \omega_1 I_{1,z} + \omega_2 I_{2,z} \right)$$
(A7)

$$+\frac{\mu}{\tau+\mu}(J_{12}\mathbf{I}_1\cdot\mathbf{I}_2). \tag{A8}$$

Adding terms together gives

$$\hat{H}_{avg} = \frac{\tau}{\tau + \mu} (\omega_1 I_{1,z} + \omega_2 I_{2,z}) + J_{12} \mathbf{I}_1 \cdot \mathbf{I}_2.$$
(A9)

This first-order average Hamiltonian is for a spin system the resonance frequencies of which have been scaled by a factor of $\tau/(\tau + \mu)$, a result that can also be obtained by scaling B_0 by the same factor. In the strong limit that $\mu \rightarrow 0$, this scaling factor disappears and the AHT first-order result approaches the conventional NMR Hamiltonian.

- J. Zhang, L. Xu, G. Bian, P. Fan, M. Li, W. Liu, and H. Yuan, Diamond nitrogen-vacancy center magnetometry: Advances and challenges, arXiv:2010.10231.
- [2] R. Schirhagl, K. Chang, M. Loretz, and C. L. Degen, Nitrogen-vacancy centers in diamond: Nanoscale sensors for physics and biology, Annu. Rev. Phys. Chem. 65, 83 (2014).
- [3] E. V. Levine, M. J. Turner, P. Kehayias, C. A. Hart, N. Langellier, R. Trubko, D. R. Glenn, R. R. Fu, and R. L. Walsworth, Principles and techniques of the

quantum diamond microscope, Nanophotonics 8, 1945 (2019).

- [4] H. J. Mamin, M. Kim, M. H. Sherwood, C. T. Rettner, K. Ohno, D. D. Awschalom, and D. Rugar, Nanoscale nuclear magnetic resonance with a nitrogen-vacancy spin sensor, Science 339, 557 (2013).
- [5] T. Staudacher, F. Shi, S. Pezzagna, J. Meijer, J. Du, C. A. Meriles, F. Reinhard, and J. Wrachtrup, Nuclear magnetic resonance spectroscopy on a (5-nanometer)³ sample volume, Science 339, 561 (2013).
- [6] S. J. DeVience, L. M. Pham, I. Lovchinsky, A. O. Sushkov, N. Bar-Gill, C. Belthangady, F. Casola, M. Corbett, H. Zhang, M. Lukin, H. Park, A. Yacoby, and R. L. Walsworth, Nanoscale NMR spectroscopy and imaging of multiple nuclear species, Nat. Nanotechnol. 10, 129 (2015).
- [7] M. Loretz, S. Pezzagna, J. Meijer, and C. L. Degen, Nanoscale nuclear magnetic resonance with a 1.9-nm-deep nitrogen-vacancy sensor, Appl. Phys. Lett. 104, 033102 (2014).
- [8] D. Rugar, H. J. Mamin, M. H. Sherwood, M. Kim, C. T. Rettner, K. Ohno, and D. D. Awschalom, Proton magnetic resonance imaging using a nitrogen-vacancy spin sensor, Nat. Nanotechnol. 10, 120 (2015).
- [9] D. R. Glenn, D. B. Bucher, J. Lee, M. D. Lukin, H. Park, and R. L. Walsworth, High-resolution magnetic resonance spectroscopy using a solid-state spin sensor, Nature 555, 351 (2018).
- [10] N. Aslam, M. Pfender, P. Neumann, R. Reuter, A. Zappe, F. Fávaro de Oliveira, A. Denisenko, H. Sumiya, S. Onoda, J. Isoya *et al.*, Nanoscale nuclear magnetic resonance with chemical resolution, Science **357**, 67 (2017).
- [11] S. Schmitt, T. Gefen, F. M. Stürner, T. Unden, G. Wolff, C. Müller, J. Scheuer, B. Naydenov, M. Markham, S. Pezzagna, J. Meijer, I. Schwarz, M. Plenio, A. Retzker, L. P. McGuinness, and F. Jelezko, Submillihertz magnetic spectroscopy performed with a nanoscale quantum sensor, Science 356, 832 (2017).
- [12] P. Kehayias, A. Jarmola, N. Mosavian, I. Fescenko, F. M. Benito, A. Laraoui, J. Smits, L. Bougas, D. Budker, A. Neumann, S. R. J. Brueck, and V. M. Acosta, Solution nuclear magnetic resonance spectroscopy on a nanostructured diamond chip, Nat. Commun. 8, 188 (2017).
- [13] D. B. Bucher, D. R. Glenn, H. Park, M. D. Lukin, and R. L. Walsworth, Hyperpolarization-enhanced NMR spectroscopy with femtomole sensitivity using quantum defects in diamond, Phys. Rev. X 10, 021053 (2020).
- [14] C. Munuera-Javaloy, A. Tobalina, and J. Casanova, Highresolution NMR spectroscopy at large fields with nitrogen vacancy centers, Phys. Rev. Lett. 130, 133603 (2023).
- [15] C. P. Slichter, *Principles of Magnetic Resonance* (Springer-Verlag, Berlin, 1990), p. 242.
- [16] I. Solomon, Relaxation processes in a system of two spins, Phys. Rev. 99, 559 (1955).
- [17] J. Schloss, Doctoral dissertation, School of Science, Massachusetts Institute of Technology, 2019.
- [18] N. Arunkumar, K. S. Olsson, J. T. Oon, C. A. Hart, D. B. Bucher, D. R. Glenn, M. D. Lukin, H. Park, D. Ham, and R. L. Walsworth, Quantum logic enhanced sensing in

solid-state spin ensembles, Phys. Rev. Lett. 131, 100801 (2023).

- [19] A. Schweiger and R. Ernst, Pulsed ESR with longitudinal detection. A novel recording technique, J. Magn. Reson. (1969) 77, 512 (1988).
- [20] G. Whitfield and A. Redfield, Paramagnetic resonance detection along the polarizing field direction, Phys. Rev. 106, 918 (1957).
- [21] N. Bar-Gill, L. M. Pham, A. Jarmola, D. Budker, and R. L. Walsworth, Solid-state electronic spin coherence time approaching one second, Nat. Commun. 4, 1743 (2013).
- [22] G. A. Morris, N. P. Jerome, and L.-Y. Lian, Realtime chemical-shift scaling in high-resolution NMR spectroscopy, Angew. Chem. (International Ed. in English) 42, 823 (2003).